



COLLEGE of AMERICAN
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Molecular Diagnostic Testing in Myelodysplastic Syndrome

CAP PHC Webinar

Adam Seegmiller, MD, PhD
Michael Savona, MD

4/24/2018

Vanderbilt University
Medical Center

Webinar Host

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Adam Seegmiller, MD, PhD, FCAP

- **Associate Professor and Vice Chair, Department of Pathology, Microbiology, and Immunology at Vanderbilt University School of Medicine**
- **Executive Medical Director, Clinical Pathology at Vanderbilt University Medical Center**
- **Leads the divisions of Laboratory Medicine and Hematopathology, and actively practices clinical hematopathology**



Michael Savona, MD

- **Director, Hematology Research and Director, Hematology Early Therapy Program Vanderbilt-Ingram Cancer Center**
- **Associate Professor of Medicine, Vanderbilt University School of Medicine**
- **Translational scientist and medical oncologist specializing in myeloid malignancies, and a leader in development of novel therapies for these diseases patients**



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Learning Objectives

1. To understand the current approach to the diagnosis of MDS.
2. To describe the potential and the limitations of applying molecular diagnostic testing in MDS.
3. To discuss the impact of molecular diagnostic testing on the prognosis and treatment of patients with MDS.

Part 1: Genetic Testing in the Diagnosis and Evaluation of Myelodysplastic Syndrome

Adam Seegmiller, MD, PhD

Case Presentation

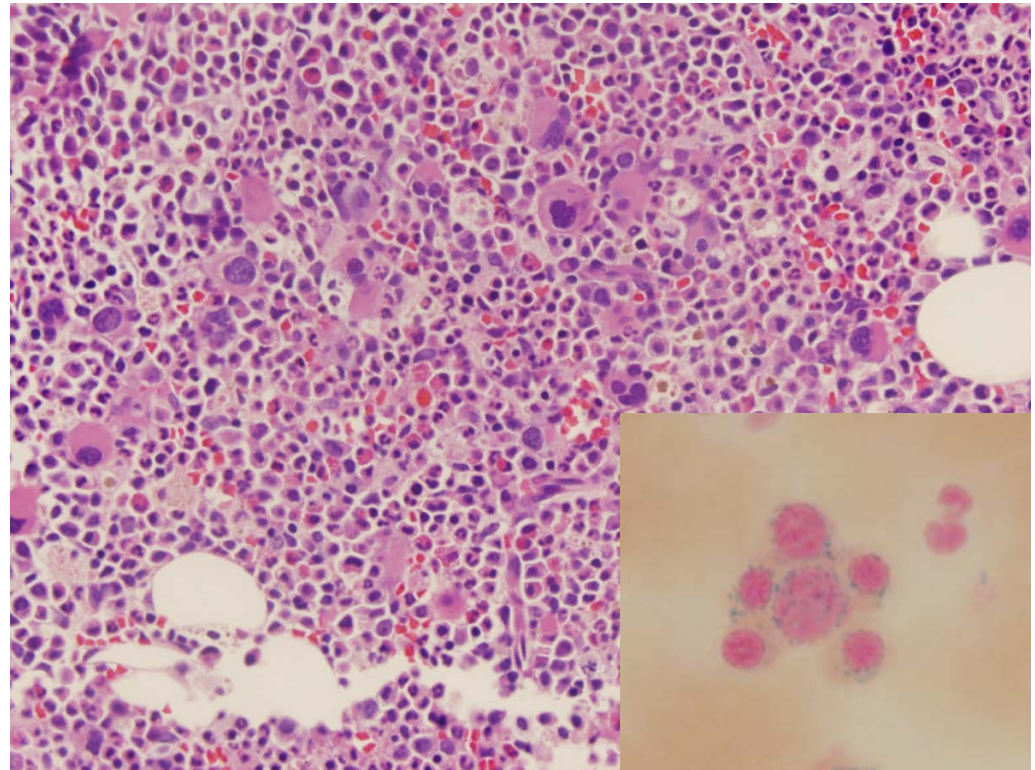
- **A 65 year-old female with history of follicular thyroid carcinoma treated with radioiodine.**
- **Presents to the emergency department complaining of shortness of breath, weakness, and fatigue.**
- **Physical examination was significant for tachycardia and pallor.**

Case Presentation

- **Complete blood count**
 - Profound macrocytic anemia (Hbg 4.0 g/dL, MCV 113 fL)
 - Mild leukocytosis (WBC $17.7 \times 10^3/\mu\text{L}$)
 - Mild thrombocytosis ($470 \times 10^3/\mu\text{L}$)

Bone Marrow Biopsy

- **Hypercellular (90%)**
- **Increased abnormal megakaryocytes**
- **Dysplastic erythroid precursors**
- **Ring sideroblasts**



Myelodysplastic Syndrome (MDS)

- **Clonal neoplasm of myeloid precursors**
- **Characterized by:**
 - Ineffective hematopoiesis → usually hypercellular bone marrow with peripheral cytopenias
 - Morphologic dysplasia in one or more myeloid cell line (granulocytic, erythroid, or megakaryocytic)
 - Increased risk for development of acute myeloid leukemia

Hasserjian RP, et al. *WHO Classification*. 2017:98
Arber DA, et al. *Blood*. 2016;127(20):2391.
Steensma DP. *Mayo Clin Proc*. 2015;90(7):969

Myelodysplastic Syndrome (MDS)

- Incidence is 10,000-40,000 per year in the US
- Mostly a disease of older individuals, due to accumulation of mutations over a lifetime
- Toxic exposures increase risk – 10-15% of cases are due to prior chemotherapy
- Some inherited hematopoietic disorders predispose to MDS

Hasserjian RP, et al. *WHO Classification*. 2017:98
Arber DA, et al. *Blood*. 2016;127(20):2391.
Steensma DP. *Mayo Clin Proc*. 2015;90(7):969

Diagnosis of MDS

- **Diagnosis of MDS requires the following:**
 1. **At least one peripheral blood cytopenia*:**
 - Anemia (Hbg <10 g/dL)
 - Neutropenia (absolute count <1.8x10³/μL)
 - Thrombocytopenia (platelets <100x10³/μL)
 2. **Morphologic dysplasia: >10% dysplastic blood or bone marrow cells in any myeloid lineage**
 3. **<20% blasts in blood and bone marrow**

Hasserjian RP, et al. *WHO Classification*. 2017:98
Arber DA, et al. *Blood*. 2016;127(20):2391.
Steensma DP. *Mayo Clin Proc*. 2015;90(7):969

Differential Diagnosis

- **Cytopenias with dysplasia can be seen in many non-clonal reactive conditions:**
 - Infections
 - Nutritional deficiencies
 - Drug effects
 - Immune or inflammatory disorders
 - Congenital syndromes
- **These should be ruled out before a definitive diagnosis is made.**

Classification of MDS

Classification is based on:

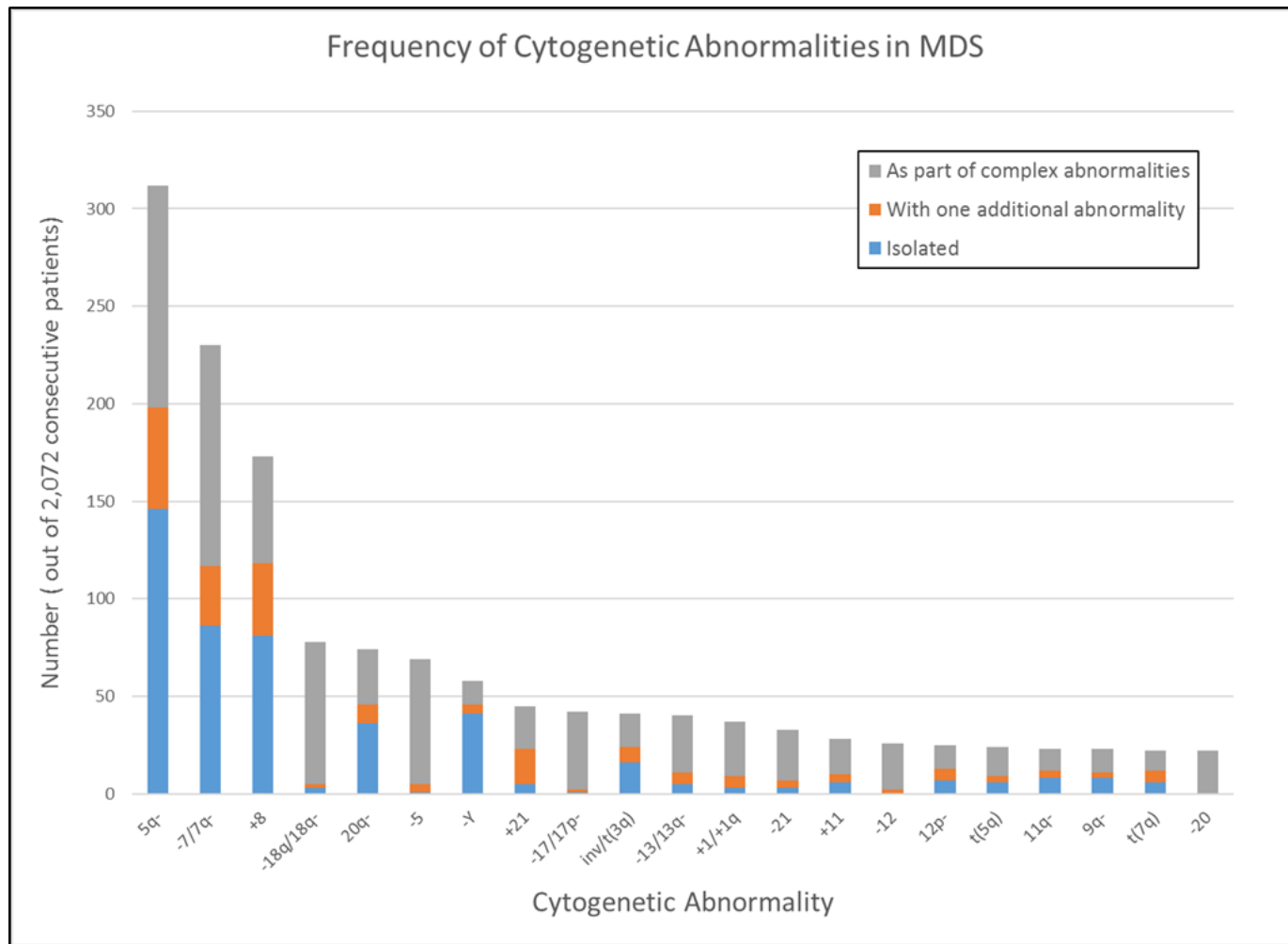
1. Number of hematopoietic lineages affected.
2. Percentage of marrow or blood blasts
3. Presence/absence of ring sideroblasts
4. Cytogenetic profile

Classification of MDS

Name	Dysplastic lineages	Cytopenias*	Ring sideroblasts as % of marrow erythroid elements	BM and PB blasts	Cytogenetics by conventional karyotype analysis
MDS with single lineage dysplasia (MDS-SLD)	1	1 or 2	<15%/<5%†	BM <5%, PB <1%, no Auer rods	Any, unless fulfills all criteria for MDS with isolated del(5q)
MDS with multilineage dysplasia (MDS-MLD)	2 or 3	1-3	<15%/<5%†	BM <5%, PB <1%, no Auer rods	Any, unless fulfills all criteria for MDS with isolated del(5q)
MDS with ring sideroblasts (MDS-RS)					
MDS-RS with single lineage dysplasia (MDS-RS-SLD)	1	1 or 2	≥15%/≥5%†	BM <5%, PB <1%, no Auer rods	Any, unless fulfills all criteria for MDS with isolated del(5q)
MDS-RS with multilineage dysplasia (MDS-RS-MLD)	2 or 3	1-3	≥15%/≥5%†	BM <5%, PB <1%, no Auer rods	Any, unless fulfills all criteria for MDS with isolated del(5q)
MDS with isolated del(5q)	1-3	1-2	None or any	BM <5%, PB <1%, no Auer rods	del(5q) alone or with 1 additional abnormality except -7 or del(7q)
MDS with excess blasts (MDS-EB)					
MDS-EB-1	0-3	1-3	None or any	BM 5%-9% or PB 2%-4%, no Auer rods	Any
MDS-EB-2	0-3	1-3	None or any	BM 10%-19% or PB 5%-19% or Auer rods	Any
MDS, unclassifiable (MDS-U)					
with 1% blood blasts	1-3	1-3	None or any	BM <5%, PB = 1%,‡ no Auer rods	Any
with single lineage dysplasia and pancytopenia	1	3	None or any	BM <5%, PB <1%, no Auer rods	Any
based on defining cytogenetic abnormality	0	1-3	<15%§	BM <5%, PB <1%, no Auer rods	MDS-defining abnormality
Refractory cytopenia of childhood	1-3	1-3	None	BM <5%, PB <2%	Any

Hasserjian RP, et al. *WHO Classification*. 2017:98
 Arber DA, et al. *Blood*. 2016;127(20):2391.

Cytogenetics in MDS



Haase D. *Ann Hematol.* 2008;87(7):515-25.

Cytogenetics and Diagnosis

- **MDS-defining abnormalities allow for a diagnosis of MDS even in the absence of definitive morphologic dysplasia:**
 - Loss of chromosomes 7 or 13
 - del(5q), del(7q), del(9q), del(11q), del(12p), del(13q)
 - t(1;3), t(2;11), inv(3)/t(3;3), t(3;21), t(6;9), t(11;16), or translocations involving 12p or 17p
 - i(17q), idic(X)(q13)
- **Presence of –Y, +8, or del(20q) as a sole abnormality is insufficient for diagnosis.**

Hasserjian RP, et al. *WHO Classification*. 2017:98
Arber DA, et al. *Blood*. 2016;127(20):2391.

Case Study Patient

Abnormal female karyotype:

46,XX,del(5)(q22q35)[11]/46,XX[9]



Image from atlasgeneticsoncology.org

MDS with isolated del(5q)

- Only cytogenetic abnormality that defines a distinct diagnostic entity.
- Most common in older women.
- Typically presents with anemia, often severe and macrocytic; thrombocytosis is common.
- Marrow morphology usually shows erythroid hypoplasia and abnormal megakaryocytes with hypolobated nuclei.
- Generally favorable outcomes with low risk of transformation compared with other MDS subtypes.

Hasserjian RP, et al. *WHO Classification*. 2017:98
Arber DA, et al. *Blood*. 2016;127(20):2391.

Other Cytogenetic/Morphologic Correlates

- **inv(3)/t(3;3) – abnormal megakaryocytes and thrombocytosis**
- **del(17p) – pseudo-Pelger-Huët anomaly, small vacuolated neutrophils**
- **del(20q) – dysmegakaryopoiesis and thrombocytopenia**

Rogers HJ, et al. *Hematologica*. 2014;99:821
Lai JL, et al. *Leukemia*. 1995;9:370.
Braun T, et al. *Leuk Res*. 2011;35:863

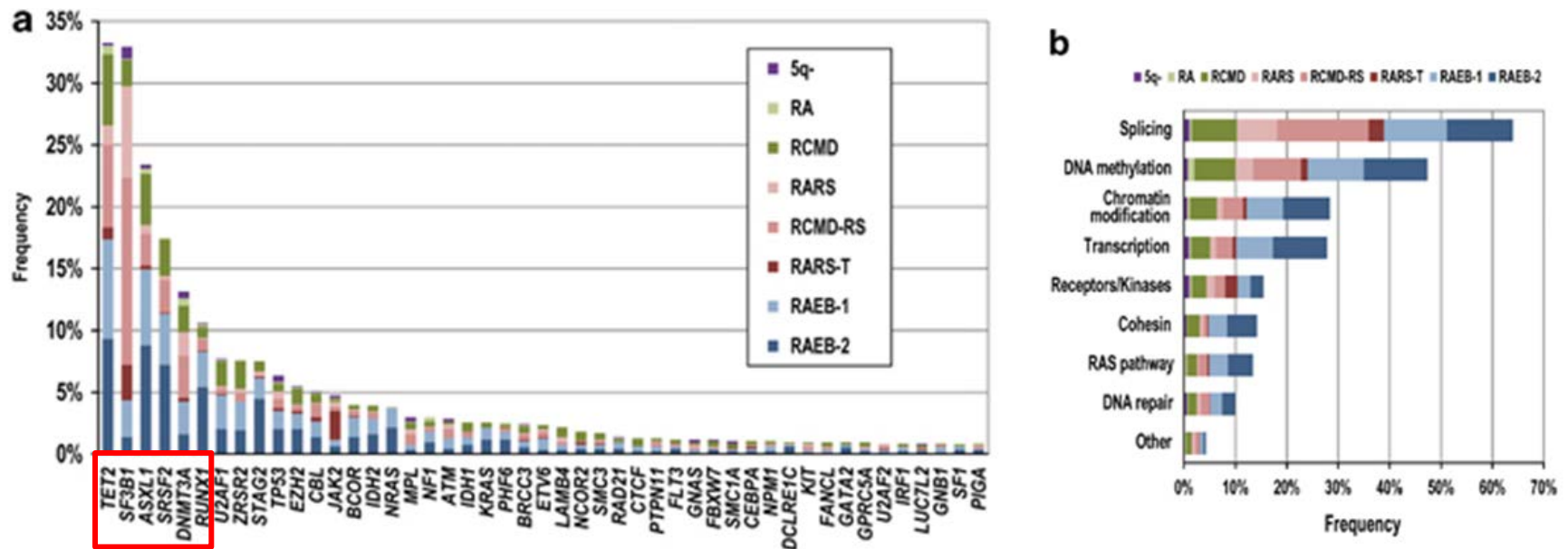
Cytogenetics and Prognosis

- Cytogenetic findings are categorized according to impact on median survival.
- These are a component of the Revised International Prognostic Scoring System (IPSS-R) for assessment of primary MDS.

Prognostic Subgroup	Cytogenetic Abnormalities	Median Survival (y)
Very Good	-Y, del(11q)	5.4
Good	Normal, del(5q), del(12p), del(20q), double including del(5q)	4.8
Intermediate	del(7q), +8, +19, i(17q), any other single or double independent clones	2.7
Poor	-7, inv(3)/t(3q)/del(3q), double including -7/del(7q), complex: 3 abnormalities	1.5
Very Poor	Complex: >3 abnormalities	0.7

Gene Mutations in MDS

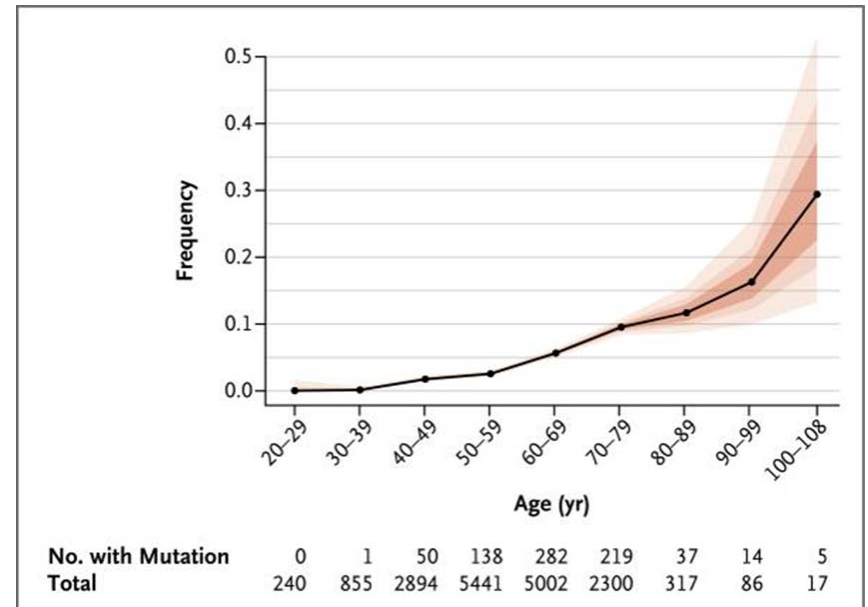
- 80-90% of MDS patients carry gene mutations



Haerlach T, et al. *Leukemia*. 2014;28(2):241-7.
 Papaemmanuil E, et al. *Blood*. 2013;122(22):3616-27.
 Malcovati L, et al. *Blood*. 2017;129(25):3371-78.

Gene Mutations and Diagnosis

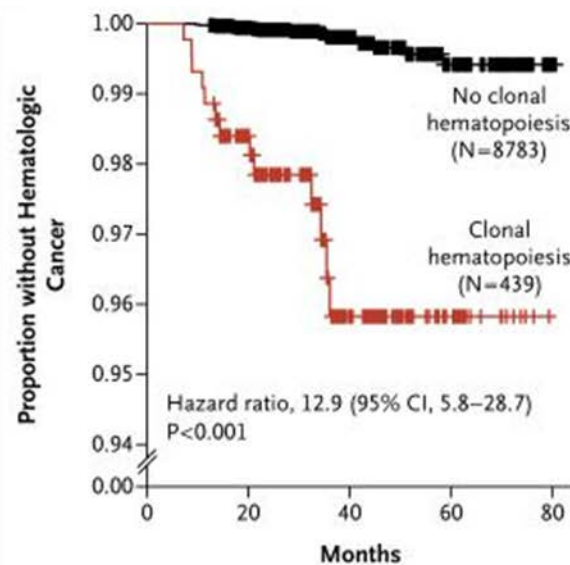
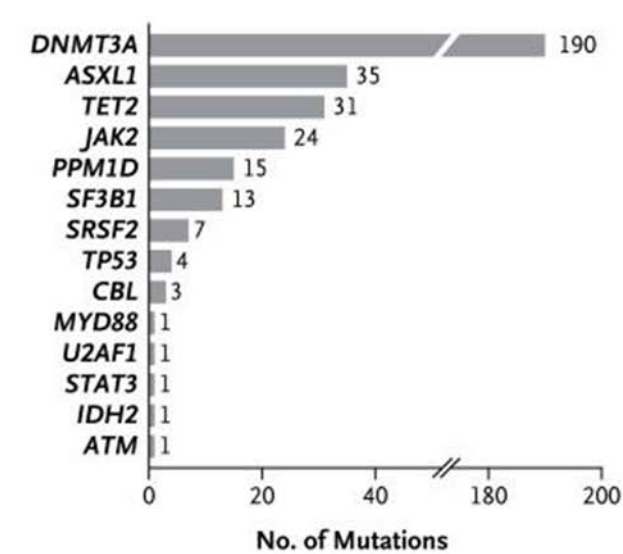
- Can gene mutations be used as *de facto* evidence of MDS in the absence of dysplasia, like certain cytogenetic abnormalities?
- Problem: many otherwise healthy older adults carry low-level somatic mutations in the same genes (CHIP or ARCH).



Jaiswal S, et al. *N Engl J Med.* 2014;371(26):2488-98.

Age-related Clonal Hematopoiesis

- Present in ~10% of healthy patients >65.
- Mostly in genes commonly mutated in MDS.
- These patients are more likely to develop subsequent hematologic malignancy, and have greater risk of CVD.



Genovese G. *N Engl J Med.* 2014;371(26):2477-87.
Jaiswal S, et al. *N Engl J Med.* 2014;371(26):2488-98.
Jaiswal S, et al. *N Engl J Med.* 2017;377(2):111-121

New Diagnostic Categories

	Traditional ICUS			MDS by WHO 2008	
	'Non-clonal' ICUS	CHIP	CCUS	Lower Risk MDS	Higher Risk MDS
Clonality	—	+	+	+	+
Dysplasia	—	—	—	+	+
Cytopenias	+	—	+	+	+
BM Blast %	< 5%	< 5%	< 5%	< 5%	< 19%
Overall Risk	Very Low	Very Low	Low (?)	Low	High
Treatments	Obs/BSC	Observation	Obs/BSC/GF	Obs/BSC/GF IMiD/IST	HMA/HCST

Clonal Cytopenias

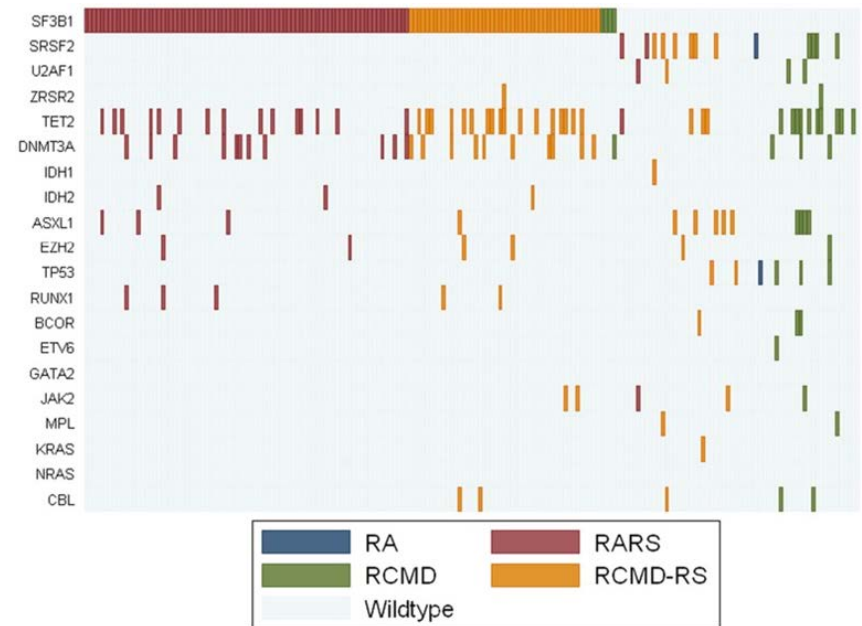
Gene Mutations and Diagnosis

- **Gene mutations cannot be used to make a diagnosis of MDS without dysplasia.**
- **Are there clues in the genotype that might predict increased likelihood of MDS?**
 - Number of mutations: ≥ 2 mutations in 64% of pre-clinical MDS vs. 8% in CHIP; PPV for MDS of ≥ 2 mutations = 0.88.
 - Median variant allele fraction (VAF): 40% in pre-clinical MDS vs. 9-10% in CHIP; PPV for MDS of one mutation with $\geq 10\%$ VAF = 0.86.
 - Particular genes: Mutations in spliceosome genes (*SF3B1*, *SRSF2*, *U2AF1*), *JAK2*, and *RUNX1* are most predictive of MDS, while mutations in *DNMT3A*, *ASXL1* or *TET2* are much less specific.

Cargo CA, et al. *Blood*. 2015;126(21):2362-5.
Malcovati L, et al. *Blood*. 2017;129(25):3371-78.

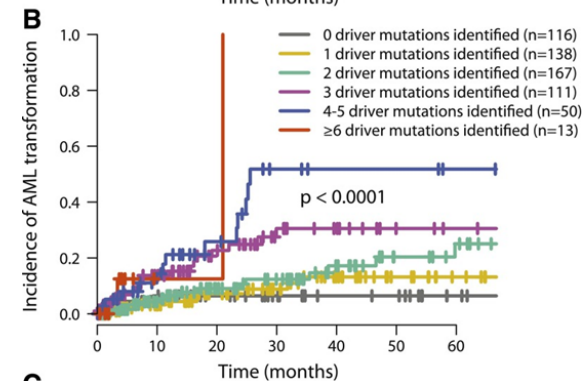
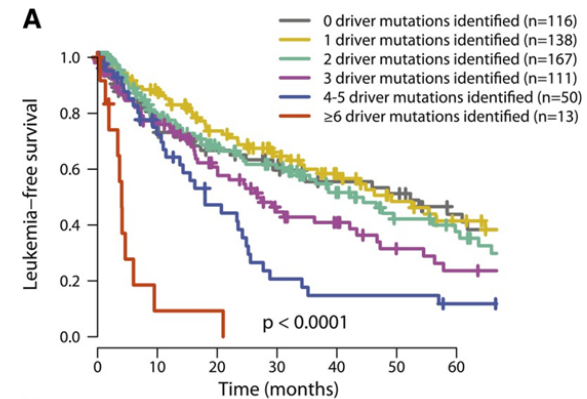
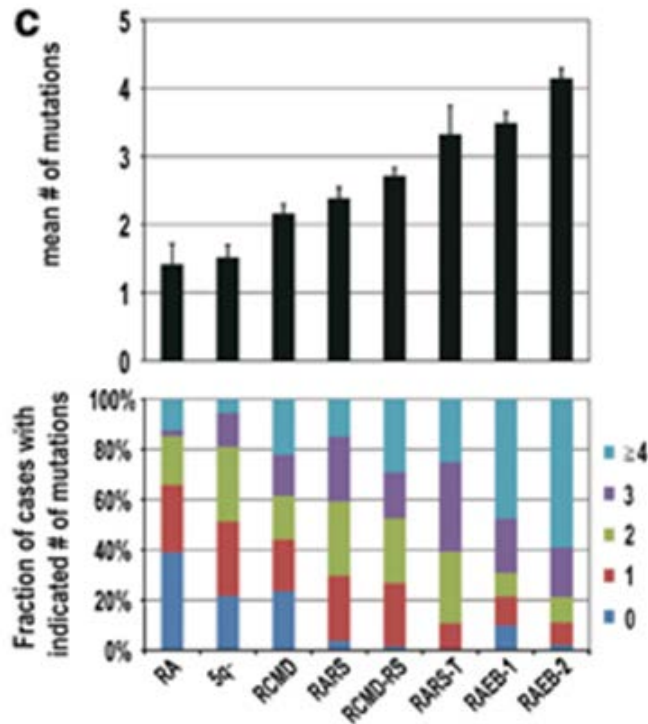
Morphologic Correlates

- Mutations in *SF3B1* are highly associated with the presence of ring sideroblasts and more favorable prognosis.
- The 2016 WHO update reduced the threshold percentage of RS required to make a diagnosis from 15% to 5% in the presence of *SF3B1* mutations.



Malcovati L, et al. *Blood*. 2015;126(2):233-41.
Papaemmanuil E, et al. *N Engl J Med*. 2011;365(15):1384-95.
Hasserjian RP, et al. *WHO Classification*. 2017:98

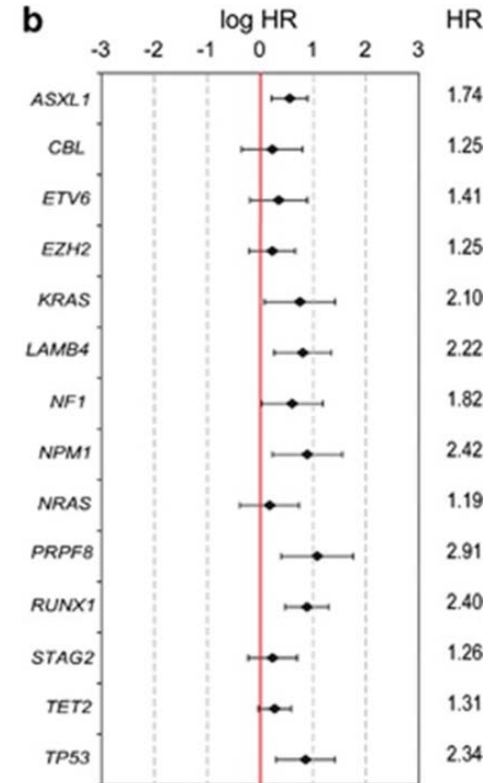
Gene Mutations and Prognosis



Haferlach T, et al. *Leukemia*. 2014;28:241
 Papaemmanuil E, et al. *Blood*. 2013;122:3616

Gene Mutations and Prognosis

- Mutations in particular genes impact prognosis:
 - Most consistently associated with poor outcome: *ASXL1*, *RUNX1*, and *TP53*
 - In particular, *TP53* mutations often lead to genomic instability, complex abnormal karyotype, and high risk of progression to AML.
 - Patients with *SF3B1* mutations have better outcomes.



Haferlach T, et al. *Leukemia*,. 2014;28:241
Bejar R, et al. *J Clin Oncol*. 2012;30:3376
Papaemmanuil E, et al. *Blood*. 2013;122:3616

Case Study Patient

- SF3B1 p.Lys700Glu (47%)
- DNMT3A p.Arg882Pro (46%)
- RUNX1 p.? (44%)
- BCORL1 p.Ser706* (14%)

Ring sideroblasts

RUNX1 +
4 mutations =
poor prognosis?

Patient rapidly progressed: bone marrow biopsy 4 months later showed increased blasts. Referred to hospice and passed away a few weeks later.

Summary – Part 1

- MDS is frequently associated with recurrent clonal somatic genetic abnormalities, detectable by karyotype and/or molecular studies.
- These have some utility in MDS diagnosis:
 - Certain clonal cytogenetic abnormalities can indicate MDS even in the absence of dysplasia.
 - Gene mutations can help diagnostically, but should be interpreted with caution due to age-related clonal hematopoiesis.
- Particular cytogenetic and molecular diagnostic findings are associated with distinctive morphologic findings and predict clinical outcomes.

Part 2: Genetic Testing in the Management and Therapy of Myelodysplastic Syndrome

Michael Savona, MD

Use of Molecular Testing in Clinical Care of MDS

- **Establishment of clonality and prognosis to guide therapy**
- **Targeted agents (clinical trials)***
- **?Future? Guided targeted therapy, and refinement of understanding of disease evolution**

Clonality and Prognosis and Molecular Lesions

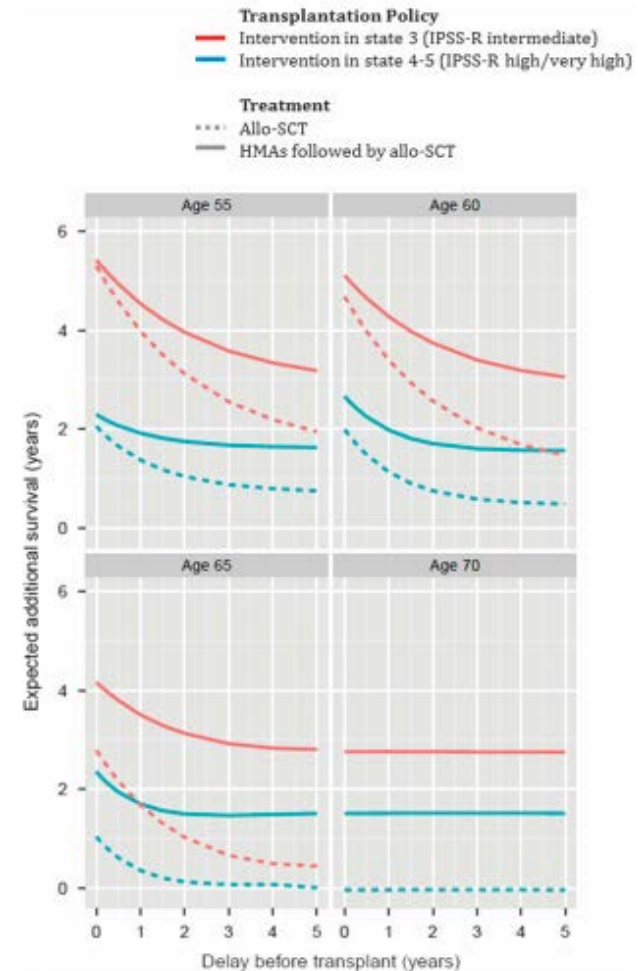
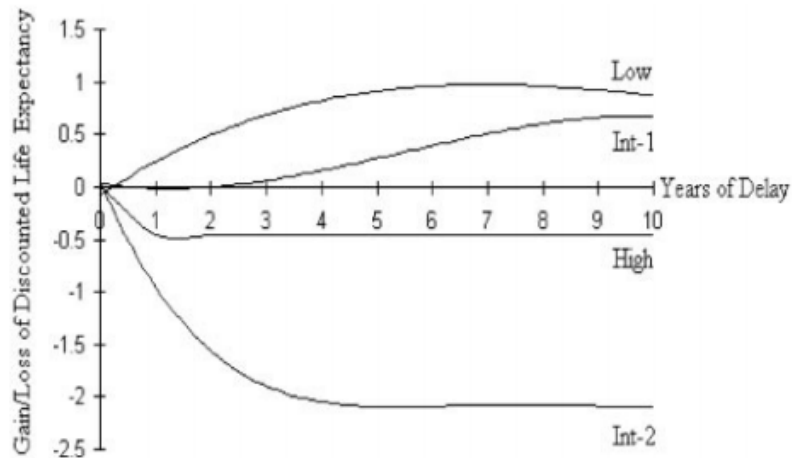
- **Most mutations have negative influence on prognosis in MDS**
 - *SF3B1* (isolated) is associated with MDS-RS /favorable risk MDS
 - Combinatory influences are TBD (eg, *SF3B1*+*RUNX1*)
- **Poor risk mutations often occur at loci shared in CHIP**
 - **so what does that mean?**
 - Large majority of CHIP does not evolve to neoplasia
 - However, presence of CCUS with higher risk mutations changes surveillance

Allogeneic Hematopoietic Stem Cell Transplant and Molecular Lesions

- **Usually, increased mutations occur in correlation to complexity of cytogenetic lesions and/or increased blasts (but not always)**
- **Number of mutations, high risk mutations account for high risk disease**
 - Should this lead to HSCT?
 - Should mutations disqualify from HSCT?

Decision Analysis in MDS is Largely Based on IPSS

- Biased by deficiencies of IPSS
- If higher risk MDS (by IPSS) is enriched for mutations → logically, transplant patients with high risk defined by mutations



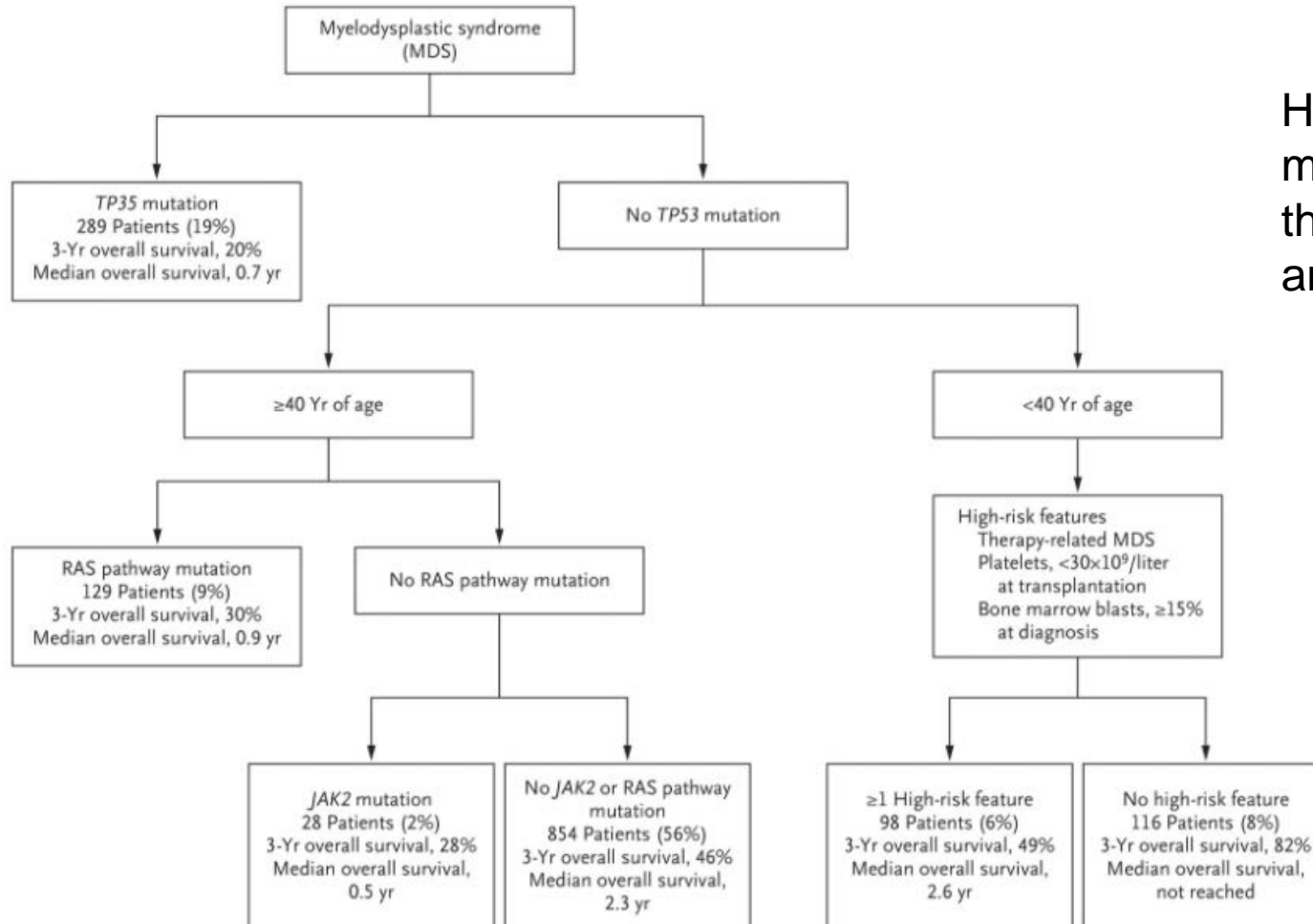
Cutler et al. *Blood*. 2004
Della Porta et al. *Leukemia*. 2017

Mutations added to AlloHSCT Decision analysis

- Higher IPSS Risk may be associated with worse molecular profile, but not always
- As mentioned, shorter survival is associated with:
 - More mutations
 - Allele burden
 - High risk mutations

Haferlach T, et al. *Leukemia*, 2014;28:241
Bejar R, et al. *J Clin Oncol*. 2012;30:3376
Papaemmanuil E, et al. *Blood*. 2013;122:3616

Isolated High Risk Mutations Imply Miserable alloH SCT Outcomes



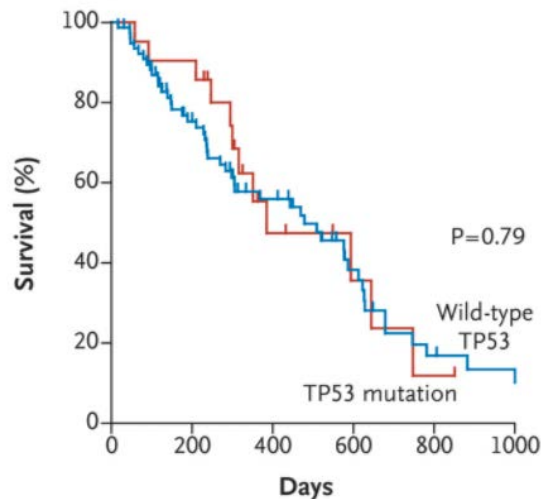
How to add mutational status to the HSCT decision analysis?

Lindsley et al, *NEJM*. 2017.

TP53 Mutations Respond to Decitabine*10days

- Phase 2 – 84 patients
- 21/21 patients (100%) with *TP53* mutations with mCR/CR

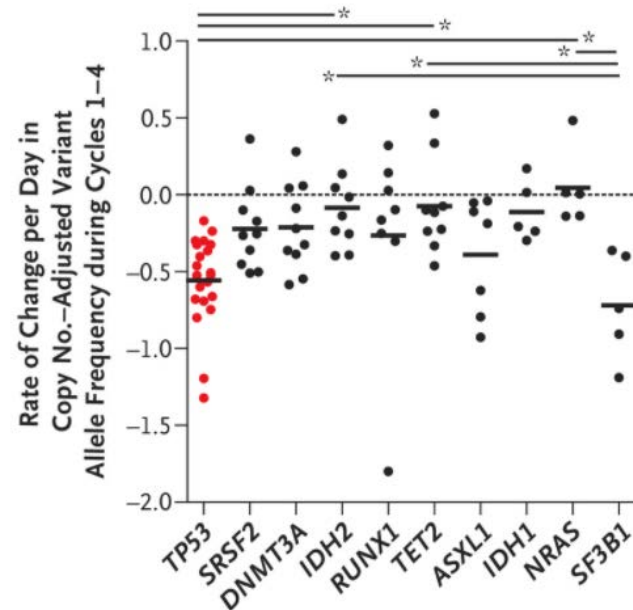
D Survival According to *TP53* Mutation



No. at Risk

<i>TP53</i> mutation	21	20	7	4	2
Wild-type <i>TP53</i>	78	51	31	16	7

E Clearance of Mutations



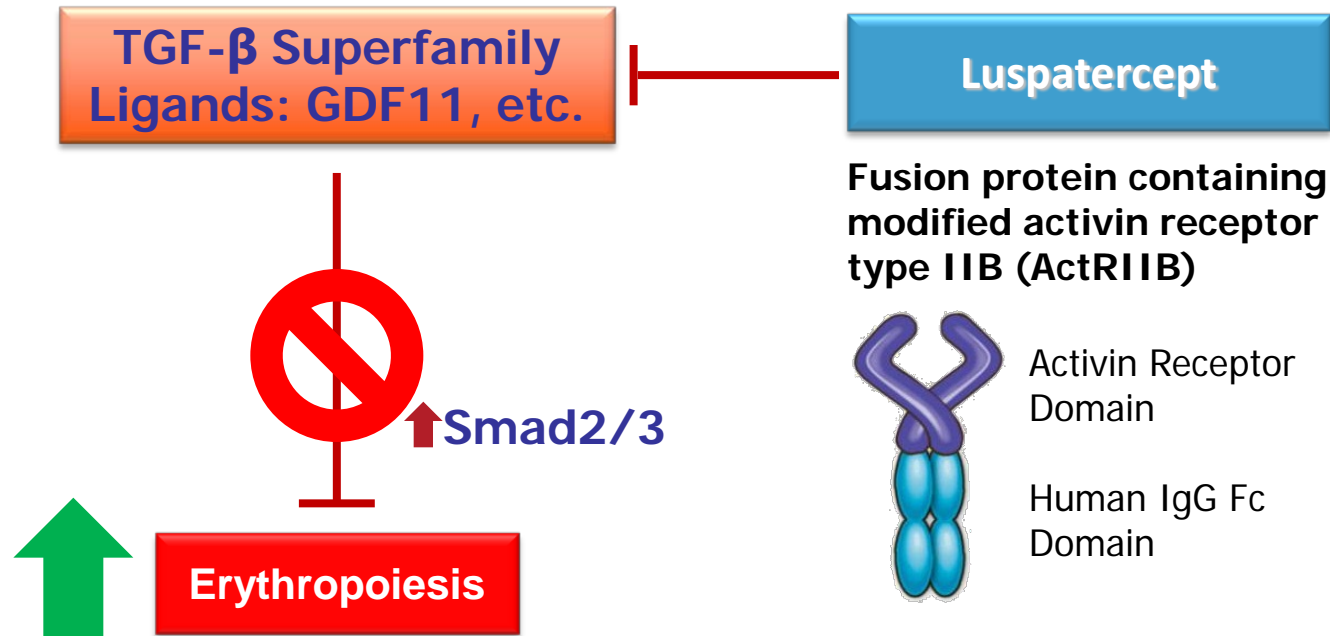
Welch et al, *NEJM*. 2016.

The Future

SF3B1 – Targeting “Good Risk”

- ***SF3B1* mutations are associated with RARS and favorable risk MDS, but are associated with chronic anemia, transfusion dependence, iron overload, and diminished quality of life**
- **Luspatercept / Sotatercept TGFb ligand trap analogues**

Luspatercept in MDS

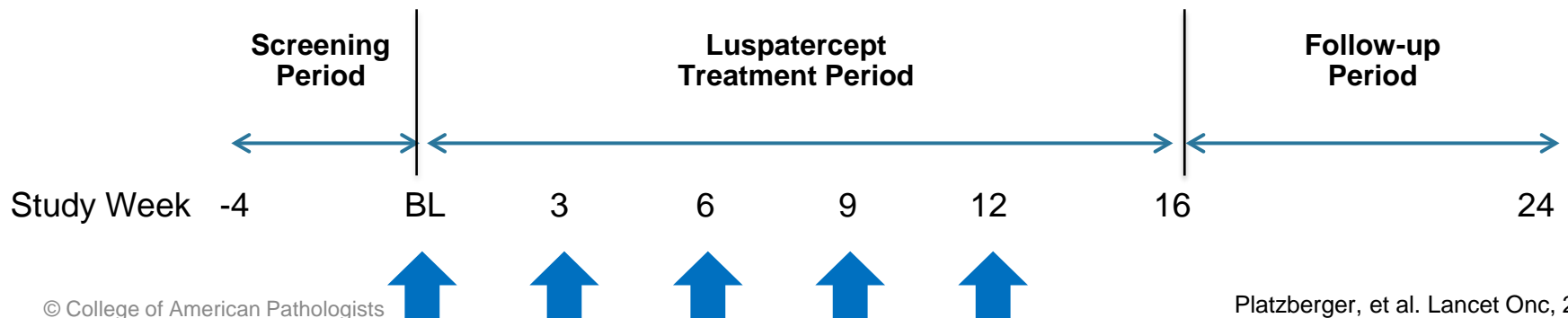


- Mechanism is distinct from erythropoietin
- Acts on late-stage erythropoiesis to increase mature RBCs in the circulation

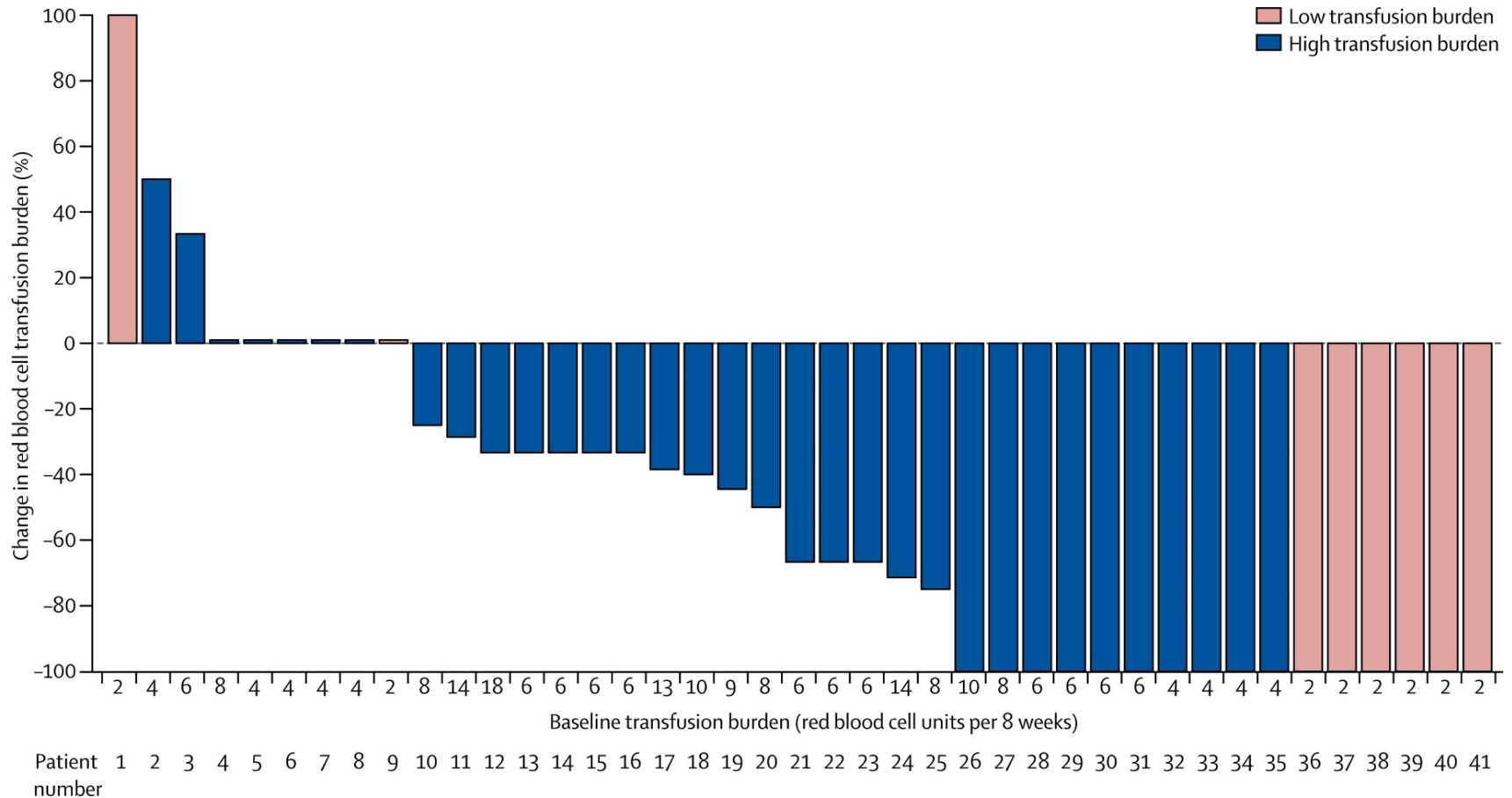
Suragani R, et al. Nature Med 2014
Zhou L, et al., Blood 2008

Luspatercept PACE-MDS Study Overview

- Phase 2, multicenter, open-label, dose-finding study in IPSS low/int-1 MDS
- Eligibility criteria:** EPO >500 U/L or nonresponsive/refractory to ESA; no prior azacitidine or decitabine; no current lenalidomide, ESA, G-CSF
- Primary efficacy endpoints**
 - Low Transfusion Burden (LTB, <4U RBC/8 weeks, Hgb <10 g/dL): Hemoglobin increase of ≥ 1.5 g/dL for ≥ 2 weeks
 - High Transfusion Burden (HTB, ≥ 4 U RBC/8 weeks): Reduction of ≥ 4 U or $\geq 50\%$ units transfused over 8 weeks
- Luspatercept administered SC every 3 weeks for 3 months (base study → extension)

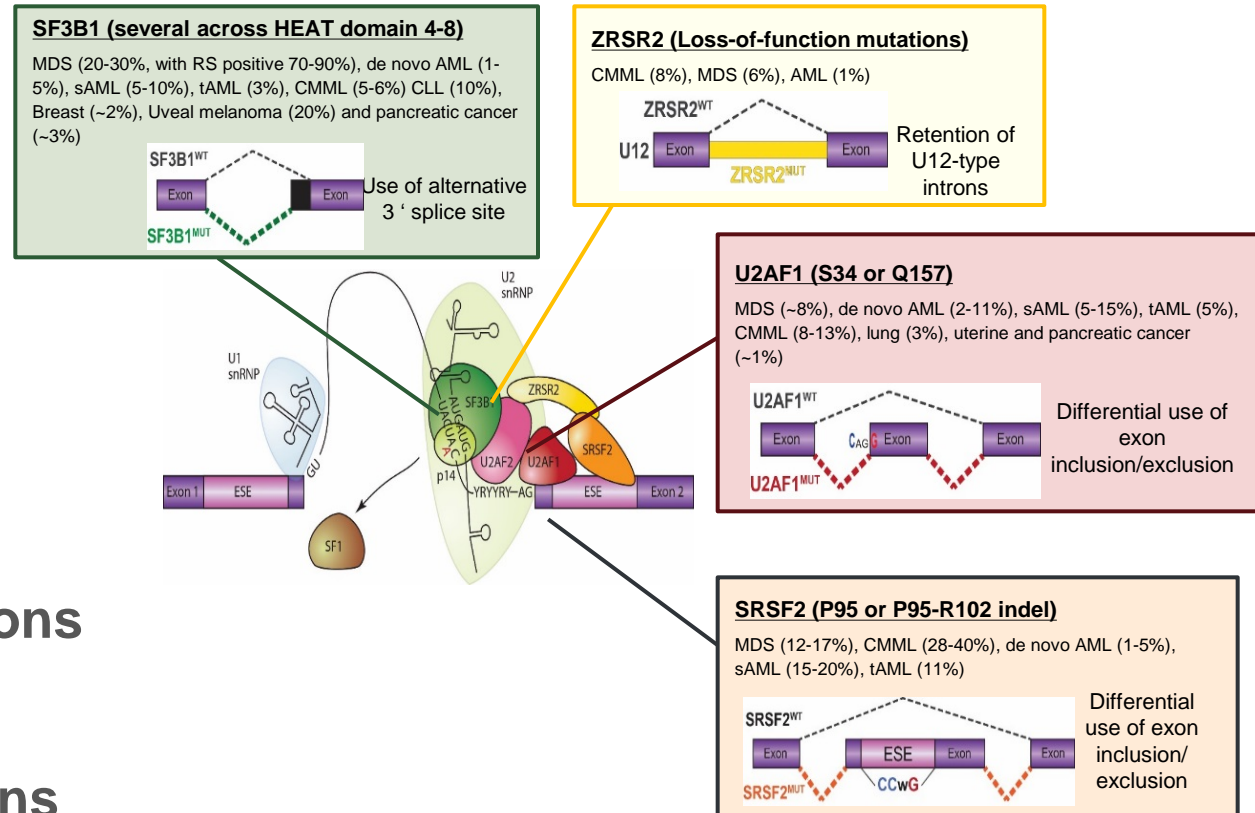


MAXIMUM PERCENTAGE CHANGE IN RBC TRANSFUSION BURDEN

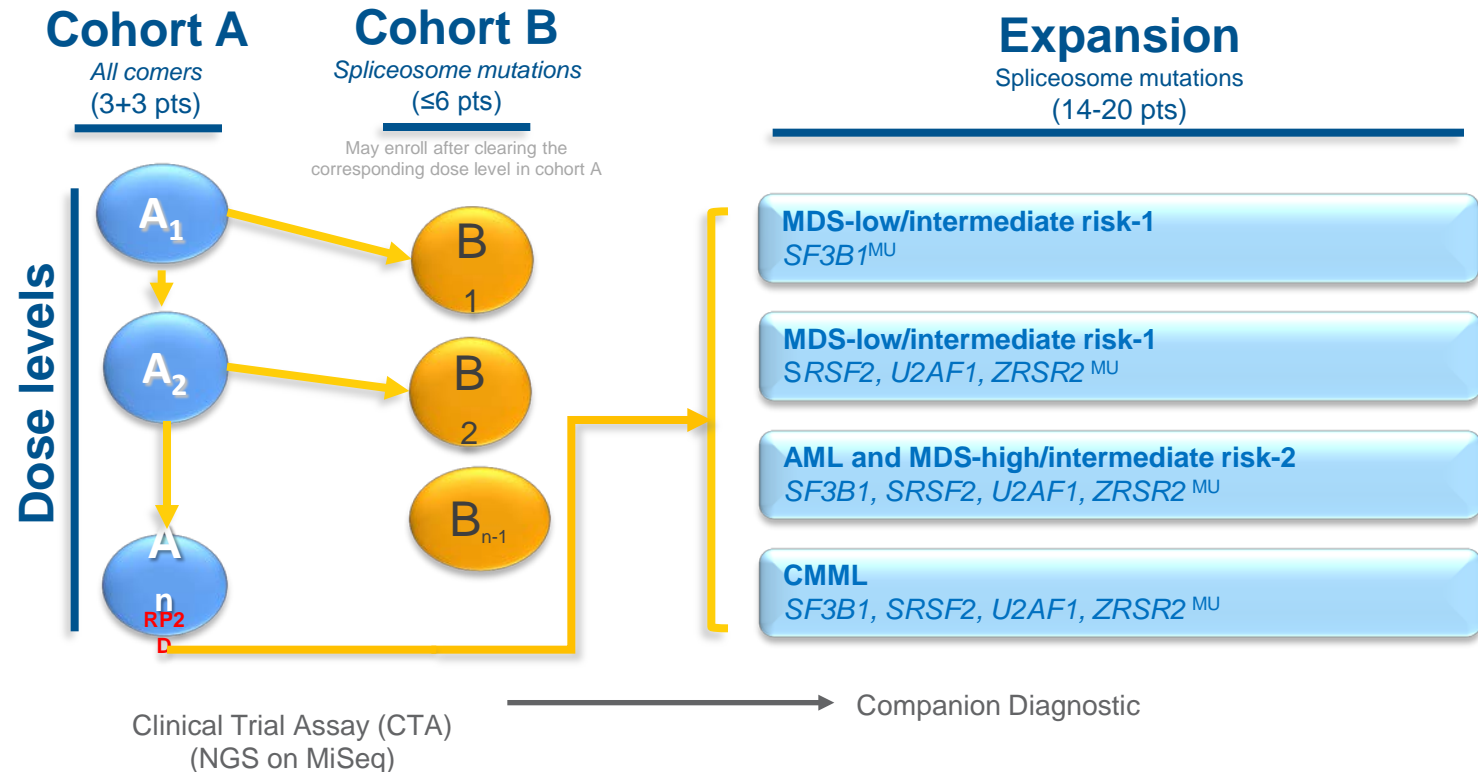


Addressing Spliceosome Malfunction

- **Recurrent mutations observed in *SF3B1*, *U2AF1*, *SRSF2* and *ZRSR2***
- **Heterozygous mutations & mutually exclusive**
- **Spliceosome mutations cause aberrant splicing**

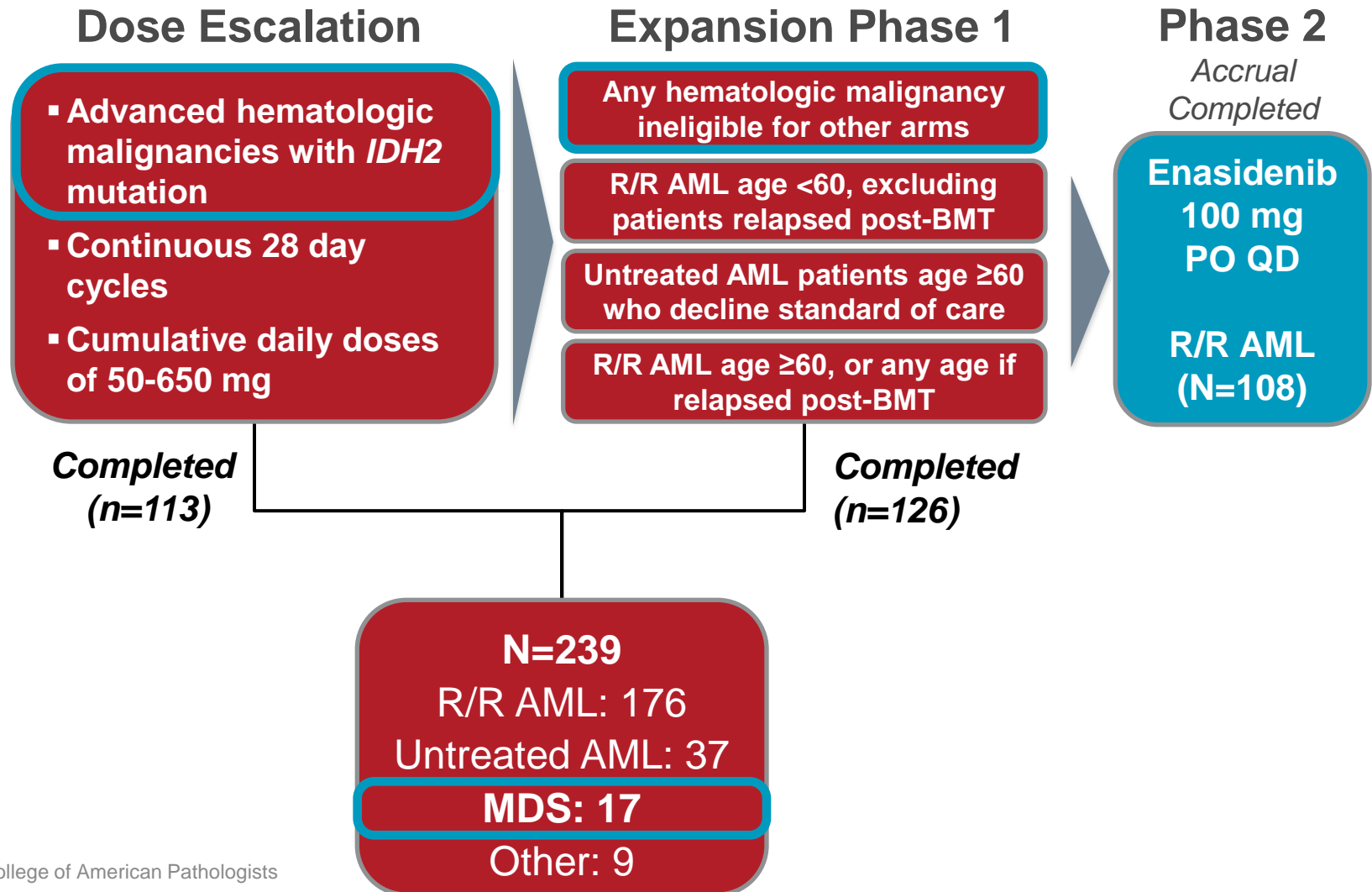


H3B-8800 Phase I Study



- **Eligibility:** Cohort A - AML, MDS, CMML; Cohort B – spliceosome mutant.

Enasidenib (IDH2 inhibitor) Phase 1/2 Dose-escalation and Expansion



Response

	MDS Patients (N=17) n/N (%)
Overall response rate (CR + PR + mCR + HI)	10/17 (59)
Best Response	
Complete remission*	1/11 (9)
Partial remission*	1/11 (9)
Marrow CR*	3/11 (27)
Any hematologic improvement (HI) [†]	5/17 (29)
HI-E	3/15 (20)
HI-P	4/12 (33)
HI-N	4/10 (40)
<p>*Investigator-assessed; evaluable patients had ≥5% bone marrow blasts at baseline</p> <p>[†]HI was programmatically adjudicated per IWG 2006 criteria for MDS; denominators reflect eligibility for response</p> <p>CR, complete remission; PR, partial remission; mCR, marrow CR; HI, hematologic improvement</p>	

- **Of 13 patients who had received prior HMA therapy, 7 (54%) had a response with enasidenib**
- **Of patients who attained HI, 2 had trilineage and 2 had bilineage improvement**
- **Median time to response was 21 days (range 10-87)**

Summary – Part 2

- Currently, the mutational profile of MDS has some impact on patient management:
 - Surveillance in patients with CCUS
 - Prognosis, risk of progression, and transplant decision
 - Decitabine in patients with TP53 mutations
- The future holds promise for targeting pathways involved in the pathogenesis of MDS

Questions?

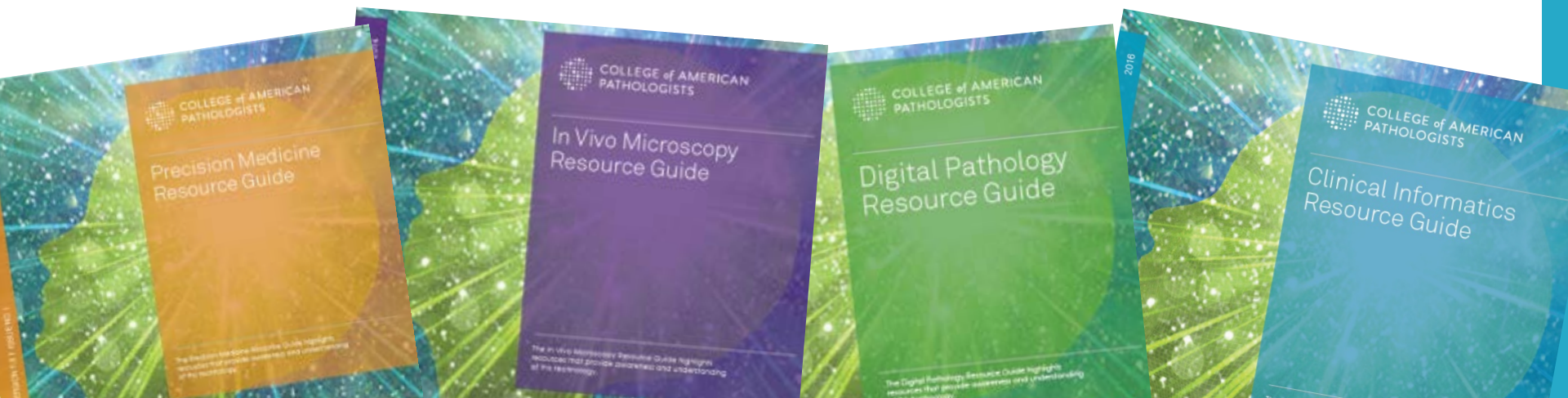
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DATE	TOPIC	SPEAKER(s)
May 2, 2018	HPV Testing on Head and Neck Carcinomas: A Review of the CAP Guidelines	Justin Bishop, MD
June 13, 2018	New Guideline for Lung Cancer Biomarker Testing: Essentials and Applications	Philip Cagle, MD & Eric Bernicker, MD

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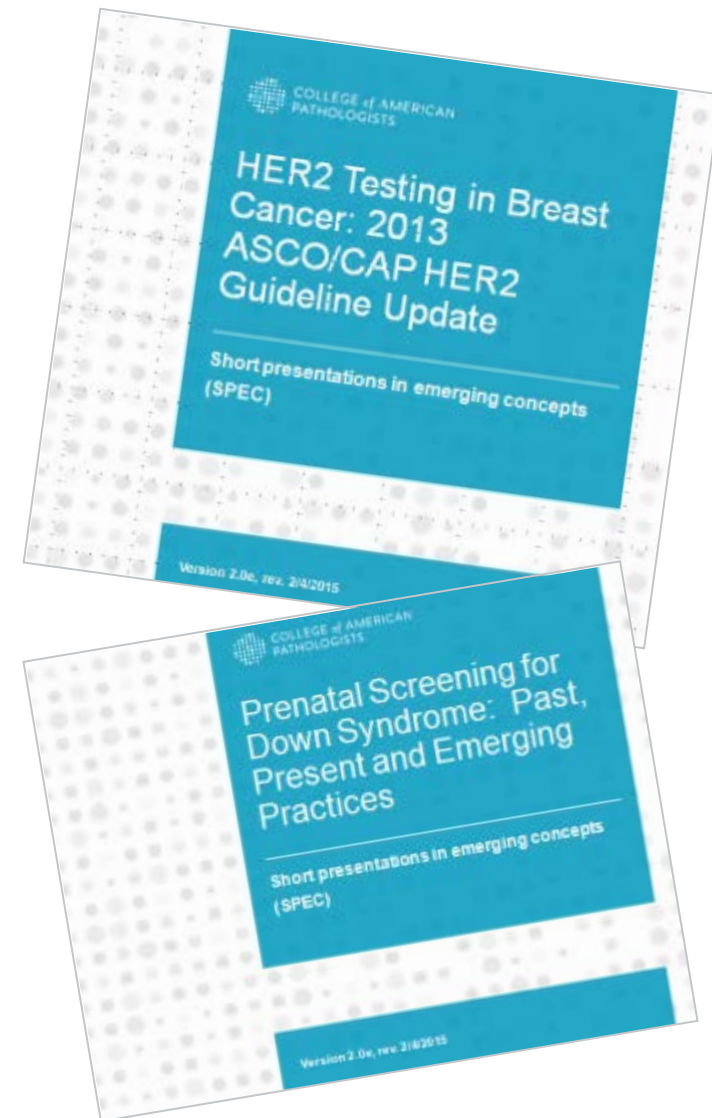
CAP's Pathology Resource Guide: Precision Medicine

- The CAP has created the Pathology Resource Guides to assist pathologists in understanding key emerging technologies.
 - Printed guides are now available for members (\$39) and non-members (\$69)
 - The digital copy of the Resource Guides are a complimentary member benefit
 - Access them www.cap.org > Resources and Publications



Short Presentations on Emerging Concepts (SPECS)

- Pathology SPECs are:
 - short PowerPoints, created for pathologists
 - Focused on diseases where molecular tests play a key role in patient management
- **Topics include** Renal Tumors, cell free DNA (cfDNA), and PD-L1 as well as other emerging topics
- Access them www.cap.org > Resources and Publications





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THANK YOU!

Thank you for attending our webinar,
**“Molecular Diagnostic Testing in Myelodysplastic Syndrome” by
Adam Seegmiller, MD, PhD &
Michael Savona, MD**

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