

# Protocol for the Examination of Biopsy Specimens from Patients with Invasive Carcinoma of the Breast

**Version:** 1.1.1.2

Protocol Posting Date: September 2022

The use of this protocol is recommended for clinical care purposes but is not required for accreditation

purposes.

## This protocol may be used for the following procedures AND tumor types:

Procedure	Description
Biopsy	Includes specimens designated needle biopsy, fine needle aspiration and others (for excisional biopsy, see below)
Tumor Type	Description
Invasive breast carcinoma of any	Includes microinvasive carcinoma and carcinoma with neuroendocrine features
type, with or without ductal	
carcinoma in situ (DCIS)	

## The following should NOT be reported using this protocol:

Procedure
Resection (consider Breast Invasive Carcinoma Resection protocol)
Excisional biopsy (consider Breast Invasive Carcinoma Resection protocol)
Tumor Type
Ductal carcinoma in situ (DCIS) without invasive carcinoma (consider the DCIS Biopsy protocol)
Paget disease of the nipple without invasive carcinoma (consider the DCIS Biopsy protocol)
Encapsulated or solid papillary carcinoma without invasion (consider the Breast DCIS Biopsy protocol)
Phyllodes tumor
Lymphoma (consider the Hodgkin or non-Hodgkin Lymphoma protocols)
Sarcoma (consider the Soft Tissue protocol)

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With guidance from the CAP Cancer and CAP Pathology Electronic Reporting Committees.

## **Accreditation Requirements**

The use of this case summary is recommended for clinical care purposes but is not required for accreditation purposes. The core and conditional data elements are routinely reported. Non-core data elements are indicated with a plus sign (+) to allow for reporting information that may be of clinical value.

<sup>\*</sup> Denotes primary author.

# **Summary of Changes**

# v 1.1.1.2

• Updated Explanatory Note A

# **Reporting Template**

**Protocol Posting Date: September 2022** 

Select a single response unless otherwise indicated.

# CASE SUMMARY: (INVASIVE CARCINOMA OF THE BREAST: Biopsy)

This template is recommended for reporting biopsy specimens, but is not required for accreditation purposes.

	, ,
SPECIMEN	
Procedure	
Needle biopsy	
Fine needle aspiration	
Other (specify):	
Not specified	
Specimen Laterality	
Right	
Left	
Not specified	
TUMOR	
+Tumor Site (select all that apply)	
Upper outer quadrant	
Lower outer quadrant	
Upper inner quadrant	
Lower inner quadrant	
Central	
Nipple	
Clock position	
Specify Clock Position (select all that apply)	
1 o'clock	
2 o'clock	
3 o'clock	
4 o'clock	
5 o'clock	
6 o'clock	
7 o'clock	
8 o'clock	
9 o'clock	
10 o'clock	
11 o'clock	
12 o'clock	
Specify distance from nipple in Centimeters (cm):	cm
Other (specify):	
Not specified	

Histo	ologic Type (Note <u>A</u> )
	No residual invasive carcinoma
	Invasive carcinoma of no special type (ductal)
I	Micro-invasive carcinoma
	Invasive lobular carcinoma
	Invasive carcinoma with mixed ductal and lobular features
	Invasive carcinoma with features of (specify):
	Tubular carcinoma
	Invasive cribriform carcinoma
	Mucinous carcinoma
	Invasive micropapillary carcinoma
	Apocrine adenocarcinoma
	Metaplastic carcinoma
	Encapsulated papillary carcinoma with invasion
	Solid papillary carcinoma with invasion
	Intraductal papillary adenocarcinoma with invasion
	Adenoid cystic carcinoma
I	Neuroendocrine tumor
ا	Neuroendocrine carcinoma
	Invasive carcinoma, type cannot be determined:
(	Other histologic type not listed (specify):
+}	Histologic Type Comment:
I	Not applicable (no residual carcinoma or microinvasion only) Nottingham Score landular (Acinar) / Tubular Differentiation
	_ Score 1 (greater than 75% of tumor area forming glandular / tubular structures)
	_ Score 2 (10% to 75% of tumor area forming glandular / tubular structures)
	_ Score 3 (less than 10% of tumor area forming glandular / tubular structures)
	Score cannot be determined:
N	uclear Pleomorphism
_	_ Score 1 (Nuclei small with little increase in size in comparison with normal breast epithelial cells
re	gular outlines, uniform nuclear chromatin, little variation in size)
	Score 2 (Cells larger than normal with open vesicular nuclei, visible nucleoli, and moderate
va	ariability in both size and shape)
_	_ Score 3 (Vesicular nuclei, often with prominent nucleoli, exhibiting marked variation in size and
sh	nape, occasionally with very large and bizarre forms)
	Score cannot be determined:
	itotic Rate
	e Table 1 in CAP Protocol.
	_ Score 1
_	_ Score 2
	_ Score 3
	Score cannot be determined:verall Grade
O.	Grade 1 (scores of 3, 4 or 5)
	_ Glade 1 (300163 01 3, 4 01 3)

Grade 2 (scores of 6 or 7)
Grade 3 (scores of 8 or 9)
Score cannot be determined (explain):
+Tumor Size
Microinvasion only (less than or equal to 1 mm)
Greatest dimension of largest invasive focus greater than 1 mm (specify exact measurement in
Millimeters (mm)): mm
+Additional Dimension in Millimeters (mm): x mm
Tumor size cannot be determined (explain):
Ductal Carcinoma In Situ (DCIS) (Note C)
Not identified
Present
Architectural Patterns (select all that apply)
Comedo
Paget disease (DCIS involving nipple skin)
Cribriform
Micropapillary
Papillary
Solid
Other (specify):
Nuclear Grade
Grade I (low)
Grade II (intermediate)
Grade III (high)
Other (specify):
Cannot be determined:
Necrosis
Not identified
Present, focal (small foci or single cell necrosis)
Present, central (expansive "comedo" necrosis)
Other (specify):
Cannot be determined:
Cannot be excluded
+Lymphovascular Invasion
Not identified
Present
Cannot be determined:
+Microcalcifications (Note D) (select all that apply)
Not identified
Present in DCIS
Present in invasive carcinoma
Present in non-neoplastic tissue
Other (specify):
Onler (appenity)

ADDITIONAL FINDINGS (Note <u>E</u> )
+Additional Findings (specify):
SPECIAL STUDIES  For hormone receptor and HER2 reporting, the CAP Breast Biomarker Template should be used. www.cap.org/cancerprotocols.
+Breast Biomarker Studies (specify pending studies):
COMMENTS
Comment(s):

# **Explanatory Notes**

# A. Histologic Type

This protocol applies to all invasive carcinomas of the breast. The World Health Organization (WHO)¹ classification of breast carcinoma is recommended, although the protocol does not preclude the use of other classifications or histologic types. Carcinomas may be classified based on the H&E appearance without the use of immunohistochemical studies.

A modified list is presented in the case summary based on the most frequent types of invasive carcinomas and terminology that is in widespread usage. The modified list is intended to capture the majority of tumors and reduce the frequency of tumors being reported as "other." Choices are added for tumors with mixed features and those with some but not all features of specific histologic types.

WHO Classification of Invasive Carcinoma of the Breast <sup>1</sup>
No residual invasive carcinoma
Invasive carcinoma of no special type (ductal)
Micro-invasive carcinoma
Invasive lobular carcinoma
Invasive carcinoma with mixed ductal and lobular features
Invasive carcinoma with mixed features (specify):
Tubular carcinoma
Invasive cribriform carcinoma
Mucinous carcinoma
Invasive micropapillary carcinoma
Apocrine adenocarcinoma
Metaplastic Carcinoma
Metaplastic carcinoma NOS
Low grade adenosquamous carcinoma
Fibromatosis-like metaplastic carcinoma
Spindle cell carcinoma
Squamous cell carcinoma
Metaplastic carcinoma with mesenchymal differentiation
Neuroendocrine Tumor
Neuroendocrine tumor NOS
Neuroendocrine tumor, grade 1
Neuroendocrine tumor, grade 2
Neuroendocrine Carcinoma
Neuroendocrine carcinoma NOS
Neuroendocrine carcinoma, small cell
Neuroendocrine carcinoma, large cell
Invasive carcinoma, type cannot be determined
Other histologic type (specify):
Invasive papillary carcinoma
Encapsulated papillary carcinoma with invasion

Solid papillary carcinoma with invasion
Intraductal papillary adenocarcinoma with invasion
Oncocytic carcinoma
Lipid-rich carcinoma
Glycogen-rich carcinoma
Sebaceous carcinoma
Mucinous cystadenocarcinoma NOS
Acinar cell carcinoma
Classic adenoid cystic carcinoma
Solid-basaloid adenoid cystic carcinoma
Adenoid cystic carcinoma with high-grade transformation
Secretory carcinoma
Mucoepidermoid carcinoma
Polymorphous adenocarcinoma
Tall cell carcinoma with reversed polarity
Adenomyoepithelioma with carcinoma
Epithelial-myoepithelial carcinoma
Other type not listed (specify):

#### References

1. WHO Classification of Tumours Editorial Board. <em>Breast tumours</em>. Lyon (France): International Agency for Research on Cancer; 2019. (WHO classification of tumours series, 5th ed.; vol. 2)

## **B.** Histologic Grade

All invasive breast carcinomas should be graded. The Nottingham combined histologic grade (Elston-Ellis modification of Scarff-Bloom-Richardson grading system) should be used for reporting. Within each stage grouping there is a relation between histologic grade and outcome.

The Nottingham combined histologic grade evaluates the amount of tubule formation, the extent of nuclear pleomorphism, and the mitotic count (or mitotic rate). Each variable is given a score of 1, 2, or 3, and the scores are added to produce a grade. The mitotic score is determined by the number of mitotic figures found in 10 consecutive high-power fields (HPF) in the most mitotically active part of the tumor. Only clearly identifiable mitotic figures should be counted; hyperchromatic, karyorrhectic, or apoptotic nuclei are excluded. Because of variations in field size, the HPF size must be determined for each microscope and the appropriate point score determined accordingly. It is recommended that the size be measured by using a micrometer. However, the diameter of an HPF can also be calculated by using the method below.

## Measuring the Size of a High-Power Field (HPF) With a Ruler

Use a clear ruler to measure the diameter of a low-power field. This number can be used to calculate a constant based on the following formula:

Eyepiece Magnification x Objective Magnification x Microscopic Field Diameter = A Constant

When the value of the constant is known, the diameter of an HPF can be calculated for other objectives by using the following formula:

# <u>Unknown Field Diameter = Constant/(Eyepiece Magnification x Objective Magnification)</u>

Half of the field diameter is the radius of the field (r), which can then be used to calculate the area of the HPF:

## 3.1415 x r<sup>2</sup>= Area of Microscopic Field

If the microscopic field diameter or the area of the field is known, Table 1 can be used to determine the number of mitoses corresponding to different scores.

Table 1. Score Categories According to Field Diameter and Mitotic Count

Scoring Categories of Mitotic Counts				
Number of mitoses per 10 fields corresponding to:				esponding to:
Field diameter (mm)	Area (mm²)	Score 1	Score 2	Score 3
0.40	0.125	≤4	5 to 9	≥10
0.41	0.132	≤4	5 to 9	≥10
0.42	0.139	≤5	6 to 10	≥11
0.43	0.145	≤5	6 to 10	≥11
0.44	0.152	≤5	6 to 11	≥12
0.45	0.159	≤5	6 to 11	≥12
0.46	0.166	≤6	7 to 12	≥13
0.47	0.173	≤6	7 to 12	≥13
0.48	0.181	≤6	7 to 13	≥14
0.49	0.189	≤6	7 to13	≥14
0.50	0.196	≤7	8 to 14	≥15
0.51	0.204	≤7	8 to 14	≥15
0.52	0.212	≤7	8 to 15	≥16
0.53	0.221	≤8	9 to 16	≥17
0.54	0.229	≤8	9 to 16	≥17
0.55	0.238	≤8	9 to 17	≥18
0.56	0.246	≤8	9 to 17	≥18
0.57	0.255	≤9	10 to 18	≥19
0.58	0.264	≤9	10 to 19	≥20
0.59	0.273	≤9	10 to 19	≥20
0.60	0.283	≤10	11 to 20	≥21
0.61	0.292	≤10	11 to 21	≥22
0.62	0.302	≤11	12 to 22	≥23
0.63	0.312	≤11	12 to22	≥23
0.64	0.322	≤11	12 to 23	≥24
0.65	0.332	≤12	13 to 24	≥25
0.66	0.342	≤12	13 to 24	≥25
0.67	0.353	≤12	13 to 25	≥26
0.68	0.363	≤13	14 to 26	≥27
0.69	0.374	≤13	14 to 27	≥ 28

From Pathology Reporting of Breast Disease. 2 Copyright 2005 National Health Service Cancer Screening Programme and The Royal College of Pathologists. Adapted with permission.

#### References

- 1. Ellis IO, Elston CW. Histologic grade. In: O'Malley FP, Pinder SE, eds. *Breast Pathology*. Philadelphia, PA: Elsevier; 2006:225-233.
- Pathology Reporting of Breast Disease. A Joint Document Incorporating the Third Edition of the NHS Breast Screening Programme's Guidelines for Pathology Reporting in Breast Cancer Screening and the Second Edition of The Royal College of Pathologists' Minimum Dataset for Breast Cancer Histopathology Published by the NHS Cancer Screening Programmes jointly with The Royal College of Pathologists. NHSBSP Publication No 58. January 2005. http://www.cancerscreening.nhs.uk/ breastscreen/publications/nhsbsp58.html. Accessed April 8, 2009.

#### C. Ductal Carcinoma In Situ

**Nuclear Grade of DCIS** 

The nuclear grade of DCIS is determined using 6 morphologic features (Table 1).1

Table 2. Nuclear Grade of Ductal Carcinoma in Situ

Feature	Grade I (Low)	Grade II (Intermediate)	Grade III (High)
Pleomorphism	Monotonous (monomorphic)	Intermediate	Markedly pleomorphic
Size	1.5 to 2 x the size of a normal red blood cell or a normal duct epithelial cell nucleus	Intermediate	>2.5 x the size of a normal red blood cell or a normal duct epithelial cell nucleus
Chromatin	Usually diffuse, finely dispersed chromatin	Intermediate	Usually vesicular with irregular chromatin distribution
Nucleoli	Only occasional	Intermediate	Prominent, often multiple
Mitoses	Only occasional	Intermediate	May be frequent
Orientation	Polarized toward luminal spaces	Intermediate	Usually not polarized toward the luminal space

#### **Necrosis**

The presence of necrosis is correlated with the finding of mammographic calcifications (i.e., most areas of necrosis will calcify). Ductal carcinoma in situ that presents as mammographic calcifications often recurs as calcifications. Necrosis can be classified as follows:

<u>Central ("comedo")</u>: The central portion of an involved ductal space is replaced by an area of expansive necrosis that is easily detected at low magnification. Ghost cells and karyorrhectic debris are generally present. Although central necrosis is generally associated with high-grade nuclei (i.e., comedo DCIS), it can also occur with DCIS of low or intermediate nuclear grade.

<u>Focal</u>: Small foci, indistinct at low magnification, or single cell necrosis.

Necrosis should be distinguished from secretory material, which can also be associated with calcifications, but does not include nuclear debris.

#### References

1. Schwartz GF, Lagios MD, Carter D, et al. Consensus conference on the classification of ductal carcinoma in situ. *Cancer.* 1997;80:1798-1802.

#### D. Microcalcifications

Cancer found in biopsies performed for microcalcifications will almost always be at the site of the calcifications or in close proximity. The presence of the targeted calcifications in the specimen should be confirmed by specimen radiography. The pathologist must be satisfied that the specimen has been sampled in such a way that the lesion responsible for the calcifications has been examined microscopically. The relationship of the radiologic calcifications to the invasive carcinoma and the DCIS should be indicated.

If calcifications can be seen in the specimen radiograph but not in the initial histologic sections, deeper levels should be examined. If needed, radiographs of the paraffin block(s) may be obtained to detect calcifications remaining in the block(s). If microcalcifications cannot be confirmed by routine microscopic evaluation, polarized light may be helpful, since calcium oxalate crystals are refractile and polarizable but usually clear or tinged yellow in H&E sections. On rare occasions, calcifications do not survive tissue processing or prolonged fixation in formalin. Foreign material can sometimes simulate calcifications (e.g., metallic fragments after surgery or trauma).

#### E. Additional Findings

In some cases, additional pathologic findings are important for the clinical management of patients. If multiple invasive carcinomas are present and differ in histologic type, grade, or the expression of ER, PgR, or HER2, this information should be included as text in this section.