**Protocol for the Examination of Biopsy Specimens From Patients With Wilms and Other Pediatric Renal Tumors**

**Version:** 5.0.0.0

**Protocol Posting Date:** September 2023

**CAP Laboratory Accreditation Program Protocol Required Use Date:** June 2024

The changes included in this current protocol version affect accreditation requirements. The new deadline for implementing this protocol version is reflected in the above accreditation date.

**For accreditation purposes, this protocol should be used for the following procedures AND tumor types:**

|  |  |
| --- | --- |
| **Procedure** | **Description** |
| Biopsy | Includes specimens designated core biopsy, incisional biopsy, or other |
| **Tumor Type** | **Description** |
| Wilms tumors | Includes pediatric patients with Wilms and other renal tumors  |

**The following should NOT be reported using this protocol:**

|  |
| --- |
| **Procedure** |
| Resection (consider Wilms Tumor Resection protocol) |
| **Tumor Type** |
| Renal cell carcinoma (consider the Kidney protocol) |
| Lymphoma (consider the Hodgkin or non-Hodgkin Lymphoma protocols) |

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**Accreditation Requirements**

This protocol can be utilized for a variety of procedures and tumor types for clinical care purposes. For accreditation purposes, only the definitive primary cancer resection specimen is required to have the core and conditional data elements reported in a synoptic format.

* Core data elements are required in reports to adequately describe appropriate malignancies. For accreditation purposes, essential data elements must be reported in all instances, even if the response is “not applicable” or “cannot be determined.”
* Conditional data elements are only required to be reported if applicable as delineated in the protocol. For instance, the total number of lymph nodes examined must be reported, but only if nodes are present in the specimen.
* Optional data elements are identified with “+” and although not required for CAP accreditation purposes, may be considered for reporting as determined by local practice standards.

The use of this protocol is not required for recurrent tumors or for metastatic tumors that are resected at a different time than the primary tumor. Use of this protocol is also not required for pathology reviews performed at a second institution (i.e., secondary consultation, second opinion, or review of outside case at second institution).

**Synoptic Reporting**

All core and conditionally required data elements outlined on the surgical case summary from this cancer protocol must be displayed in synoptic report format. Synoptic format is defined as:

* Data element: followed by its answer (response), outline format without the paired Data element: Response format is NOT considered synoptic.
* The data element should be represented in the report as it is listed in the case summary. The response for any data element may be modified from those listed in the case summary, including “Cannot be determined” if appropriate.
* Each diagnostic parameter pair (Data element: Response) is listed on a separate line or in a tabular format to achieve visual separation. The following exceptions are allowed to be listed on one line:
	+ Anatomic site or specimen, laterality, and procedure
	+ Pathologic Stage Classification (pTNM) elements
	+ Negative margins, as long as all negative margins are specifically enumerated where applicable
* The synoptic portion of the report can appear in the diagnosis section of the pathology report, at the end of the report, or in a separate section, but all Data element: Responses must be listed together in one location

Organizations and pathologists may choose to list the required elements in any order, use additional methods in order to enhance or achieve visual separation, or add optional items within the synoptic report. The report may have required elements in a summary format elsewhere in the report IN ADDITION TO but not as replacement for the synoptic report i.e., all required elements must be in the synoptic portion of the report in the format defined above.

**Summary of Changes**

**v 5.0.0.0**

* Protocol updated for accreditation requirement

**Reporting Template**

**Protocol Posting Date: September 2023**

**Select a single response unless otherwise indicated.**

**CASE SUMMARY: (KIDNEY, PEDIATRIC RENAL TUMORS: Biopsy)**

*For bilateral tumors, complete a separate checklist for each kidney.*

**EXPERT CONSULTATION**

**+Expert Consultation (Note** [**A**](#N11237)**)**

\_\_\_ Pending - Completion of this CAP Cancer Protocol is awaiting expert consultation

\_\_\_ Completed - This CAP Cancer Protocol or some elements have been performed following expert

 consultation

\_\_\_ Not applicable

**SPECIMEN**

**Procedure (Note** [**B**](#N11236)**)**

\_\_\_ Core biopsy

\_\_\_ Incisional biopsy

\_\_\_ Other (specify): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

\_\_\_ Not specified

**Specimen Laterality  (select all that apply)**

\_\_\_ Right

\_\_\_ Left

\_\_\_ Not specified

**TUMOR**

**Histologic Type (Note** [**C**](#N11240)**)**

\_\_\_ Wilms tumor, favorable histology

\_\_\_ Wilms tumor, diffuse anaplasia

\_\_\_ Congenital mesoblastic nephroma (cellular, classic, or mixed)

\_\_\_ Clear cell sarcoma of kidney

\_\_\_ Rhabdoid tumor

\_\_\_ Other (specify): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

\_\_\_ Malignant neoplasm, type cannot be determined (explain): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

**+Histologic Type Comment: \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_**

**+Relevant Immunohistochemistry (Note** [**C**](#N11240)**)**

\_\_\_ Not performed

\_\_\_ Specify findings: \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

\_\_\_ Pending

**+Ancillary Studies (Note** [**D**](#N11238)**) (select all that apply)**

\_\_\_ Microarray

\_\_\_ Specify findings: \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

\_\_\_ Pending

\_\_\_ FISH

\_\_\_ Specify probe and findings: \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

\_\_\_ Pending

\_\_\_ Next generation sequencing (NGS)

\_\_\_ Specify findings: \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

\_\_\_ Pending

\_\_\_ Other (specify): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

**ADDITIONAL FINDINGS**

**+Additional Findings (specify) (Notes** [**D**](#N11238)**,**[**E**](#N11239)**): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_**

**COMMENTS**

**Comment(s): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_**

**Explanatory Notes**

**A. Expert Consultation**

Expert consultation is not required. This question has been added to annotate, if so desired, that the case has been sent out for consultation and thus items of the CAP protocol could not be completed pending expert consultation. Completion of the CAP protocol will then be performed following consultation.

**B. Frozen Section and Biopsy Handling**

With pediatric renal tumors, there are many issues that can interfere with making accurate diagnostic and staging decisions. The following guidelines are recommended to ensure that the necessary diagnostic features are preserved and properly examined.[1](#R48248)

Because of the high number of false-positives, intraoperative frozen sections should be avoided unless the operative procedure will be altered by the result. Biopsies of pediatric renal tumors present significant potential for diagnostic error, even on permanent section. However, frozen sections from the bivalved nephrectomy specimen—to ensure tumor viability or to prompt other differential diagnostic studies—may be of value. See resection template notes for more detail.

For future potential molecular studies, viable tumor (1 gram or more) should be snap-frozen (liquid nitrogen or cold isopentane) in 2 or more vials, along with a separate portion of nonneoplastic kidney (at least 1 vial).[1](#R48248) The latter serves as a useful control in molecular genetic studies and helps determine whether any detected genomic abnormalities are germline or intratumoral mutations. Nephrogenic rests may also be sampled and frozen for the same reasons.

References

1. Knezevich SR, Garnett MJ, Pysher TJ, et al. ETV6-NTRK3 gene fusion and trisomy 11 establish a histogenetic link between mesoblastic nephroma and congenital fibrosarcoma. Cancer Res. 1998;58(22):5046-5048.

**C. Microscopic Examination: Histology and Immunohistochemistry**

Favorable Histology Wilms Tumor

Classic Wilms tumors present with a mixture of blastemal, stromal, and epithelial cell types. A common difficulty faced by pathologists interpreting a pediatric renal mass is the distinction between a hyperplastic perilobar nephrogenic rest and a Wilms tumor because these may be cytologically identical. The most helpful histologic feature is the absence of a peritumoral fibrous capsule in perilobar nephrogenic rests.

Many other neoplasms may have a histologic appearance similar to blastemal-predominant Wilms tumors. The most common tumors misdiagnosed as Wilms tumors are undifferentiated neuroblastoma, Ewing sarcoma, and synovial sarcoma. The most helpful feature that favors the diagnosis of Wilms tumor is the presence of overlapping nuclei with finely dispersed chromatin. Similarly, epithelial-prominent Wilms tumors show considerable histologic overlap with papillary renal cell carcinoma and metanephric adenoma. A more detailed diagnosis of pediatric renal tumors is provided elsewhere.[1,](#R48280)[2,](#R48281)[3](#R48282)

Immunohistochemistry

For diagnosis of Wilms tumor, no single or panel markers can with 100% confidence either prove or exclude the diagnosis of Wilms tumor. WT1 is commonly positive in blastemal and epithelial elements but may be negative in up to 20% of Wilms tumors. PAX8/PAX2 are expressed in Wilms tumor and this expression may exclude small blue cell tumors of the soft tissue that happen to present in the kidney.[3](#R48282) Almost any other immunohistochemical marker may be found in Wilms tumors in the correct pathologic context.

Although some studies suggest that p53 immunostaining may be a more sensitive predictor of poor outcomes than histologic assessment of anaplasia[1](#R48280), such studies are fraught with difficulties in interpreting the outside limits of “positivity” as well as with interinstitutional variability in immunostaining techniques. Furthermore, some p53 mutations by their nature do not result in abnormal protein accumulation. However, strong, unequivocal p53 protein accumulation identified in a tumor that is suspicious for anaplasia may contribute to the diagnosis.[2](#R48281)

Anaplastic Wilms Tumor

Once a tumor has been diagnosed as Wilms tumor, it is necessary to determine if it is of favorable histology or if anaplasia is present. Although anaplasia is present in only 5% of all cases[4,](#R48283)[5](#R48284), it is the major prognostic indicator and will place a tumor in an unfavorable histologic category.

The presence of anaplasia is a significant prognostic factor in Wilms tumor and places the tumor in an unfavorable category. Although the mechanism for unfavorable prognosis is unclear, anaplasia may be a marker of chemotherapy resistance. A diagnosis of anaplasia requires both (1) gigantic polypoid nuclei with increased chromatin content and major diameters at least 3 times those of adjacent cells and (2) the presence of multipolar or otherwise recognizable polypoid mitotic figures. On a small biopsy, a single multipolar mitotic figure or an unequivocally gigantic tumor cell nucleus may be sufficient criteria for diagnosis. Severe nuclear unrest is defined as nuclear pleomorphism or atypia approaching the criteria of anaplasia. Care should be taken in the assessment of anaplasia cells exhibiting rhabdomyoblastic differentiation, as these cells may show nuclear enlargement, pleomorphism, and hyperchromasia akin to regenerating skeletal muscle. Such areas of “pseudoanaplasia” will have increased cytoplasmic volume and will lack atypical mitoses, as described above.

Criteria for focal versus diffuse anaplasia have been defined topographically and are rigorous.[5](#R48284) This topographic definition of focal anaplasia makes it mandatory that pathologists carefully document the exact site from which every section is obtained (e.g., on a diagram, specimen photocopy, and/or photograph of the gross specimen).

Focal Anaplasia

Diagnosis of focal anaplasia is warranted if all of the following are true:

* No anaplasia should be present in tumors within renal vessels or outside the kidney.
* Anaplasia must be confined to 1 or 2 sharply localized regions, each less than 15 mm in diameter, within the primary intrarenal tumor site.[6](#R48285)
* Each focus of anaplasia must be surrounded on all sides by nonanaplastic tissue. This may require mapping of the tumor during submission.
* The remaining nonanaplastic tumor must not show severe nuclear unrest.

(The same criteria apply to posttreatment nephrectomies. There is no evidence to suggest that either chemotherapy or radiation therapy results in anaplasia.)

Diffuse Anaplasia

Diagnosis of diffuse anaplasia is warranted if any of the following are true:

* Anaplasia is present in tumors in any extrarenal site, including vessels of the renal sinus, extracapsular infiltrates, or nodal or distant metastases. Also, anaplasia is present in intrarenal vascular involvement by tumor.
* Anaplasia is present in a random biopsy.
* Anaplasia is unequivocally identified, but the tumor fails any of the above criteria for focal anaplasia.

Posttherapy Classification of Wilms Tumor:

The response of a Wilms tumor to prior therapy may help guide the subsequent therapeutic strategy. For this reason, the Children’s Oncology Group is using the overall categories (low, medium, high risk) utilized by the International Society of Paediatric Oncology (SIOP) when categorizing posttherapy tumors.[7](#R48286) As outlined above, these categories are based on the proportion of the tumor that is viable and blastemal, and in COG only apply in favorable histology Wilms tumor. It is acknowledged that such quantitative analysis is quite difficult to reproduce and is highly dependent on how representative of the entire tumor the sections submitted are. The overall concept is that tumor that remains highly undifferentiated and proliferative following therapy will require more aggressive therapy going forward. Pathologists should, as always, use their best judgment. Staging of posttherapy nephrectomy specimens should be based on the resection specimen only. A prior pretherapy biopsy is not a criterion for assigning stage III to a post-therapy specimen.

Impact of Necrotic Tumor on Wilms Tumor Staging:

Necrosis outside of the kidney may be present in pretherapy resection specimens or posttherapy specimens. The presence of necrotic tumor or chemotherapy-induced change (in the absence of viable tumor) in the renal sinus and/or within the perirenal fat is not regarded as a reason for upstaging, providing the tumor (either viable or necrotic) is completely excised and does not reach the resection margins. In contrast, the presence of necrotic tumor or chemotherapy-induced changes in a lymph node or at the resection margins is regarded as proof of previous tumor with potential microscopic residual disease, and therefore the tumor is assigned stage III.

Congenital Mesoblastic Nephroma

Congenital Mesoblastic Nephroma (CMN), a tumor of infancy, represents 2 morphologically/genetically distinct tumors: classic and cellular forms. The classic subtype is characterized by a whirled gross appearance and is composed of bland spindle cells with a low mitotic rate that are arranged in long, sweeping fascicles.[8,](#R48287)[9](#R48288) The cellular subtype has a fleshy and hemorrhagic gross appearance and is characterized by more densely cellular plump spindle cells with shorter to haphazard fascicles and a higher mitotic rate. Cases may also show a “mixed” histologic appearance with features of both classic and cellular CMN.[10,](#R48289)[11](#R48290) Cellular CMN may be positive for PAX8 and desmin and negative for CD34 and cytokeratins, but immunohistochemical markers are overall nonspecific.[11,](#R48290)[12](#R48291)

Approximately 10% of CMNs recur. Virtually all CMNs that recur are of the cellular subtype.[8,](#R48287)[9,](#R48288)[13](#R48292) Recurrences occur very rapidly, often within the first month of diagnosis. Virtually all recurrences occur by the first year of age.[14](#R48293) More than half are local recurrences; however, pulmonary metastases have been identified in 20% of patients who relapse. However, the primary determinant of outcome is the completeness of the excision. Surgeons should be educated and encouraged to secure wide margins, particularly medial margins, when resecting renal tumors in infants. Nonetheless, one can rarely be sure that the medial margin is clear; therefore, all patients should be followed closely. Monthly abdominal ultrasounds should be performed for 1 year, with the hope of catching recurrences early enough to surgically excise them. Adjuvant chemotherapy is required when there is gross residual tumor.[13,](#R48292)[14](#R48293) Radiation has no demonstrable effect.

Clear Cell Sarcoma of the Kidney

Clear cell sarcoma of the kidney (CCSK) is capable of mimicking, or being mimicked by, every other major neoplastic entity in the pediatric kidney. Immunohistochemical stains for CCSK are non-specific but may be helpful to aid the diagnosis. CCSK may show variable but consistent expression for BCOR cyclin D1, and NGFR; other IHC may be useful to exclude other diagnoses.[15,](#R48294)[16,](#R48295)[17](#R48296) Molecular features are described in Note G.

The histologic spectrum and clinical outcome of patients with CCSK have been reported by the National Wilms Tumor Study Group.[18](#R48297) Nearly all patients with stage I CCSK survive. Conversely, patients with more advanced disease have a propensity for local recurrence and metastasis. Recurrences can occur from years to decades after the initial presentation, sometimes demonstrating bland histology that differs from the primary tumor. The metastatic pattern tends to be more widespread than that of Wilms tumor and includes bone, brain, and soft tissue. There is a high recurrence rate and death rate even when treated by combination chemotherapy, but survival can be greatly improved after treatment with doxorubicin, 11 which underscores the importance of identifying this neoplasm to facilitate early administration of more effective chemotherapy regimes.

There are several variants of CCSK, among which the following are most important:

Classic Pattern

The classic pattern of CCSK presents an evenly dispersed network of fine, arborizing vessels accompanied by a variable amount of spindle-cell stroma, subdividing the tumor into nests or cords of regular size, usually about 8 to 12 cells in width. The tumor cells are of regular size, usually with stellate cytoplasm, which often surrounds clear vacuoles. The nuclei are notably regular in size, with finely dispersed chromatin, and usually inconspicuous nucleoli. Mitotic activity may be sparse. Scattered preexisting tubules or glomeruli often are dispersed through the peripheral regions of the tumor. This pattern of growth, which isolates and separates individual nephronic units or collecting tubules, is an important clue that one is not dealing with Wilms tumor. The latter almost always has a sharply defined, “pushing” border.

Hyalinizing Pattern

The hyalinizing pattern of CCSK often has an osteoid-like, nonbirefringent matrix that separates tumor cells, giving an appearance reminiscent of osteosarcoma. A similar change may be seen in rhabdoid tumor of the kidney (RTK).

Epithelioid Pattern

The epithelioid pattern is the most deceptive of the patterns of CCSK, in which the tumor cells align themselves along vessels in a manner mimicking the tubules of Wilms tumor. Often these cells form filigree-like strands.

Rhabdoid Tumor of the Kidney

This distinctive renal neoplasm most commonly is encountered in infants younger than 1 year of age and is uncommon in patients older than 5 years.[19,](#R48298)[20,](#R48299)[21](#R48300) It is extremely aggressive and is the prognostically unfavorable neoplasm of the kidney in early life. Rhabdoid tumors continue to present significant diagnostic challenges, particularly when they do not show overt rhabdoid features. However, the growing appreciation that this tumor arises in sites other than the kidney and the central nervous system, and the increased appreciation of the wide histologic spectrum of rhabdoid tumors, have contributed to a marked increase in their correct diagnosis. Rhabdoid tumors of the kidney should not be confused with the true myogenic cells, which are often found in Wilms tumors.

The most distinctive features of rhabdoid tumor of the kidney (RTK) are rather large cells with large vesicular nuclei, a prominent single nucleolus, and the presence in at least some cells of globular eosinophilic cytoplasmic inclusions composed of whorled masses of intermediate filaments. Another distinctive feature is the extremely aggressive, invasive pattern of this lesion. RTK has a diverse immunohistochemical profile. Tumors may be positive for many supposedly incompatible epitopes for epithelial (keratins), myogenous, neural, and mesenchymal cell types. Epithelial membrane antigen (EMA) should be included in the routine panel applied to small blue cell tumors, largely because of the typical focal strong positivity for EMA (as well as a multiple of other markers) that rhabdoid tumors demonstrate.[2](#R48281) RTKS consistently show loss of expression of INI-1/BAF47 (see Note G).[2](#R48281)

References

1. Knezevich SR, Garnett MJ, Pysher TJ, et al. ETV6-NTRK3 gene fusion and trisomy 11 establish a histogenetic link between mesoblastic nephroma and congenital fibrosarcoma. Cancer Res. 1998;58(22):5046-5048.
2. Hoot AC, Russo P, Judkins AR, Perlman EJ, Biegel JA. Immunochemical analysis of hSNF5/INI1 distinguishes renal and extra-renal malignant rhabdoid tumors from other pediatric soft tissue tumors. Am J Surg Pathol. 2004;28(11):1485-1491.
3. Murphy WM, Perlman EJ, Grignon D. Tumors of the kidney, bladder, and related urinary structures. Atlas of Tumor Pathology. 4th Series. Washington DC: Armed Forces Institute of Pathology, 2004.
4. Zuppan CW. Handling and evaluation of pediatric renal tumors. Am J Clin Pathol. 1998;109(4 suppl 1): S31-S37.
5. Faria P, Beckwith JB, Mirshra K, et al. Focal versus diffuse anaplasia in Wilms tumor-new definitions with prognostic significance: a report from the National Wilms Tumor Study Group. Am J Surg Pathol. 1996;20(8):909-920.
6. Vujanic GM, Gessler M, Ooms AHAG, Collini P, Coulomb-l’Hermine A, D’Hooghe E, de Krijger RR, Perotti D, Pritchard-Jones K, Vokuhl C, van den Heuvel-Eibrink MM, Graf N; International Society of Paediatric Oncology-Renal Tumour Study Group (SIOP-RTSG). The UMBRELLA SIOP-RTSG 2016 Wilms tumor pathology and molecular biology protocol. Nat Rev Urol. 2018 Nov;15(11):693-701. doi: 10.1038/s41585-081-0100-3. Erratum in: Nat Rev Urol. 2019 Sep;16(9):563. PMID:30310143.
7. Vujanic GM, Sandstedt B. The pathology of Wilms’ tumor (nephroblastoma): the International Society of Paediatric Oncology Approach. J Clin Pathol. 2010;63:102-109.
8. Gooskens SL, Houwing ME, Vujanic GM. Congenital mesoblastic nephroma 50 years after its recognition: a narrative review. Pediatr Blood Cancer. 2017;64(7).
9. Vokuhl C, Nourkami-Tutdibi N, Furtwangler R, et al. ETV6-NTRK3 in congenital mesoblastic nephroma: A report of the SIOP/GPOH nephroblastoma study. Pediatr Blood Cancer. 2018;65(4).
10. Bolande RP. Congenital mesoblastic nephroma of infancy. Perspect Pediatr Pathol. 1973;1:227-50.
11. Pettinato G., Manivel JC, Wick MR, et al. Classical and cellular (atypical) congenital mesoblastic nephroma: a clinicopathologic, ultrastructural, immunohistochemical, and flow cytometric study. Hum Pathol. 1989;20(7):682-90.
12. Demellawy DE, Cundiff CA, Nasr A, et al. Congenital mesoblastic nephroma: a study of 19 cases using immunohistochemistry and ETV6-NTRK3 fusion gene rearrangement. Pathology. 2016;48(1)47-50.
13. Furtwaengler R, Reinhard H, Leuscher I, et al. Mesobalstic nephroma – a report from the Gesellschaft fur Padiatrische Onkologie und Hamatologie (GPOH). Cancer. 2006;106(10):2275-83.
14. Jehangir S, Kurian JJ, Selvarajah D. Recurrent and metastatic congenital mesoblastic nephroma: where does the evidence stand? Peiatr Surg Int. 2017;33:1183-1188.
15. Mirkovic J, Calicchio M, Fletcher CD, Perez-Atayde AR. Diffuse and strong cyclin D1 immunoreactivity in clear cell sarcoma of the kidney. Histopathology. 2015;67:306-12.
16. Roy A, Kumar V, Zorman B, et al. Recurrent internal tandem duplications of BCOR in clear cell sarcoma of the kidney. Nat Commun. 2015:6:8891.
17. Arva NC, Bonadio J, Perlman EJ, Cajaiba MM. Diagnostic utility of Pax8, Pax2, and NGFR immunohistochemical expression in pediatric renal tumors. Appl Immunohistochem Mol Morphol. 2017. doi: 10.1097 PMID: 28426529.
18. Argani P, Perlman EJ, Breslow NE, et al. Clear cell sarcoma of the kidney: a review of 351 cases from the National Wilms Tumor Study Group Pathology Center. Am J Surg Pathol. 2000;24(1):4-18.
19. Weeks DA, Beckwith JB, Mierau GW, wt al. Rhabdoid tumor of kidney. A report of 111 cases from the National Wilms’ Tumor Study Pathology Center. Am J Surg Pathol. 1989;13(6):439-58.
20. Thomlinson GE, Breslow NE, Dome J, et al. Rhabdoid tumor of the kidney in National Wilms’ Tumor Study: age at diagnosis as a prognostic factor. J Clin Oncol. 2005-;23:7641-7645.
21. van den Heuvel-Eibrink MM, Grundy P, Graf N, et al. Characteristics and survival of 750 children diagnosed with a renal tumor in the first seven months of life: A collaborative study by the SIOP/GPOH/SFOP, NWTSG, and UKCCSG Wilms tumor study groups. Pediatr Blood Cancer. 2008;50(6):1130-4.

**D. Ancillary Studies**

The diagnosis of primary renal tumors in children remains largely based on examination of hematoxylin-eosin (H&E)-stained sections. However, a few ancillary studies may be employed for diagnostic or prognostic importance.

**Wilms tumor molecular testing:**

Molecular tests such as loss of heterozygosity (LOH) at chromosomes 1p and 16q, 1q gain, and 11p15 loss have prognostic significance in certain patient populations. Augmentation of therapy has been shown to be effective for WT with combined LOH at 1p and 16q, therefore analysis of these loci, most commonly by targeted or genome-wide SNP array, has become routine practice in North America**.**[1,](#R48260)[2,](#R48261)[3](#R48262)  While 1q gain is associated with adverse prognosis, the benefit of increased therapy is an area of active investigation. LOH and loss of imprinting of 11p15 have been associated with increased risk of relapse in young patients with stage I favorable histology WT that is treated with nephrectomy alone without adjuvant therapy.[4,](#R48263)[5,](#R48264)[6](#R48265)

The molecular etiology of Wilms tumor is heterogeneous and more than a dozen genes have been found to be recurrently mutated in Wilms tumor tissue including genes involved in transcriptional regulation (WT1, MYCN, SIX1, SIX2, MLLT1), microRNA processing (DGCR8, DROSHA, DICER1, and XPO5), and the WNT signaling pathway (AMER1 and CTNNB1).  TP53 mutations have been detected in 50-75% of anaplastic histology Wilms tumors. Additionally, approximately 70% of Wilms tumors have evidence of IGF2 overexpression, which may arise via genetic or epigenetic changes at chromosome locus 11p15.[7](#R48266) Some of these genes may also have germline mutations, which has implications for Wilms tumor predisposition and genetic counseling.

**Other tumor molecular testing:**

Congenital Mesoblastic Nephroma

CMN represents 2 genetically distinct tumors that correspond to the histologic subtypes. “Classic” CMN (24% of cases), which histologically resembles a type of fibromatosis has recently been recognized to harbor aEGFR activating mutations (most often internal tandem duplications).[8,](#R48267)[9](#R48268) These alterations may be detected by next generation sequencing (NGS). “Cellular” CMN (66% of cases), which is analogous to the soft tissue tumor, infantile fibrosarcoma, most commonly contains an ETV6-NTRK3 gene fusion. However, a variety of other variant MAP kinase pathway activating translocations or mutations may also be present.[10](#R48269) ETV6-NTRK3 fusions may be detected by FISH or NGS, while the less common alternative alterations may be detected by comprehensive NGS for mutations and fusions. Genetically, “mixed” CMN have most frequently demonstrated EGFR alterations similar to the classic subtype, with rare cases with genetic overlap to cellular CMN.[9](#R48268)

Clear Cell Sarcoma of the Kidney

CCSK is characterized molecularly by BCOR internal tandem duplications or YWHAE-NUTM2B fusions.[11](#R48270) The YWHAE-NUTM2B fusion was the first reported recurrent alteration in CCSK, but only accounts for approximately 15% of cases.[12](#R48271) With increased use of NGS, the presence of the BCOR internal tandem duplication was confirmed in the majority of tumors that are negative for the fusion.[11,](#R48270)[13](#R48272)  Rare renal tumors with a CCSK morphology have also been detected with BCOR gene fusions.[13,](#R48272)[14,](#R48273)[15](#R48274) Fusions may be detected by FISH or by NGS-based RNA sequencing. The BCOR internal tandem duplication may be detected by NGS or by targeted PCR assays.

Rhabdoid Tumor of the Kidney

Both renal and extrarenal rhabdoid tumors carry homozygous deletions and/or mutations of the SMARCB1 gene located at 22q11.2, which is a member of the SWI/SNF chromatin remodeling complex.[16](#R48275) Furthermore, germline mutations have been identified in individuals with both renal and central nervous system rhabdoid tumors. The SMARCB1 (INI1) gene causes conformational changes in the nucleosome, thereby altering histone-DNA binding and facilitating transcription factor access. Mutations in SMARCB1 correspond to the loss of expression by immunohistochemistry using the INI-1/BAF47 antibody.[17](#R48276) This antibody shows strong nuclear expression in normal tissues; however, nuclear expression is lost (aberrant expression) in rhabdoid tumor nuclei. Additionally, a variety of other tumors may also show loss of INI-1 by immunohistochemistry to include renal medullary carcinoma, epithelioid sarcoma, among several others. Molecular testing for SMARCB1 mutations is not necessary for the diagnosis of RTK, but may be utilized in the workup, particularly to identify germline mutations.

References

1. Grundy PE, Breslow NE, Perlman E, et al. The National Wilms Tumor Study Group. Loss of heterozygosity for chromosomes 1p and 16q is an adverse prognostic factor in favorable-histology Wilms tumor: a report from the National Wilms Tumor Study Group. J Clin Oncol. 2005;23:7312-7321.
2. Dome JS, Mullen EA, Dix DB, Gratias EJ, Ehrlich PF, Daw NC, Geller JI, Chintagumpala M, Khanna G, Kalapurakal JA, Renfro LA, Perlman EJ, Grundy PE, Fernandez CV. Impact of the First Generation of Children's Oncology Group Clinical Trials on Clinical Practice for Wilms Tumor. J Natl Compr Canc Netw. 2021 Aug 1;19(8):978-985.
3. Dix DB, Fernandez CV, Chi YY, Mullen EA, Geller JI, Gratias EJ, Khanna G, Kalapurakal JA, Perlman EJ, Seibel NL, Ehrlich PF, Malogolowkin M, Anderson J, Gastier-Foster J, Shamberger RC, Kim Y, Grundy PE, Dome JS; AREN0532 and AREN0533 study committees. Augmentation of Therapy for Combined Loss of Heterozygosity 1p and 16q in Favorable Histology Wilms Tumor: A Children's Oncology Group AREN0532 and AREN0533 Study Report. J Clin Oncol. 2019 Oct 20;37(30):2769-2777.
4. Gratias EJ, Dome JS, Jennings LJ, et al. Association of chromosome 1q gain with inferior survival in favorable histology Wilms tumor. J Clin Oncol. 2016;34(26):3189-3194.
5. Perlman EJ, Grundy P, Anderson JR, et al. WT1 mutation and 11p loss of heterozygosity predict relapse in very low risk Wilms tumors treated by surgery alone. J Clin Oncol. 2011;29:698-703.
6. Fernandez CV, Perlman EJ, Mullen EA, Chi YY, Hamilton TE, Gow KW, Ferrer FA, Barnhart DC, Ehrlich PF, Khanna G, Kalapurakal JA, Bocking T, Huff V, Tian J, Geller JI, Grundy PE, Anderson JR, Dome JS, Shamberger RC. Clinical Outcome and Biological Predictors of Relapse After Nephrectomy Only for Very Low-risk Wilms Tumor: A Report From Children's Oncology Group AREN0532. Ann Surg. 2017 Apr;265(4):835-840.
7. Gadd S, Huff V, Walz AL, Ooms AHAG, Armstrong AE, Gerhard DS, Smith MA, Auvil JMG, Meerzaman D, Chen QR, Hsu CH, Yan C, Nguyen C, Hu Y, Hermida LC, Davidsen T, Gesuwan P, Ma Y, Zong Z, Mungall AJ, Moore RA, Marra MA, Dome JS, Mullighan CG, Ma J, Wheeler DA, Hampton OA, Ross N, Gastier-Foster JM, Arold ST, Perlman EJ. A Children's Oncology Group and TARGET initiative exploring the genetic landscape of Wilms tumor. Nat Genet. 2017 Oct;49(10):1487-1494.
8. Lei L, Stohr BA, Berry S, Lockwood CM, Davis JL, Rudzinski ER, Kunder CA. Recurrent EGFR alterations in NTRK3 fusion negative congenital mesoblastic nephroma. Pract Lab Med. 2020 May 16;21:e00164.
9. Wegert J, Vokuhl C, Collord G, et al. Recurrent intragenic rearrangements of EGFR and BRAF in soft tissue tumors of infants. Nat Commun. 2018;9(1):2378.
10. Knezevich SR, Garnett MJ, Pysher TJ, et al. ETV6-NTRK3 gene fusion and trisomy 11 establish a histogenetic link between mesoblastic nephroma and congenital fibrosarcoma. Cancer Res. 1998;58(22):5046-5048.
11. Ueno-Yokohata H, Okita H, Nakasato K, et al. Consistent in-frame internal tandem duplications of BCOR characterize clear cell sarcoma of the kidney. Nat Genet. 2015;47(8):861-863.
12. O’Meara E, Stack D, Lee CH, et al.  Characterization of the chromosomal translocation t(10;17)(q22;p13) in clear cell sarcoma of kidney. J Pathol. 2012;227:72-80.
13. Roy A, Kumar V, Zorman B, et al.  Recurrent internal tandem duplications of BCOR in clear cell sarcoma of the kidney. Nat Commun. 2015;6:8891.
14. Wong MK, NgCCY, Kuick CH.  Clear cell sarcomas of the kidney are characterized by BCOR gene abnormalities, including exon 15 internal tandem duplications and BCOR-CCNB3 gene fusion. Histopathology. 2018;72(2):320-329.
15. Han H, Betrannd KC, Patel KR, et al.  BCOR-CCNB3 fusion-positive clear cell sarcoma of the kidney. Pediatr Blood Cancer. 2019;26:e28151.
16. Biegel JA, Zhou J-Y, Rorke LB, et al. Germline and acquired mutations of INI1 in atypical teratoid and rhabdoid tumors. Cancer Res.1999;59(1):74-79.
17. Hoot AC, Russo P, Judkins AR, Perlman EJ, Biegel JA. Immunohistochemical analysis of hSNF5/INI1 distinguishes renal and extra-renal malignant rhabdoid tumors from other pediatric soft tissue tumors. Am J Surg Pathol. 2004;28(11):1485-1491.

**E. Syndromes Associated with Wilms Tumor**

The following syndromes are associated with Wilms tumor[1,](#R48277)[2,](#R48278)[3](#R48279):

* REST-related Wilms tumor
* TRIM28-related Wilms tumor
* WT1 disorder
* 1p15-related Wilms tumor (Beckwith-Wiedemann syndrome, hemi-hyperplasia)
* WAGR syndrome
* Perlman familial nephroblastomatosis syndrome
* Denys-Drash syndrome
* Trisomy 18
* Neurofibromatosis
* Bloom syndrome
* etc.

References

1. Beckwith JB. Nephrogenic rests and the pathogenesis of Wilms tumor: developmental and clinical considerations. Am J Med Genet. 1998;79(4):268-273.
2. Charles AK, Brown KW, Berry PJ. Microdissecting the genetic events in nephrogenic rests and Wilms tumor development. Am J Pathol. 1998;153(3):991-1000.
3. Turner JT, Brzezinski J, Dome JS. Wilms Tumor Predisposition. 2003 Dec 19