Protocol for the Examination of Resection Specimens From Pediatric Patients With Ewing Sarcoma

Version: 4.1.1.1
Protocol Posting Date: September 2022

The use of this protocol is recommended for clinical care purposes but is not required for accreditation purposes.

This protocol should be used for the following procedures AND tumor types:

<table>
<thead>
<tr>
<th>Procedure</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>Resection</td>
<td>Includes specimens designated resection, amputation, limb salvage procedure, or other</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Tumor Type</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ewing sarcoma</td>
<td>Includes pediatric patients with osseous and extraosseous Ewing sarcoma family of tumors</td>
</tr>
</tbody>
</table>

The following should NOT be reported using this protocol:

<table>
<thead>
<tr>
<th>Procedure</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>Needle, incisional or skin biopsies (consider Pediatric Ewing Sarcoma Biopsy protocol)</td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Tumor Type</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>Adult Ewing sarcoma*</td>
<td>(consider using Bone or Soft Tissue protocols)</td>
</tr>
<tr>
<td>Round cell sarcoma with EWSR1-non-ETS fusions, CIC-rearranged sarcoma, or sarcoma with BCOR-genetic alterations (consider using Bone or Soft Tissue protocols)</td>
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</tbody>
</table>

*Ewing sarcoma in adults may be treated differently than pediatric Ewing sarcoma and use of the AJCC TNM staging system remains appropriate for adult patients.

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With guidance from the CAP Cancer and CAP Pathology Electronic Reporting Committees.
* Denotes primary author.

Accreditation Requirements
The use of this case summary is recommended for clinical care purposes but is not required for accreditation purposes. The core and conditional data elements are routinely reported. Non-core data elements are indicated with a plus sign (+) to allow for reporting information that may be of clinical value.
Summary of Changes
v 4.1.1.1

- Corrected spelling error and format in cover page
- The ‘Expert Consultation’ question was made optional
- Added Explanatory ‘Note A’ and associated with the ‘Expert Consultation’ question
Reporting Template  
Protocol Posting Date: September 2022  
Select a single response unless otherwise indicated.

CASE SUMMARY: (EWING SARCOMA: Resection)

EXPERT CONSULTATION

+Expert Consultation (Note A)  
___ Pending - Completion of this CAP Cancer Protocol is awaiting expert consultation  
___ Completed - This CAP Cancer Protocol or some elements have been performed following expert consultation  
___ Not applicable

CLINICAL

Preresection Treatment (select all that apply)  
___ No known preresection therapy  
___ Chemotherapy performed  
___ Radiation therapy performed  
___ Therapy performed, type not specified  
___ Not specified

SPECIMEN (Note B)

Procedure (Note C)  
___ Resection  
___ Amputation (specify type): _________________  
___ Limb salvage procedure (specify type): _________________  
___ Other (specify): _________________  
___ Not specified

TUMOR

Multiple Primary Sites  
___ Not applicable  
___ Present: ____________________  
Please complete a separate checklist for each primary site

Tumor Site  
___ Osseous  
___ Long bones of upper limb, scapula and associated joints (specify): _________________  
___ Short bones of upper limb and associated joints (specify): _________________  
___ Long bones of lower limb and associated joints (specify): _________________  
___ Short bones of lower limb and associated joints (specify): _________________  
___ Overlapping lesion of bones, joints and articular cartilage of limbs (specify): _________________  
___ Bone of limb, NOS (specify): _________________
___ Bones of skull and face and associated joints (excluding mandible C41.1) (specify):

___ Mandible (specify): ____________________________

___ Vertebral column (excluding sacrum and coccyx C41.4) (specify): ______________________________

___ Rib, sternum, clavicle and associated joints (specify): ________________________________

___ Pelvic bones, sacrum, coccyx and associated joints (specify): ______________________________

___ Overlapping lesion of bones, joints and articular cartilage (specify): ______________________________

___ Bone, NOS: ______________________________

___ Extraosseous

___ Heart / mediastinum

___ Heart (specify): ______________________________

___ Anterior mediastinum (specify): ______________________________

___ Posterior mediastinum (specify): ______________________________

___ Mediastinum, NOS: ______________________________

___ Overlapping lesion of heart, mediastinum and pleura (specify): ______________________________

___ Peritoneum and / or retroperitoneum

___ Retroperitoneum: ______________________________

___ Peritoneum, including omentum and mesentery (specify parts): ______________________________

___ Peritoneum, NOS: ______________________________

___ Peritoneum and / or retroperitoneum

___ Other soft tissue

___ Head, face, and neck (specify): ______________________________

___ Upper limb and shoulder (specify): ______________________________

___ Lower limb and hip (specify): ______________________________

___ Thorax (specify): ______________________________

___ Abdomen (specify): ______________________________

___ Pelvis (specify): ______________________________

___ Trunk (specify): ______________________________

___ Overlapping lesion (specify): ______________________________

___ Other, NOS: ______________________________

___ Not specified

Tumor Size (Note C)

___ Greatest dimension in Centimeters (cm): ________________ cm

+Additional Dimension in Centimeters (cm): _____ x _____ cm

___ Cannot be determined (explain): ______________________________

Site(s) Involved by Direct Tumor Extension (select all that apply)

___ Epiphysis or apophysis

___ Metaphysis

___ Diaphysis

___ Cortex

___ Medullary cavity

___ Surface

___ Joint

___ Adjacent soft tissue: ______________________________

___ Other (specify): ______________________________

___ Cannot be determined: ______________________________
Lymphovascular Invasion (Note D)
___ Not identified
___ Present
___ Cannot be determined: ____________________

Treatment Effect (Note E)
*Treatment effect includes necrosis, fibrosis and other treatment related changes.*
___ Not applicable (no preresection therapy)
___ Not identified
___ Present
___ Percentage of Treatment Effect
   ___ Specify percentage: ____________________ %
   ___ Other (specify): ____________________
   ___ Cannot be determined
___ Cannot be determined: ____________________

+Tumor Comment: ____________________

MARGINS (Note F)

Margin Status
___ All margins negative for tumor

Closest Margin(s) to Tumor (select all that apply)
___ Bone (specify): ____________________
___ Soft tissue (specify): ____________________
___ Parenchymal (specify): ____________________
___ Other (specify): ____________________
___ Cannot be determined (explain): ____________________

Distance from Tumor to Closest Margin
*Specify in Centimeters (cm)*
___ Exact distance: ____________________ cm
___ Greater than: ____________________ cm
___ Other (specify): ____________________
___ Cannot be determined: ____________________
___ Tumor present at margin

Margin(s) Involved by Tumor
___ Specify involved margin(s): ____________________
___ Cannot be determined (explain): ____________________
___ Cannot be determined: ____________________

+Margin Comment: ____________________
REGIONAL LYMPH NODES

Regional Lymph Node Status
___ Not applicable (no regional lymph nodes submitted or found)
___ Regional lymph nodes present
    ___ All regional lymph nodes negative for tumor
    ___ Tumor present in regional lymph node(s)

Number of Lymph Nodes with Tumor
    ___ Exact number (specify): _________________
    ___ At least (specify): _________________
    ___ Other (specify): _________________
    ___ Cannot be determined (explain): _________________

Number of Lymph Nodes Examined
    ___ Exact number: _________________
    ___ At least (specify): _________________
    ___ Other (specify): _________________
    ___ Number cannot be determined (explain): _________________

+Regional Lymph Node Comment: _________________

DISTANT METASTASIS

Distant Site(s) Involved, if applicable (select all that apply)
___ Not applicable
___ Lung: _________________
___ Bone: _________________
___ Other (specify): _________________
___ Cannot be determined: _________________

PATHOLOGIC STAGE CLASSIFICATION (pTNM, AJCC 8th Edition) (Note G)
The AJCC staging systems for bone and soft tissue based tumors may be used for pathologic staging if desired.

SPECIAL STUDIES (Note H)
Results of these studies may not be available at the time of the final report

+Immunohistochemistry (specify): _________________

Cytogenetic Findings
___ Not performed
___ Pending
___ EWSR1 rearrangement, fusion partner not known
___ EWSR1-FLI1 gene rearrangement
___ EWSR1-ERG gene rearrangement
___ Other EWSR1 gene rearrangement (specify): _________________
___ Non-EWSR1 variant translocation (specify): _________________
___ Other (specify): _________________
___ No rearrangement identified
___ Not known

**Method for Cytogenetic Studies**
___ Not applicable (Cytogenetic Studies not performed)
___ Conventional karyotyping
___ Fluorescent in situ hybridization (FISH)
___ Reverse transcriptase polymerase chain reaction (RT-PCR)
___ Other (specify): _________________
___ Not known

**ADDITIONAL FINDINGS**

+Additional Findings (specify): _________________

**COMMENTS**

Comment(s): _________________
Explanatory Notes

A. Expert Consultation
Expert consultation is not required. This question has been added to annotate, if so desired, that the case has been sent out for consultation and thus items of the CAP protocol could not be completed pending expert consultation. Completion of the CAP protocol will then be performed following consultation.

B. Tissue Handling
Tissue specimens optimally are received fresh/unfixed because of the importance of ancillary studies, such as cytogenetics and molecular testing, which may prefer fresh tissue. First priority should always be given to formalin-fixed tissues (FFPE) for morphologic evaluation. Ideally, some tissue can be submitted for FFPE without decalcification or following decalcification in EDTA or ETDA+acid decalcification solutions to preserve nucleic acids for molecular testing, to including FISH, RT-PCR, and/or next generation sequencing (NGS). Decalcification in pure acid decalcification solutions degrade nucleic acids and limit molecular testing. Following submission of FFPE, submission of fresh tissue for cytogenetics and/or snap freezing a minimum of 100 mg of viable tumor may be needed potential molecular studies and/or COG study purposes. Molecular testing on formalin-fixed paraffin-embedded tissue may be performed for FISH evaluation of EWSR1 rearrangement, for RT-PCR evaluation of EWSR1-FLI1, EWSR1-ERG, and other ES translocations, or NGS. When the amount of tissue is limited, the pathologist can keep the frozen tissue aliquot used for frozen section (usually done to determine sample adequacy and viability) in a frozen state (-70°C is preferable). Translocations may be detected using RT-PCR on frozen or fixed paraffin-embedded tissue, or FISH on touch preparations made from fresh tissue or formalin-fixed paraffin-embedded tissue.

Note that classification of many subtypes of sarcoma is not always dependent upon special studies, such as cytogenetics or molecular genetics, but frozen tissue may be required to enter patients into treatment protocols. Discretion should be used in triaging tissue from sarcomas. Adequate tissue should be submitted for conventional light microscopy before tissue has been taken for cytogenetics, electron microscopy, or molecular analysis.

References

C. Procedures
Tumor Resection
Resection specimens may be intralesional, marginal, wide, or radical in extent. Intrallesional resections extend through tumor planes, with gross or microscopic residual tumor identifiable at surgical margins. A marginal resection involves a margin formed by reactive tissue surrounding the tumor. A wide radical resection has surgical margins that extend through normal tissue, usually external to the anatomic compartment containing the tumor. For all types of resections, marking (tattoo with ink followed by use of a mordant) and orientation of the specimen (prior to cutting) by the surgeon are highly recommended for accurate pathologic evaluation. Full representative mapping of the specimen is also recommended, as discussed below.

A full sagittal section of a bone tumor resection specimen, as illustrated in Figure 1, allows for mapping of the entire central face of the tumor and adjacent marginal tissue. Sectioning the specimen in a
longitudinal plane that allows for evaluation of the tumor in its greatest cross-sectional dimension is important. Soft tissue and bone marrow margins should be inked and taken prior to sectioning the specimen with both amputation and limb salvage specimens. Freezing of the specimen prior to cutting with a bone saw (with intraosseous specimens) is the preferred method at some institutions. This face of the specimen should be documented using digital imaging photography or alternatively by a photocopy of the specimen when sealed in a plastic bag. As shown in Figure 1 of an amputation specimen with soft tissue in place, the central full face of the specimen and lesional region can be mapped and blocked following fixation and with adequate decalcification for complete microscopic examination, including estimate of percentage of tumor necrosis. If possible, at least one section of tumor without decalcification or decalcification with less harsh decalcification methods to include EDTA or ETDA+formic acid is recommended to preserve integrity of nuclei acids.

Figure 1. Grid diagram of histologic sections taken, superimposed on photograph of a sagittally-sectioned amputation specimen including the distal femur and proximal tibia.

References

D. Lymphovascular Invasion (LVI)
Lymphovascular invasion (LVI) indicates whether microscopic lymphovascular invasion is identified in the pathology report. LVI includes lymphatic invasion, vascular invasion, or lymphovascular invasion.
Evaluation of LVI may require immunohistochemical staining for endothelial markers (CD31, CD34, D240, etc). By American Joint Committee on Cancer (AJCC) and International Union Against Cancer (UICC) convention, LVI does not affect the T category indicating local extent of tumor unless specifically included in the definition of the T category.

E. Prognostic Factors
Typically, ES has a lobular growth pattern consisting of tumor cells that are distinctly monotonous in their nuclear uniformity. Nuclei measure 10 µm to 15 µm in diameter with distinct nuclear membranes, finely granular chromatin, and 1 to 2 inconspicuous nucleoli. Cytoplasm is poorly defined, scant, pale-staining, and may be vacuolated due to irregular glycogen deposition. Some cases of ES may show increased nuclear size, more pronounced atypia, and increased mitotic activity. Multinucleated giant cells are not seen. Large areas of tumor necrosis with “ghost-like tumor cells” may be striking and in some biopsy specimens may represent the majority of the tumor. Areas of neuroectodermal differentiation (Homer-Wright rosettes; rarely Flexner-Wintersteiner rosettes, ganglionic differentiation or primitive neuroepithelium) may be evident in some tumors. Some cases may show extensive epithelial differentiation, in particular the adamantinoma-like variant most commonly seen in the head-neck region. Currently, extraosseous Ewing sarcoma receives identical therapy as intraosseous Ewing sarcoma. There are no histopathologic ES subtypes that possess an established prognostic importance.

A summary of the prognostic factors is detailed below.\(^1\) Of all prognostic factors, age at onset, tumor size, site, and stage have proven to be the most important in predicting outcome.

<table>
<thead>
<tr>
<th>Factor</th>
<th>Favorable Prognosis</th>
<th>Unfavorable Prognosis</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age</td>
<td>Less than 10 years</td>
<td>Greater than or equal to 18 years</td>
</tr>
<tr>
<td></td>
<td>(EFS 69%); 10-17 years (EFS 74%)</td>
<td>(EFS 44%)</td>
</tr>
<tr>
<td>Site</td>
<td>Distal extremity</td>
<td>Pelvis (EFS 50%)</td>
</tr>
<tr>
<td></td>
<td>(EFS 74%); Proximal extremity (EFS 62%)</td>
<td></td>
</tr>
<tr>
<td>Size</td>
<td>Less than 8 cm greatest diameter (EFS 75%)</td>
<td>Greater than or equal to 8 cm in greatest dimension (EFS 55%)</td>
</tr>
<tr>
<td>Stage</td>
<td>Nonmetastatic tumor</td>
<td>Metastatic tumor (EFS approximately 20%)</td>
</tr>
<tr>
<td></td>
<td>(EFS approximately 70%)</td>
<td></td>
</tr>
</tbody>
</table>

Definition: EFS, event-free survival.

Histologic response to chemotherapy is an excellent predictor of outcome in osteosarcomas and may also be of value in ES. However, the evaluation of percentage necrosis in ES can be difficult, because unlike osteosarcoma, there is no residual acellular osteoid framework left to demarcate the original tumor bed. Furthermore, data regarding correlation of necrosis with outcome in extraosseous ES is not available. Currently, histologic assessment of percentage necrosis is not used formally to guide therapy in ES; however, it is recommended that the report includes the estimated percentage of necrosis.

References

F. Margins
The extent of resection (ie, gross residual disease versus complete resection with negative margins) has the strongest influence on local control of malignancy.\(^1\) The definition of what constitutes a sufficiently
“wide” margin of normal tissue in the management of ES and the significance of reactive and/or necrotic tissue at the margin are current study questions for the Children’s Oncology Group, and may evolve in the future. Currently, any tumor at the margin, whether viable, nonviable, or treated, is considered positive. The significance of treated tumor at the margin when there has been an excellent chemotherapeutic response (ie, greater than 90% tumor necrosis) remains unclear. There is currently no consensus as to whether margins involved by treated tumor require further treatment, and this is considered a negative margin on some studies. The presence of treated tumor at the margin should be reported, however, and can be included in the comment section of the checklist. The following margins are considered adequate:

- Cortical bone margin: 2 to 5 cm
- Fascia, periosteum, and intermuscular septa: 2 mm
- Fat, muscle, and medullary bone: 5 mm

With Ewing sarcoma involving an encapsulated organ, surgical margins are considered to be negative if the organ’s capsule is not surgically violated or breached by the tumor.

References

G. TNM and Stage Groupings
The AJCC TNM staging system for bone or soft tissue tumors may be used for pathologic staging of Ewing sarcoma and can be reported in the Comment section. However, the presence or absence of metastatic disease (a feature that may not be known to the pathologist) is the primary factor in the staging and treatment of pediatric patients with Ewing sarcoma.

References

H. Ancillary Studies
Immunohistochemistry
Immunohistochemistry with monoclonal antibodies against the cell surface glycoprotein CD99 is positive in virtually all cases of ES. This glycoprotein is diffusely expressed in the vast majority of cases in a membranous pattern (Figure 2). The results of staining using monoclonal antibodies O13, HBA71, and 12E7 are similar, but individual tumors may exhibit better staining with one of these antibodies versus other antibodies.
Figure 2. CD99 staining in Ewing sarcoma shows strong, diffuse, membranous staining. (CD99 antibody O13 with hematoxylin counterstain.)

Lymphoblastic lymphomas/leukemias, rhabdomyosarcomas, synovial sarcomas, solitary fibrous tumors, rhabdoid tumors, neuroendocrine tumors, desmoplastic small round cell tumors, and mesenchymal chondrosarcomas may also demonstrate immunoreactivity to CD99. In some of these tumors, CD99 immunostaining is often weakly granular and intracytoplasmic; in others (lymphoblastic lymphoma/leukemia, occasional cases of poorly differentiated synovial sarcoma, alveolar rhabdomyosarcoma), distinct membrane staining is present, as seen in ES. Because these other tumors with small round cell morphology can exhibit CD99 expression, it is very important to consider including other immunohistochemical stains such as muscle markers (desmin, muscle-specific actin, myoD1, myogenin), S-100, epithelial markers (epithelial membrane antigen, cytokeratin), INI-1, and lymphoid markers (CD45, CD30, Tdt, T-cell and/or B-cell markers) when CD99 is performed to properly exclude CD99-expressing tumors. Cytokeratin positivity may be seen in ES and may be diffusely positive in the adamantinoma-like variant of Ewing sarcoma.\textsuperscript{3,4} Newer immunohistochemical antibodies, such as NKX2.2, may also be useful for the diagnosis of ES, although NKX2.2 staining may rarely be seen in other small round cell tumors.\textsuperscript{4} The value of other immunohistochemical markers for diagnosis, such as Ki-67, p53, and C-kit (CD117), has not been established.

Chromosomal Translocations
The 2020 World Health Organization (WHO) classification of bone and soft tissue tumors defines Ewing sarcoma as a round cell sarcoma harboring a FET-ETS gene fusion. FET represents a family of genes to include FUS, EWSR1, and TAF15; whereas the ETS gene family is a large family of transcription factors involved in cell cycle regulation, cellular differentiation, among other functions. In relation to Ewing sarcoma, the characteristic translocations involve the EWSR1 gene at 22q12, most often either the FLI1 gene at 11q24 (90-95%) or the ERG gene at 21q22 (5-10%). These two fusions account for the vast majority of genetic alterations in ES. It should be emphasized that there are numerous other EWSR1 or FUS gene partners that occur in a minority (5%-10%) of ES. The failure to identify an EWSR1-FLI or EWSR1-ERG translocation by RT-PCR or cytogenetics does not exclude ES from the diagnosis. If RT-PCR is negative, in the context of a tumor suspicious for ES, other molecular studies (cytogenetics, NGS) may be important for identification of the less common ES translocations and for discovering novel EWSR1 translocations in ES. Some of the less common ES translocations involve FUS (ch16) rather than EWSR1, or involve other ETS partners including ETV1, ETV4, or FEV. FISH analysis for EWSR1 (or FUS) is helpful as a first step and may confirm the diagnosis in those tumors with histomorphologic
features and immunohistochemical phenotypes of ES. Because other small round cell tumors of childhood can have $EWSR1$ rearrangements with specific tumor-defining partners, $EWSR1$ FISH positivity alone is not diagnostic of ES. Some of these tumors with $EWSR1$ rearrangement include angiomatoid fibrous histiocytoma, clear cell sarcoma of soft parts, desmoplastic round cell tumor, and extraskeletal myxoid chondrosarcoma, as well as a subset of myxoid liposarcomas and myoepithelial carcinoma. This underscores the necessity for histologic and immunohistochemical correlation with FISH and/or cytogenetic data.5

Therefore, considerations when choosing testing methodologies may include, classic versus non-classic histomorphology, immunophenotype, need to confirm translocation partner, turnaround time, cost, and ultimately may be depend on the availability of testing modalities at each institution. While obtaining evidence of a diagnostic fusion is recommended, it should be noted that absence of a fusion can either result from 1) true lack of fusion, 2) test failure (eg. FISH for EWSR-ERG fusions can miss rearrangements) or 3) mismatch between the testing approach and the fusion present (eg. EWSR1-ERG present and test is for RT-PCR for EWSR1-FLI1).

Of note, the specific $EWSR1$ translocation and subtype based upon exon fusion type do not influence treatment, prognosis, or outcome.6

References