COLLEGE of AMERICAN PATHOLOGISTS

Initial Diagnostic Workup of Acute Leukemia

Guideline from the College of American Pathologists (CAP) and the American Society of Hematology (ASH)

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Background

The diagnosis and prognostic determination of acute leukemia currently incorporate

- Clinical information
- Morphology
- Immunophenotyping
- Cytogenetics
- Molecular genetics



Background continued

Why have guidelines for the work-up of acute leukemia?

Clinical practice guidelines have the potential to:

- Standardize care across all settings
- Reduce duplication of testing
- Support the use of specific tests
- Improve patient care and outcomes



Introduction

 The College of American Pathologists (CAP) and the American Society for Hematology (ASH) formed an expert panel to review the relevant literature and to establish a guideline for appropriate laboratory testing as well as for the clinical information necessary for the initial diagnosis of acute leukemia, including AML, ALL, and acute leukemias of ambiguous lineage.



Introduction continued

- The panel closely followed the Institute of Medicine <u>Clinical Practice Guidelines We Can Trust</u> standards for guideline development
 - 1. Establish transparency
 - 2. Manage conflicts of interest
 - 3. Establish a multi-disciplinary panel
 - 4. Perform systematic review
 - 5. Rate strength of recommendations
 - 6. Articulate the recommendations
 - 7. Include external review

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Guideline expert panel members

- Daniel A. Arber, MD, CAP co-chair
- James W. Vardiman, MD, ASH co-chair
- Melissa Cessna, MD
- Joan Etzell, MD
- Sa Wang, MD
- Karl Theil, MD, PhD
- Michael J. Borowitz, MD, PhD
- Robert Hasserjian, MD
- Kathryn Foucar, MD



Guideline staff

- Nicole E. Thomas, MPH, CT(ASCP)^{cm}, CAP Senior Manager, Center Guideline Development
- Robert Plovnick, MD, ASH Director of Quality Improvement
- Robert Kunkle, ASH Senior Manager, Practice Guidelines
- Kendall Alexander, MPH, ASH Clinical Quality Projects
 Specialist
- R. Bryan Rumble, MSc, Methodology Consultant



• Tony Smith, MLS, ECMS (AIIM), Medical Librarian

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Systematic evidence review

- Identify key questions
- Literature search
- Data extraction
- Develop proposed recommendations
- Open comment period
- Considered judgment process
 - Consider risks and benefits, cost, regulatory requirements, preferences, etc.



Key questions

- 1. What clinical and laboratory information should be available during the initial diagnostic evaluation of a patient with acute leukemia?
- 2. What specimens and sample types should be evaluated during the initial work-up of a patient with acute leukemia?
- 3. At the time of diagnosis, what tests are required for all patients for the initial evaluation of an acute leukemia?



Key questions continued

- 4. Which tests should be performed only on a subset of patients, including in response to results of initial tests and morphology?
- 5. Where should laboratory testing be performed?
- 6. How should test results and the diagnosis be correlated and reported?



Systematic evidence review results

- Initial literature search conducted for studies from 1/2005– 9/2011; 2 literature refreshes to include studies from 9/2011 – 8/2015
- 4,901 articles identified for abstract review
 - 1,182 articles submitted for full text review
 - 119 articles underwent data extraction and quality assessment analysis
 - Additional references from expert panel
- Panel had face-to-face meetings and conference



calls to develop 29 draft statements

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Definition of strength of recommendations

Grades for Strength of Recommendations		
Designation	Recommendation	Rationale
Strong Recommendation	Recommend for or against a particular	Supported by convincing (high) or adequate
	practice (Can include "must" or	(intermediate) quality of evidence and clear benefit
	"should")	that outweighs any harms
Recommendation	Recommend for or against a particular	Some limitations in quality of evidence (adequate
	practice (Can include "should" or	[intermediate] or inadequate [low]), balance of
	"may")	benefits and harms, values, or costs but panel
		concludes that there is sufficient evidence and/or
		benefit to inform a recommendation
Expert Consensus Opinion	Recommend for or against a particular	Serious limitations in quality of evidence (inadequate
	practice (Can include "should" or	[low] or insufficient), balance of benefits and harms,
	"may")	values or costs, but panel consensus is that a
		statement is necessary
No Recommendation	No recommendation for or against a	Insufficient evidence or agreement of the balance of
	practice	benefits and harms, values, or costs to provide a
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Systematic evidence review results continued

- Results from the open comment period
 - August 10-31, 2015: 29 draft statements were presented
 - 162-200 respondents for each draft statement
 - 789 written comments
 - 26 draft statements achieved more than 90% agreement, two achieved 80 - 90% agreement, and one received 70-80% agreement
- Final guideline includes 27 guideline statements (recommendations)



Strong Recommendation - The treating clinician should provide relevant clinical data or ensure that this is readily accessible by the pathologist.

Note: These data include, but is not limited to the patient's age, gender, ethnicity, history of any hematologic disorder or known predisposing conditions or syndromes, any prior malignancy, exposure to cytotoxic therapy, immunotherapy, radiotherapy, or other possibly toxic substances, and any additional clinical findings of diagnostic or prognostic importance. The treating clinician should also include any history of possibly confounding factors, such as recent growth factor therapy, transfusions or other medications that might obscure or mimic the features of acute leukemia. The treating clinician should also obtain and provide information regarding any family history of any hematologic disorder or other malignancies.



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Guideline Statement 1 rationale

- Clinical information is essential in AL diagnosis
- Detection of germline predisposition disorders is critical for treatment and prognosis
- Pathologist must know about confounding factors such as G-CSF therapy or other prior therapy



Recommendation - The treating clinician should provide relevant physical examination and imaging findings or ensure that these are readily accessible by the pathologist.

Note: This includes, but is not limited to, neurologic exam findings and the presence of tumor masses (e.g., mediastinal), other tissue lesions (e.g., cutaneous), and/or organomegaly.



Guideline Statement 2 rationale

- Extramedullary sites of disease could be biopsied, if necessary
- Diagnostic evaluation could be performed on an extramedullary specimen
- Certain type of tumor masses (e.g. mediastinal mass) could provide clues as to a specific AL subtype



Strong Recommendation - The pathologist should review recent or concurrent complete blood counts (CBCs) and leukocyte differentials and evaluate a peripheral blood smear.



Strong Recommendation - The treating clinician or pathologist should obtain a fresh bone marrow aspirate for all patients suspected of acute leukemia, a portion of which should be used to make bone marrow aspirate smears for morphologic evaluation. If performed, the pathologist should evaluate an adequate bone marrow trephine core biopsy, bone marrow trephine touch preparations, and/or marrow clots, in conjunction with the bone marrow aspirates.

Note: If bone marrow aspirate material is inadequate or if there is compelling clinical reason to avoid bone marrow examination, peripheral blood may be used for diagnosis and ancillary studies if sufficient numbers of blasts are present. If a bone marrow aspirate is unobtainable, touch imprint preparations of a core biopsy should be prepared and evaluated, and an additional core biopsy may be submitted unfixed in tissue culture medium for disaggregation for flow and genetic studies. Optimally, the same physician should interpret the BM aspirate smears and the core biopsy specimens, or the interpretations of these specimens should be correlated if performed by different physicians.



Guideline Statement 4 rationale

 Specialized testing requires viable cells so adequate specimens for both routine assessment and specialized testing are essential



Strong Recommendation - In addition to morphologic assessment (blood and bone marrow), the pathologist or treating clinician should obtain sufficient samples and perform conventional cytogenetic analysis (i.e., karyotype), appropriate molecular genetic and/or fluorescent in-situ hybridization (FISH) testing, and flow cytometric immunophenotyping (FCI). The flow cytometry panel should be sufficient to distinguish acute myeloid leukemia (including acute promyelocytic leukemia), Tcell acute lymphoblastic leukemia (T-ALL) (including early T-cell precursor leukemias), B-cell precursor ALL (B-ALL), and acute leukemia of ambiguous lineage on all patients diagnosed with acute leukemia. Molecular genetic and/or FISH testing does not, however, replace conventional cytogenetic analysis.

Note: If sufficient bone marrow aspirate or peripheral blood material is not available for FCI, immunohistochemical studies may be used as an alternative method for performing limited immunophenotyping. In addition, a second bone marrow core biopsy can be obtained and submitted unfixed in tissue culture media for disaggregation for genetic studies and flow cytometry.



Expert Consensus Opinion - For patients with suspected or confirmed acute leukemia, the pathologist may request and evaluate cytochemical studies to assist in the diagnosis and classification of acute myeloid leukemia (AML).



Guideline Statement 6 rationale

- Cytoechemical MPO staining is a rapid and sensitive method to confirm myeloid blast lineage
- Cytochemical MPO is especially valuable in the rapid diagnosis of acute promyelocytic leukemia



Recommendation - The treating clinician or pathologist may use cryopreserved cells or nucleic acid, formalin fixed, non-decalcified paraffinembedded (FFPE) tissue, or unstained marrow aspirate or peripheral blood smears obtained and prepared from peripheral blood, bone marrow aspirate or other involved tissues for molecular or genetic studies in which the use of such material has been validated. Such specimens must be properly identified and stored under appropriate conditions in a laboratory that is in compliance with regulatory and/or accreditation requirements.



Guideline Statement 7 rationale

 Both banked, viable cell specimens and formalinfixed, paraffin-embedded specimens are useful for various types of testing if fresh tissue is not available



Strong Recommendation - For patients with acute lymphoblastic leukemia (ALL) receiving intrathecal therapy, the treating clinician should obtain a cerebrospinal fluid (CSF) sample. The treating clinician or pathologist should ensure that a cell count is performed and that examination/enumeration of blasts on a cytocentrifuge preparation is performed and is reviewed by the pathologist.



Guideline Statement 8 rationale

- Monitoring of CSF is a standard practice in patients with AL, especially acute lymphoblastic leukemias
- The pathologist must assess these CSF specimens for recognition and enumeration of blasts



Expert Consensus Opinion - For patients with acute leukemia other than those with ALL receiving intrathecal therapy, the treating clinician may, under certain circumstances, obtain a cerebrospinal fluid (CSF) sample when there is no clinical contraindication. The treating clinician or pathologist should ensure that a cell count is performed and that examination/enumeration of blasts on a cytocentrifuge preparation is performed and is reviewed by the pathologist.

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Recommendation - For patients with suspected or confirmed acute leukemia, the pathologist may use flow cytometry in the evaluation of CSF.



Strong Recommendation - For patients who present with extramedullary disease without bone marrow or blood involvement, the pathologist should evaluate a tissue biopsy and process it for morphologic, immunophenotypic, cytogenetic, and molecular genetic studies, as recommended for the bone marrow.

Note: Additional biopsies may be indicated to obtain fresh material for ancillary testing.



Strong Recommendation - For patients with suspected or confirmed acute leukemia, the pathologist or treating clinician should ensure that flow cytometry analysis or molecular characterization is comprehensive enough to allow subsequent detection of minimal residual disease (MRD).



Guideline Statement 12 rationale

- Flow cytometric immunophenotyping is required for lineage confirmation in all cases of acute leukemia.
- Since persistent leukemia after therapy is a key prognostic factor for some types of AL, minimal residual disease monitoring by either flow cytometry or genetic testing is often required.



Strong Recommendation - For pediatric patients with suspected or confirmed B-ALL, the pathologist or treating clinician should ensure that testing for t(12;21)(p13.2;q22.1); *ETV6-RUNX1*, t(9;22)(q34.1;q11.2); *BCR-ABL1*, *KMT2A(MLL)* translocation, iAMP 21, and trisomy 4 and 10 is performed.



Strong Recommendation - For testing for t(9;22)(q34.1;q11.2) and *BCR-ABL1*; Recommendation for testing for *KMT2A* (*MLL*) translocations

For adult patients with suspected or confirmed B-ALL, the pathologist or treating clinician should ensure that testing for t(9;22)(q34.1;q11.2); *BCR-ABL1* is performed. In addition, testing for *KMT2A* (*MLL*) translocations may be performed.



Recommendation - For patients with suspected or confirmed ALL, the pathologist or treating clinician may order appropriate mutational analysis for selected genes that influence diagnosis, prognosis, and/or therapeutic management that includes, but is not limited to: PAX5, JAK1, JAK2, and/or IKZF1 for B-ALL and NOTCH1 and/or FBXW7 for T-ALL. Testing for overexpression of CRLF2 may also be performed for **B-ALL**.



Guideline Statements 13-17 rationale

 Genetic testing provides both diagnostic and prognostic information in many types of AL including precursor B-ALL and all AML subtypes.



Strong recommendation - for testing for *FLT3*-ITD; Recommendation for testing for other mutational analysis –

For pediatric and adult patients with suspected or confirmed acute myeloid leukemia (AML) of any type, the pathologist or treating clinician should ensure that testing for *FLT3*-ITD is performed. The pathologist or treating clinician may order mutational analysis that includes but is not limited to: *IDH1, IDH2, TET2, WT1, DNMT3A*, and/or *TP53* for prognostic and/or therapeutic purposes.



Strong Recommendation - for testing for *KIT* mutation in adult patients with CBF AML; Expert Consensus Opinion for testing for *KIT* mutation in pediatric patients with CBF AML –

For adult patients with confirmed core binding factor (CBF) AML (AML with t(8;21)(q22;q22.1); *RUNX1-RUNX1T1* or inv(16)(p13.1q22) /t(16;16)(p13.1;q22); *CBFB-MYH11*), the pathologist or treating clinician should ensure that appropriate mutational analysis for *KIT* is performed. For pediatric patients with confirmed core binding factor AML (AML with t(8;21)(q22;q22.1); *RUNX1-RUNX1T1* or inv(16)(p13.1q22) /t(16;16)(p13.1;q22); *CBFB-MYH11*), the pathologist or treating clinician may ensure that appropriate mutational analysis for *KIT* is performed.



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Strong Recommendation - For patients with suspected acute promyelocytic leukemia (APL), the pathologist or treating physician should also ensure that rapid detection of *PML-RARA* is performed. The treating physician should also order appropriate coagulation studies to evaluate for disseminated intravascular coagulation (DIC).



Guideline Statement 18 rationale

- Because of imminent risk of life-threatening hemorrhage, the rapid diagnosis of acute promyelocytic leukemia is essential.
- Implementation of ATRA therapy as early as possible is critical.



Strong Recommendation - For patients other than those with confirmed core binding factor AML, APL, or AML with myelodysplasia-related cytogenetic abnormalities, the pathologist or treating clinician should also ensure that mutational analysis for *NPM1, CEBPA*, and *RUNX1* is also performed.



No Recommendation - For patients with confirmed acute leukemia, no recommendation is made for or against the use of global/gene specific methylation, micro RNA (miRNA) expression, or gene expression analysis for diagnosis or prognosis.



Strong Recommendation - For patients with confirmed mixed phenotype acute leukemia (MPAL), the pathologist or treating clinician should ensure that testing for t(9;22)(q34.1;q11.2); *BCR-ABL1*, and *KMT2A* (*MLL*) translocations is performed.



Strong Recommendation - All laboratory testing performed for the initial work-up and diagnosis of a patient with acute leukemia must be performed in a laboratory that is in compliance with regulatory and/or accreditation requirements.



Strong Recommendation - If after examination of a peripheral blood smear, it is determined that the patient will require immediate referral to another institution with expertise in the management of acute leukemia for treatment, the initial institution should, whenever possible, defer invasive procedures including bone marrow aspiration and biopsies to the treatment center to avoid duplicate procedures, associated patient discomfort, and additional costs.



Strong Recommendation - If a patient is referred to another institution for treatment, the primary institution should provide the treatment center with all laboratory results, pathology slides, flow cytometry data, cytogenetic information, and a list of pending tests at the time of the referral. Pending test results should be forwarded as they become available.



Strong Recommendation - If a patient is referred to another institution for treatment, the primary institution should provide the treatment center with all laboratory results, pathology slides, flow cytometry data, cytogenetic information, and a list of pending tests at the time of the referral. Pending test results should be forwarded as they become available.



Strong Recommendation - The pathologist and treating clinician should coordinate and ensure that all tests performed for classification, management, predicting prognosis and disease monitoring are entered into the patient's medical records.

Note: This information should include the sample source, adequacy, and collection information as applicable.



Strong Recommendation – Treating physicians and pathologists should use the current World Health Organization (WHO) terminology for the final diagnosis and classification of acute leukemia.



Conclusion

Clinical Practice Guidelines: Here to Stay

- Provide significant positive impact on patient care
- Increases standardization across practice settings
- Continuous updating is key



Link to guideline

<u>http://www.archivesofpathology.org/doi/pdf/10.5858</u>
 <u>/arpa.2016-0504-CP</u>





