

# Protocol for the Examination of Biopsy Specimens From Patients With Invasive Carcinoma of the Breast

**Version:** Breast Invasive Biopsy 1.1.0.0 **Protocol Posting Date:** February 2020

The use of this protocol is recommended for clinical care purposes but is <u>not</u> required for accreditation purposes.

This protocol may be used for the following procedures AND tumor types:

ins protocol may be used for the following procedures AND tumor types.			
Procedure	Description		
Biopsy	Includes specimens designated needle biopsy, fine needle aspiration		
	and others (for excisional biopsy, see below)		
Tumor Type	Description		
Invasive breast carcinoma of	Includes microinvasive carcinoma and carcinoma with neuroendocrine		
any type, with or without	features		
ductal carcinoma in situ			
(DCIS)			

The following should NOT be reported using this protocol:

Procedure
Resection (consider Breast Invasive Carcinoma Resection protocol)
Excisional biopsy (consider Breast Invasive Carcinoma Resection protocol)
Tumor Type
Ductal carcinoma in situ (DCIS) without invasive carcinoma (consider the DCIS Biopsy protocol)
Paget disease of the nipple without invasive carcinoma (consider the DCIS Biopsy protocol)
Encapsulated or solid papillary carcinoma without invasion (consider the Breast DCIS Biopsy protocol)
Phyllodes tumor
Lymphoma (consider the Hodgkin or non-Hodgkin Lymphoma protocols)
Sarcoma (consider the Soft Tissue protocol)

#### **Authors**

Patrick L. Fitzgibbons, MD\*; James L. Connolly\*, MD; Mary Edgerton, MD, PhD; Ross Simpson, MD. With guidance from the CAP Cancer and CAP Pathology Electronic Reporting Committees.

#### **Accreditation Requirements**

The use of this biopsy case summary is recommended for clinical care purposes, but is not required for accreditation purposes. The core and conditional data elements are routinely reported for biopsy specimens. Noncore data elements are included to allow for reporting information that may be of clinical value.

#### **Summary of Changes**

V1.1.0.0 Modified Histologic Type (WHO 2019) Included Tumor Size

<sup>\*</sup> Denotes primary author. All other contributing authors are listed alphabetically.

### **Surgical Pathology Cancer Case Summary**

Protocol posting date: February 2020

#### **INVASIVE CARCINOMA OF THE BREAST: Biopsy**

#### Notes:

This case summary is recommended for reporting biopsy specimens but is NOT REQUIRED for accreditation purposes. Core data elements are bolded to help identify routinely reported elements.

Select a single response unless otherwise indicated.

Procedure, Laterality, and Site may be listed separately or on 1 line.

Procedure
Needle biopsy
Fine needle aspiration
Other (specify):
Not specified
Specimen Laterality
Right
Left
Not specified
Tumor Site (coloct all that apply)
Tumor Site (select all that apply) Upper outer quadrant
Lower outer quadrant
Upper inner quadrant
Lower inner quadrant
Central
Central Nipple
o'clocko'clock
Distance from nipple (centimeters):cm
Other (specify):
Not specified
Not specified
Tumor Size
Microinvasion only (≤1 mm)
Greatest dimension of largest invasive focus >1 mm (millimeters): mm, or at least mm
Greatest dimension of largest invasive focus >1 mm (millimeters): at least mm
Tumor size cannot be determined (explain):
Histologic Type (Note A)
No residual invasive carcinoma
Invasive carcinoma of no special type (ductal)
Micro-invasive carcinoma
Invasive lobular carcinoma
Invasive carcinoma with mixed ductal and lobular features
Invasive carcinoma with features of (specify):
Tubular carcinoma
Invasive cribriform carcinoma
Mucinous carcinoma
Invasive micropapillary carcinoma
Apocrine adenocarcinoma

Metaplastic carcinoma Encapsulated papillary carcinoma with invasion
Solid papillary carcinoma with invasion
Intraductal papillary adenocarcinoma with invasion
Adenoid cystic carcinoma
Neuroendocrine tumor
Neuroendocrine carcinoma Invasive carcinoma, type cannot be determined
Other histologic type (specify):
Histologic Grade (Nottingham Histologic Score) (Note B)  Not applicable (microinvasion only)
Glandular (Acinar)/Tubular Differentiation
Score 1 (>75% of tumor area forming glandular/tubular structures)
Score 2 (10% to 75% of tumor area forming glandular/tubular structures)
<ul><li>Score 3 (&lt;10% of tumor area forming glandular/tubular structures)</li><li>Score cannot be determined</li></ul>
Nuclear Pleomorphism
Score 1 (nuclei small with little increase in size in comparison with normal breast epithelial cells, regular
outlines, uniform nuclear chromatin, little variation in size)
Score 2 (cells larger than normal with open vesicular nuclei, visible nucleoli, and moderate variability in both size and shape)
Score 3 (vesicular nuclei, often with prominent nucleoli, exhibiting marked variation in size and shape,
occasionally with very large and bizarre forms)
Score cannot be determined
Mitatia Data (ana Tabia 4)
Mitotic Rate (see Table 1) Score 1
Score 2
Score 3
Score cannot be determined
Overall Create
Overall Grade Grade 1 (scores of 3, 4, or 5)
Grade 2 (scores of 6 or 7)
Grade 3 (scores of 8 or 9)
Score cannot be determined (explain):
Ductal Carcinoma In Situ (DCIS) (Note C)
Not identified
Present
Cannot be excluded
Architectural Patterns (if DCIS is present select all that apply)
Comedo Paget disease (DCIS involving nipple skin)
Cribriform
Micropapillary
Papillary Papillary
Solid
Other (specify):

The routinely reported core data elements are bolded.

\_\_\_ Grade I (low)

Grade II (intermediate) Grade III (high)
Necrosis (if DCIS is present)  Not identified Present, focal (small foci or single cell necrosis) Present, central (expansive "comedo" necrosis)
Lymphovascular Invasion  Not identified  Present Cannot be determined
Additional Pathologic Findings (Note D) Specify:
Microcalcifications (select all that apply) (Note E)  Not identified Present in DCIS Present in invasive carcinoma Present in non-neoplastic tissue Other (specify):
Ancillary Studies
Note: For hormone receptor and HER2 reporting, the CAP Breast Biomarker Template should be used. www.cap.org/cancerprotocols.
Biomarker Studies Pending
Comment(s)

#### A. Histologic Type

This protocol applies to all invasive carcinomas of the breast. The World Health Organization (WHO)<sup>1</sup> classification of breast carcinoma is recommended, although the protocol does not preclude the use of other classifications or histologic types. Carcinomas may be classified based on the H&E appearance without the use of immunohistochemical studies.

A modified list is presented in the case summary based on the most frequent types of invasive carcinomas and terminology that is in widespread usage. The modified list is intended to capture the majority of tumors and reduce the frequency of tumors being reported as "other." Choices are added for tumors with mixed features and those with some but not all features of specific histologic types.

WHO Classification of Invasive Carcinoma of the Breast <sup>1</sup>
No residual invasive carcinoma
<ul><li>Invasive carcinoma of no special type (ductal)</li><li>Micro-invasive carcinoma</li></ul>
Invasive lobular carcinoma
Invasive lobular carcinoma Invasive carcinoma with mixed ductal and lobular features
Invasive carcinoma with mixed features (specify): Tubular carcinoma
Invasive cribriform carcinoma
Mucinous carcinoma
Invasive micropapillary carcinoma
Apocrine adenocarcinoma
Apocinic adenocatomorna
Metaplastic Carcinoma
Metaplastic carcinoma NOS
Low grade adenosquamous carcinoma
Fibromatosis-like metaplastic carcinoma
Spindle cell carcinoma
Squamous cell carcinoma
Metaplastic carcinoma with mesenchymal differentiation
Encapsulated papillary carcinoma with invasion
Solid papillary carcinoma with invasion
Intraductal papillary adenocarcinoma with invasion
Adenoid cystic carcinoma
Neuroendocrine Tumor
Neuroendocrine tumor NOS
Neuroendocrine tumor, grade 1
Neuroendocrine tumor, grade 2
Neuroendocrine Carcinoma
Neuroendocrine carcinoma NOS
Neuroendocrine carcinoma, small cell
Neuroendocrine carcinoma, large cell
Invasive carcinoma, type cannot be determined
Other histologic type (specify):
Invasive papillary carcinoma
Oncocytic carcinoma
Lipid-rich carcinoma
Glycogen-rich carcinoma
Sebaceous carcinoma
Mucinous cystadenocarcinoma NOS
Acinar cell carcinoma

## Background Documentation

Classic adenoid cystic carcinoma
Solid-basaloid adenoid cystic carcinoma
Adenoid cystic carcinoma with high-grade transformation
Secretory carcinoma
Mucoepidermoid carcinoma
Polymorphous adenocarcinoma
Tall cell carcinoma with reversed polarity
Adenomyoepithelioma with carcinoma
Epithelial-myoepithelial carcinoma
Other type not listed (specify):

#### References

1. WHO Classification of Tumours Editorial Board. *Breast tumours*. Lyon (France): International Agency for Research on Cancer; 2019. (WHO classification of tumours series, 5th ed.; vol. 2).

#### **B.** Histologic Grade

All invasive breast carcinomas should be graded.<sup>1</sup> The Nottingham combined histologic grade (Elston-Ellis modification of Scarff-Bloom-Richardson grading system) should be used for reporting. Within each stage grouping there is a relation between histologic grade and outcome.

The Nottingham combined histologic grade evaluates the amount of tubule formation, the extent of nuclear pleomorphism, and the mitotic count (or mitotic rate). Each variable is given a score of 1, 2, or 3, and the scores are added to produce a grade. The mitotic score is determined by the number of mitotic figures found in 10 consecutive high-power fields (HPF) in the most mitotically active part of the tumor. Only clearly identifiable mitotic figures should be counted; hyperchromatic, karyorrhectic, or apoptotic nuclei are excluded. Because of variations in field size, the HPF size must be determined for each microscope and the appropriate point score determined accordingly. It is recommended that the size be measured by using a micrometer. However, the diameter of an HPF can also be calculated by using the method below.

#### Measuring the Size of a High-Power Field (HPF) With a Ruler

Use a clear ruler to measure the diameter of a low-power field. This number can be used to calculate a constant based on the following formula:

Eyepiece Magnification x Objective Magnification x Microscopic Field Diameter = A Constant

When the value of the constant is known, the diameter of an HPF can be calculated for other objectives by using the following formula:

<u>Unknown Field Diameter = Constant/(Eyepiece Magnification x Objective Magnification)</u>

Half of the field diameter is the radius of the field (r), which can then be used to calculate the area of the HPF: 3.1415 x  $r^2$  = Area of Microscopic Field

If the microscopic field diameter or the area of the field is known, Table 1 can be used to determine the number of mitoses corresponding to different scores.

Table 1. Score Categories According to Field Diameter and Mitotic Count

Scoring Categories of	Mitotic Counts			
		Number of mitoses per 10 fields corresponding to:		
Field diameter (mm)	Area (mm²)	Score 1	Score 2	Score 3
0.40	0.125	≤4	5 to 9	≥10
0.41	0.132	≤4	5 to 9	≥10
0.42	0.139	≤5	6 to 10	≥11
0.43	0.145	≤5	6 to 10	≥11
0.44	0.152	≤5	6 to 11	≥12
0.45	0.159	≤5	6 to 11	≥12
0.46	0.166	≤6	7 to 12	≥13
0.47	0.173	≤6	7 to 12	≥13
0.48	0.181	≤6	7 to 13	≥14
0.49	0.189	≤6	7 to13	≥14
0.50	0.196	≤7	8 to 14	≥15
0.51	0.204	≤7	8 to 14	≥15
0.52	0.212	≤7	8 to 15	≥16
0.53	0.221	≤8	9 to 16	≥17
0.54	0.229	≤8	9 to 16	≥17
0.55	0.238	≤8	9 to 17	≥18
0.56	0.246	≤8	9 to 17	≥18
0.57	0.255	≤9	10 to 18	≥19
0.58	0.264	≤9	10 to 19	≥20
0.59	0.273	≤9	10 to 19	≥20
0.60	0.283	≤10	11 to 20	≥21
0.61	0.292	≤10	11 to 21	≥22
0.62	0.302	≤11	12 to 22	≥23
0.63	0.312	≤11	12 to22	≥23
0.64	0.322	≤11	12 to 23	≥24
0.65	0.332	≤12	13 to 24	≥25
0.66	0.342	≤12	13 to 24	≥25
0.67	0.353	≤12	13 to 25	≥26
0.68	0.363	≤13	14 to 26	≥27
0.69	0.374	≤13	14 to 27	≥ 28

From Pathology Reporting of Breast Disease.<sup>2</sup> Copyright 2005 National Health Service Cancer Screening Programme and The Royal College of Pathologists. Adapted with permission.

#### References

- 1. Ellis IO, Elston CW. Histologic grade. In: O'Malley FP, Pinder SE, eds. *Breast Pathology*. Philadelphia, PA: Elsevier; 2006:225-233.
- Pathology Reporting of Breast Disease. A Joint Document Incorporating the Third Edition of the NHS Breast Screening Programme's Guidelines for Pathology Reporting in Breast Cancer Screening and the Second Edition of The Royal College of Pathologists' Minimum Dataset for Breast Cancer Histopathology Published by the NHS Cancer Screening Programmes jointly with The Royal College of Pathologists. NHSBSP Publication No 58. January 2005. http://www.cancerscreening.nhs.uk/ breastscreen/publications/nhsbsp58.html. Accessed April 8, 2009.

#### C. Ductal Carcinoma In Situ

#### **Nuclear Grade of DCIS**

The nuclear grade of DCIS is determined using 6 morphologic features (Table 1).1

Table 2. Nuclear Grade of Ductal Carcinoma in Situ

Feature	Grade I (Low)	Grade II (Intermediate)	Grade III (High)
Pleomorphism	Monotonous (monomorphic)	Intermediate	Markedly pleomorphic
Size	1.5 to 2 x the size of a normal red blood cell or a normal duct epithelial cell nucleus	Intermediate	>2.5 x the size of a normal red blood cell or a normal duct epithelial cell nucleus
Chromatin	Usually diffuse, finely dispersed chromatin	Intermediate	Usually vesicular with irregular chromatin distribution
Nucleoli	Only occasional	Intermediate	Prominent, often multiple
Mitoses	Only occasional	Intermediate	May be frequent
Orientation	Polarized toward luminal spaces	Intermediate	Usually not polarized toward the luminal space

#### Necrosis

The presence of necrosis is correlated with the finding of mammographic calcifications (ie, most areas of necrosis will calcify). Ductal carcinoma in situ that presents as mammographic calcifications often recurs as calcifications. Necrosis can be classified as follows:

- <u>Central ("comedo")</u>: The central portion of an involved ductal space is replaced by an area of expansive
  necrosis that is easily detected at low magnification. Ghost cells and karyorrhectic debris are generally
  present. Although central necrosis is generally associated with high-grade nuclei (ie, comedo DCIS), it can
  also occur with DCIS of low or intermediate nuclear grade.
- Focal: Small foci, indistinct at low magnification, or single cell necrosis.

Necrosis should be distinguished from secretory material, which can also be associated with calcifications, but does not include nuclear debris.

#### References

1. Schwartz GF, Lagios MD, Carter D, et al. Consensus conference on the classification of ductal carcinoma in situ. *Cancer.* 1997;80:1798-1802.

#### D. Additional Pathologic Findings

In some cases, additional pathologic findings are important for the clinical management of patients. If multiple invasive carcinomas are present and differ in histologic type, grade, or the expression of ER, PgR, or HER2, this information should be included as text in this section.

#### E. Microcalcifications

Cancer found in biopsies performed for microcalcifications will almost always be at the site of the calcifications or in close proximity. The presence of the targeted calcifications in the specimen should be confirmed by specimen radiography. The pathologist must be satisfied that the specimen has been sampled in such a way that the lesion responsible for the calcifications has been examined microscopically. The relationship of the radiologic calcifications to the invasive carcinoma and the DCIS should be indicated.

If calcifications can be seen in the specimen radiograph but not in the initial histologic sections, deeper levels should be examined. If needed, radiographs of the paraffin block(s) may be obtained to detect calcifications remaining in the block(s). If microcalcifications cannot be confirmed by routine microscopic evaluation, polarized light may be helpful, since calcium oxalate crystals are refractile and polarizable but usually clear or tinged yellow in H&E sections. On rare occasions, calcifications do not survive tissue processing or prolonged fixation in formalin. Foreign material can sometimes simulate calcifications (eg, metallic fragments after surgery or trauma).