Protocol for the Examination of Specimens From Patients With Primary Tumors of the Ovary, Fallopian Tube, or Peritoneum

Version: Ovary Fallopian 1.1.1.0
Protocol Posting Date: February 2020
CAP Laboratory Accreditation Program Protocol Required Use Date: November 2020


For accreditation purposes, this protocol should be used for the following procedures AND tumor types:

<table>
<thead>
<tr>
<th>Procedure</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>Resection</td>
<td>Includes oophorectomy, salpingo-oophorectomy, salpingectomy, subtotal resection, or removal of tumor in fragments</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Tumor Type Description</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>Primary malignant tumors of ovary, fallopian tube or peritoneum</td>
<td>Includes all primary epithelial borderline tumors and carcinomas, carcinosarcoma, malignant germ cell tumors, and malignant sex cord-stromal tumors.</td>
</tr>
</tbody>
</table>

This protocol is NOT required for accreditation purposes for the following:

<table>
<thead>
<tr>
<th>Procedure</th>
</tr>
</thead>
<tbody>
<tr>
<td>Biopsy</td>
</tr>
<tr>
<td>Primary resection specimen with no residual cancer (eg, following neoadjuvant therapy)</td>
</tr>
<tr>
<td>Cytologic specimens</td>
</tr>
</tbody>
</table>

The following tumor types should NOT be reported using this protocol:

<table>
<thead>
<tr>
<th>Tumor Type</th>
</tr>
</thead>
<tbody>
<tr>
<td>Peritoneal mesothelioma</td>
</tr>
<tr>
<td>Lymphoma (consider the Hodgkin or non-Hodgkin Lymphoma protocols)</td>
</tr>
<tr>
<td>Sarcoma (consider the Soft Tissue protocol)</td>
</tr>
</tbody>
</table>

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With guidance from the CAP Cancer and CAP Pathology Electronic Reporting Committees.

* Denotes primary authors. All other contributing authors are listed alphabetically.

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Accreditation Requirements
This protocol can be utilized for a variety of procedures and tumor types for clinical care purposes. For accreditation purposes, only the definitive primary cancer resection specimen is required to have the core and conditional data elements reported in a synoptic format.

- **Core data elements** are required in reports to adequately describe appropriate malignancies. For accreditation purposes, essential data elements must be reported in all instances, even if the response is “not applicable” or “cannot be determined.”

- **Conditional data elements** are only required to be reported if applicable as delineated in the protocol. For instance, the total number of lymph nodes examined must be reported, but only if nodes are present in the specimen.

- **Optional data elements** are identified with “+” and although not required for CAP accreditation purposes, may be considered for reporting as determined by local practice standards.

The use of this protocol is not required for recurrent tumors or for metastatic tumors that are resected at a different time than the primary tumor. Use of this protocol is also not required for pathology reviews performed at a second institution (ie, secondary consultation, second opinion, or review of outside case at second institution).

Synoptic Reporting
All core and conditionally required data elements outlined on the surgical case summary from this cancer protocol must be displayed in synoptic report format. Synoptic format is defined as:

- Data element: followed by its answer (response), outline format without the paired “Data element: Response” format is NOT considered synoptic.

- The data element should be represented in the report as it is listed in the case summary. The response for any data element may be modified from those listed in the case summary, including “Cannot be determined” if appropriate.

- Each diagnostic parameter pair (Data element: Response) is listed on a separate line or in a tabular format to achieve visual separation. The following exceptions are allowed to be listed on one line:
  - Anatomic site or specimen, laterality, and procedure
  - Pathologic Stage Classification (pTNM) elements
  - Negative margins, as long as all negative margins are specifically enumerated where applicable

- The synoptic portion of the report can appear in the diagnosis section of the pathology report, at the end of the report or in a separate section, but all Data element: Responses must be listed together in one location. Organizations and pathologists may choose to list the required elements in any order, use additional methods in order to enhance or achieve visual separation, or add optional items within the synoptic report. The report may have required elements in a summary format elsewhere in the report IN ADDITION TO but not as replacement for the synoptic report ie, all required elements must be in the synoptic portion of the report in the format defined above.

Summary of Changes
Version 1.1.1.0
The following data elements were modified:
pT3 – Minor typographical correction
Added Neoplastic cells (applicable only to borderline / atypical proliferative tumor) to Peritoneal/Ascitis Fluid
Surgical Pathology Cancer Case Summary

Protocol posting date: February 2020

**OVARY or FALLOPIAN TUBE or PRIMARY PERITONEUM:**

*Note: Applies to primary tumors of ovarian, fallopian tube, or primary peritoneal origin. If bilateral tumors of 2 different histologic types are present, separate case summaries should be used for each tumor.*

Select a single response unless otherwise indicated.

**Procedure (select all that apply) (Note A)**

- Total hysterectomy and bilateral salpingo-oophorectomy
- Radical hysterectomy
- Simple hysterectomy
- Supracervical hysterectomy
- Bilateral salpingo-oophorectomy
- Right salpingo-oophorectomy
- Left salpingo-oophorectomy
- Salpingo-oophorectomy, side not specified
- Right oophorectomy
- Left oophorectomy
- Oophorectomy, side not specified
- Bilateral salpingectomy
- Right salpingectomy
- Left salpingectomy
- Salpingectomy, side not specified
- Omentectomy
- Peritoneal biopsies
- Peritoneal tumor debulking
- Peritoneal washing
- Pleurocentesis (pleural fluid)
- Other (specify): ____________________________

*Note: For information about lymph node sampling, please refer to the Regional Lymph Node section.*

**Hysterectomy Type**

- Abdominal
- Vaginal
- Vaginal, laparoscopic-assisted
- Laparoscopic
- Laparoscopic, robotic-assisted
- Other (specify): ___________________________
- Not specified

**Specimen Integrity (Note B)**

*Note: For primary ovarian tumors, if the ovary containing primary tumor is removed intact into a laparoscopy bag and ruptured in the bag by the surgeon without spillage into the peritoneal cavity (to allow for removal via laparoscopy port site or small incision), the specimen integrity should be listed as “capsule intact” with a comment explaining this in the report.*

**Specimen Integrity of Right Ovary (if applicable)**

- Capsule intact
- Capsule ruptured
- Fragmented
- Other (specify): ____________________________

+ Data elements preceded by this symbol are not required for accreditation purposes. These optional elements may be clinically important but are not yet validated or regularly used in patient management.
Specimen Integrity of Left Ovary (if applicable)
___ Capsule intact
___ Capsule ruptured
___ Fragmented
___ Other (specify): ____________________________

Specimen Integrity of Right Fallopian Tube (if applicable)
___ Serosa intact
___ Serosa ruptured
___ Fragmented
___ Other (specify): ____________________________

Specimen Integrity of Left Fallopian Tube (if applicable)
___ Serosa intact
___ Serosa ruptured
___ Fragmented
___ Other (specify): ____________________________

+ Morcellated specimen (specify organ): ____________________________

Tumor Site (Notes C, D, and E)
Note: Please select the primary tumor site only. Information about additional organ involvement is provided in another section.
___ Right ovary
___ Left ovary
___ Bilateral ovaries
___ Ovary, laterality cannot be determined (explain): _______________________
___ Right fallopian tube
___ Left fallopian tube
___ Bilateral fallopian tubes
___ Fallopian tube, laterality cannot be determined (explain): _______________________
___ Right tubo-ovarian
___ Left tubo-ovarian
___ Bilateral tubo-ovarian
___ Tubo-ovarian, laterality cannot be determined (explain): _______________________
___ Primary peritoneum
___ Other (specify): _______________________

Ovarian Surface Involvement (required only if applicable)
___ Present
    Specify laterality (if applicable): _______________________
___ Absent
___ Cannot be determined (explain): _______________________

Fallopian Tube Surface Involvement (required only if applicable)
___ Present
    Specify laterality (if applicable): _______________________
___ Absent
___ Cannot be determined (explain): _______________________

Tumor Size
Note: For bilateral tumors, please report maximum dimension for each primary tumor, specifying by laterality.
Greatest dimension (centimeters): ___ cm
+ Additional dimensions (centimeters): ___ x ___ cm
___ Cannot be determined (explain): _______________________

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**Histologic Type (select all that apply) (Notes F and G)**

- Serous tubal intraepithelial carcinoma (STIC)
- Serous borderline tumor/atypical proliferative serous tumor
- Serous borderline tumor/atypical proliferative serous tumor with microinvasion
- Serous borderline tumor-micropapillary variant/noninvasive low-grade serous carcinoma
- Serous carcinoma
- Endometrioid borderline tumor/atypical proliferative endometrioid tumor
- Endometrioid borderline tumor/atypical proliferative endometrioid tumor with microinvasion
- Endometrioid carcinoma
- Clear cell borderline tumor/atypical proliferative clear cell tumor
- Clear cell carcinoma
- Mucinous borderline tumor/atypical proliferative mucinous tumor
- Mucinous borderline tumor/atypical proliferative mucinous tumor with intraepithelial carcinoma
- Mucinous borderline tumor/atypical proliferative mucinous tumor with microinvasion
- Mucinous carcinoma
- Mucinous carcinoma with expansile invasive pattern
- Mucinous carcinoma with infiltrative invasive pattern
- Seromucinous borderline tumor/atypical proliferative seromucinous tumor
- Seromucinous borderline tumor/atypical proliferative seromucinous tumor with microinvasion
- Seromucinous carcinoma
- Brenner tumor, borderline/atypical proliferative Brenner tumor
- Brenner tumor, malignant
- Carcinoma, subtype cannot be determined
- Mixed epithelial borderline (atypical proliferative) tumor (specify types and percentages): ___________
- Mixed epithelial carcinoma (specify types and percentages): ________________________
- Carcinosarcoma (malignant mixed Müllerian tumor)
- Small cell carcinoma, pulmonary type
- Small cell carcinoma, hypercalcemic type
- Squamous cell carcinoma
- Transitional cell carcinoma
- Undifferentiated carcinoma
- Granulosa cell tumor, adult type
- Granulosa cell tumor, juvenile type
- Other sex cord-stromal tumor (specify type): ________________________
- Dysgerminoma
- Yolk sac tumor (endodermal sinus tumor)
- Immature teratoma
- Carcinoma arising from a teratoma (specify type): ______________________
- Mixed malignant germ cell tumor (specify types and percentages): ______________________
- Other histologic type not listed (specify): ______________________

**Histologic Grade (required for serous, endometrioid, mucinous, and seromucinous carcinomas, immature teratomas, and Sertoli-Leydig cell tumors) (Note H)**

*Note: Serous carcinomas are graded via a 2-tier system. Immature teratomas can be graded using a 2-tier or 3-tier system. Endometrioid and mucinous carcinomas are graded via a 3-tier system. Sertoli-Leydig cell tumors are graded via a modified 3-tier grading system with grade 2 tumors being termed “intermediate differentiated.” Clear cell carcinomas, borderline epithelial neoplasms, carcinosarcomas, all other malignant sex-cord stromal and germ cell tumors are not graded.*

**WHO Grading System**

- G1: Well differentiated
- G2: Moderately differentiated
- G3: Poorly differentiated
- GX: Cannot be assessed

**Two-Tier Grading System (required for immature teratomas and serous carcinomas only)**

- Low grade

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___ High grade
___ Other (specify): _____________________________
___ Not applicable

**Implants (required for advanced stage serous/seromucinous borderline tumors only) (Note I)**

*Note: Serous tumor implants that were formerly classified as "invasive implants" are now classified as low-grade serous carcinoma of the peritoneum.*

___ Not sampled
___ Not identified
___ Present (specify sites): _______________________

**Other Tissue/ Organ Involvement (select all that apply)**

*Note: Any organ not selected is either not involved or was not submitted.*

___ Not applicable
___ Not identified
___ Right ovary
___ Left ovary
___ Ovary (side not specified)
___ Right fallopian tube
___ Left fallopian tube
___ Fallopian tube (side not specified)
___ Uterus
___ Cervix
___ Pelvic peritoneum
___ Abdominal peritoneum
___ Omentum
___ Other organs/tissue (specify): _______________________
___ Cannot be determined (explain): _______________________

**Largest Extrapelvic Peritoneal Focus (required only if applicable)**

___ Microscopic
___ Macroscopic (2 cm or less)
___ Macroscopic (greater than 2 cm)
___ Cannot be determined (explain): _______________________

Specify site (if applicable): _______________________

**Peritoneal/Ascitic Fluid**

___ Not submitted/unknown
___ Negative for malignancy (normal/benign)
___ Atypical and/or suspicious (explain): _______________________
___ Neoplastic cells (applicable only to borderline/atypical proliferative tumor)
___ Malignant (positive for malignancy)
___ Unsatisfactory/nondiagnostic (explain): _______________________
___ Other (specify): _______________________
___ Results pending

**Pleural Fluid**

+ ___ Not submitted / unknown
+ ___ Negative for malignancy (normal/benign)
+ ___ Atypical and/or suspicious (explain): _______________________
+ ___ Malignant (positive for malignancy)
+ ___ Unsatisfactory/nondiagnostic (explain): _______________________
+ ___ Results pending

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Treatment Effect (required only for high-grade serous carcinomas) (Note J)

Note: Treatment effect is based on assessment of residual tumor in the omentum.

___ No known presurgical therapy
___ No definite or minimal response identified (chemotherapy response score 1 [CRS 1])
___ Moderate response identified (CRS 2)
___ Marked response with no or minimal residual cancer (CRS 3)
___ Cannot be determined

Regional Lymph Nodes

Note: For ovarian, fallopian tube, or primary peritoneal tumors, lymph nodes designated as pelvic; (parametrial, obturator, internal iliac [hypogastric], external iliac, common iliac, sacral, presacral), para-aortic, and retroperitoneal are considered regional lymph nodes. Although not specifically named by AJCC or FIGO, intra-omental and peri-intestinal lymph nodes, are also regarded as regional lymph nodes for staging purposes. Any other involved nodes should be categorized as metastases (pM1) and commented on in the distant metastasis section. Presence of isolated tumor cells no greater than 0.2 mm in regional lymph node(s) is considered N0 (+).

___ No lymph nodes submitted or found

Lymph Node Examination (required only if lymph nodes are present in the specimen)

___ All lymph nodes negative for tumor cells
___ Positive for tumor cells (select all that apply)
   Number of Nodes with Metastasis Greater than 10 mm: ______
   Number of Nodes with Metastasis 10 mm or Less (excludes isolated tumor cells): ______
   Number of Nodes with Isolated Tumor Cells (0.2 mm or less) (if applicable): ______
   Number cannot be determined (explain): ________________________________

Note: Reporting the number of lymph nodes with isolated tumor cells is required only in the absence of metastasis greater than 0.2 mm in other lymph nodes.

+ Nodal Site(s) with Tumor Cells (specify): __________________________

Size of Largest Metastatic Deposit (if applicable) (millimeters): _____ mm

Location of Largest Deposit (if applicable) (specify): __________________________

Number of Lymph Nodes Examined: ______
   Number cannot be determined (explain): ________________________________
   + Specify Site(s): __________________________

Pathologic Stage Classification (pTNM, AJCC 8th Edition) (Note K)

Note: Reporting of pT, pN, and (when applicable) pM categories is based on information available to the pathologist at the time the report is issued. Only the applicable T, N, or M category is required for reporting; their definitions need not be included in the report. The categories (with modifiers when applicable) can be listed on 1 line or more than 1 line.

TNM Descriptors (required only if applicable) (select all that apply)
___ m (multiple primary tumors)
___ r (recurrent)
___ y (posttreatment)

Primary Tumor (pT)
___ pTX: Primary tumor cannot be assessed
___ pT0: No evidence of primary tumor
___ pT1: Tumor limited to ovaries (one or both) or fallopian tube(s)
___ pT1a: Tumor limited to one ovary (capsule intact) or fallopian tube, no tumor on ovarian or fallopian tube surface; no malignant cells in ascites or peritoneal washings #

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__pT1b:__ Tumor limited to both ovaries (capsules intact) or fallopian tubes; no tumor on ovarian or fallopian tube surface; no malignant cells in ascites or peritoneal washings

__pT1c:__ Tumor limited to one or both ovaries or fallopian tubes with any of the following:

__pT1c1:__ Surgical spill
__pT1c2:__ Capsule ruptured before surgery or tumor on ovarian or fallopian tube surface
__pT1c3:__ Malignant cells in ascites or peritoneal washings

__pT2:__ Tumor involves one or both ovaries or fallopian tubes with pelvic extension below pelvic brim or primary peritoneal cancer

__pT2a:__ Extension and/or implants on uterus and/or fallopian tube(s) and/or ovaries.
__pT2b:__ Extension to and/or implants on other pelvic tissues.

__pT3:__ Tumor involves one or both ovaries or fallopian tubes, or primary peritoneal cancer, with microscopically confirmed peritoneal metastasis outside the pelvis and/or metastasis to retroperitoneal (pelvic and/or para-aortic) lymph nodes

__pT3a:__ Microscopic extra-pelvic (above the pelvic brim) peritoneal involvement with or without positive retroperitoneal lymph nodes
__pT3b:__ Macroscopic peritoneal metastasis beyond pelvis 2 cm or less in greatest dimension with or without metastasis to retroperitoneal lymph nodes
__pT3c:__ Macroscopic peritoneal metastasis beyond pelvis more than 2 cm in greatest dimension with or without metastasis to the retroperitoneal lymph nodes (includes extension of tumor to capsule of liver and spleen without parenchymal involvement of either organ)

# Note: Serous tubal intraepithelial carcinoma (STIC) should be staged as pT1a if it involves one tube only, as pT1b if it involves both tubes, and as pT1c3 if it is accompanied by positive peritoneal washing washings or ascites. Nonmalignant ascites is not classified. The presence of ascites does not affect staging unless malignant cells are present.

**Regional Lymph Nodes (pN)**

__pNX:__ Regional lymph nodes cannot be assessed
__pN0:__ No regional lymph node metastasis
__pN0(i+):__ Isolated tumor cells in regional lymph node(s) no greater than 0.2 mm
__pN1:__ Positive retroperitoneal lymph nodes only (histologically confirmed)
__pN1a:__ Metastasis up to and including 10 mm in greatest dimension
__pN1b:__ Metastasis more than 10 mm in greatest dimension

**Distant Metastasis (pM) (required only if confirmed pathologically in this case)**

__pM1:__ Distant metastasis, including pleural effusion with positive cytology; liver or splenic parenchymal metastasis; metastasis to extra-abdominal organs (including inguinal lymph nodes and lymph nodes outside the abdominal cavity); and transmural involvement of intestine

__pM1a:__ Pleural effusion with positive cytology
__pM1b:__ Liver or splenic parenchymal metastases; metastases to extra-abdominal organs (including inguinal lymph nodes and lymph nodes outside the abdominal cavity); transmural involvement of intestine

Specify site(s), if known: ____________________________

# Note: Parenchymal liver or splenic metastasis is classified as stage IV disease, whereas liver or splenic capsule metastasis is classified as stage III disease. Non-regional lymph node metastases (such as inguinal, supraclavicular, and axillary nodes) are considered M1. Involvement of surface of diaphragm is considered pT3; however, involvement of skeletal muscle of diaphragm or abdominal wall tissue beyond the peritoneum is considered distant metastasis (M1).
+ FIGO Stage (2018 FIGO Cancer Report)
+ ___ I: Tumor confined to ovaries or fallopian tube(s)
+ ___ IA: Tumor limited to 1 ovary (capsule intact) or fallopian tube; no tumor on ovarian or fallopian tube surface; no malignant cells in the ascites or peritoneal washings
+ ___ IB: Tumor limited to both ovaries (capsules intact) or fallopian tubes; no tumor on ovarian or fallopian tube surface; no malignant cells in the ascites or peritoneal washings
+ ___ IC: Tumor limited to 1 or both ovaries or fallopian tubes, with any of the following:
  + ___ IC1: Surgical spill intraoperatively
  + ___ IC2: Capsule ruptured before surgery or tumor on ovarian or fallopian tube surface
  + ___ IC3: Malignant cells present in the ascites or peritoneal washings
+ ___ II: Tumor involves 1 or both ovaries or fallopian tubes with pelvic extension (below pelvic brim) or peritoneal cancer
  + ___ IIA: Extension and/or implants on the uterus and/or fallopian tubes and/or ovaries
  + ___ IIB: Extension to other pelvic intraperitoneal tissues
+ ___ III: Tumor involves 1 or both ovaries, or fallopian tubes, or primary peritoneal cancer, with cytologically or histologically confirmed spread to the peritoneum outside the pelvis and/or metastasis to the retroperitoneal lymph nodes
  + ___ IIIA: Metastasis to the retroperitoneal lymph nodes with or without microscopic peritoneal involvement beyond the pelvis
  + ___ IIIA1: Positive retroperitoneal lymph nodes only (cytologically or histologically proven)
  + ___ IIIA1(i): Metastasis ≤10 mm in greatest dimension#
  + ___ IIIA1(ii): Metastasis >10 mm in greatest dimension#
  + ___ IIIA2: Microscopic extrapelvic (above the pelvic brim) peritoneal involvement with or without positive retroperitoneal lymph nodes
+ ___ IIIB: Macroscopic peritoneal metastases beyond the pelvic brim ≤2 cm in greatest dimension, with or without metastasis to the retroperitoneal lymph nodes
+ ___ IIIC: Macroscopic peritoneal metastases beyond the pelvic brim >2 cm in greatest dimension, with or without metastases to the retroperitoneal nodes##
+ ___ IV: Distant metastasis excluding peritoneal metastases
  + ___ IVA: Pleural effusion with positive cytology
  + ___ IVB: Metastases to extra-abdominal organs (including inguinal lymph nodes and lymph nodes outside of abdominal cavity)###

# This is tumor dimension and not lymph node dimension.
## Includes extension of tumor to capsule of liver and spleen without parenchymal involvement of either organ.
### Parenchymal metastases are stage IVB. Disease invading through the bowel wall and into the mucosa increases the stage to IVB, and transmural involvement of a visceral structure also represents stage IVB disease.

+ Additional Pathologic Findings (select all that apply) (Note L)
+ ___ None identified
+ ___ Serous tubal intraepithelial carcinoma (STIC)
+ ___ Endometriosis
+ ___ Endosalpingiosis
+ ___ Other (specify): ______________________________

+ Ancillary Studies (Note M)
+ BRCA1 mutation testing (specify result): _____________
+ BRCA2 mutation testing (specify result): _____________
+ DNA mismatch repair enzyme studies (specify result)#: ______________________________
  # Note: For clear cell and endometroid carcinomas only

+ Clinical History (select all that apply)
+ ___ BRCA1/2 family history
+ ___ Hereditary breast/ovarian cancer
+ ___ Lynch syndrome
+ ___ Other (specify): ______________________________

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Explanatory Notes

A. Suggestions for Sampling for Microscopic Examination

Ovarian Surface
Involvement of the ovarian surface is an important element in staging tumors limited to the ovary, and the presence of surface involvement may influence treatment. Therefore, careful examination of the ovarian surface is crucial. Furthermore, in patients who undergo prophylactic (salpingo-) oophorectomy because of a family history of ovarian and/or breast cancer, very small foci of involvement of the ovarian surface may be present that may be potentially lethal and may be missed if the macroscopic inspection is not optimal.1-6

Ovarian/Adnexal Tumor
One section for each centimeter of the tumor's largest dimension is generally recommended, with modification based on the degree of heterogeneity of the tumor and the difficulty of diagnosis. Borderline (atypical proliferative) serous tumor, borderline serous tumors with micropapillary features/noninvasive low-grade serous carcinoma, and borderline (atypical proliferative) mucinous tumors require more sections (2 sections for each centimeter of the tumor's largest dimension is recommended in such cases). Some sections should include the ovarian surface where it is most closely approached by tumor on gross examination, with the number of sections depending on the degree of suspicion of surface involvement. Tumor adhesions and sites of rupture should be sampled and labeled specifically for microscopic identification.

Risk Reducing Salpingo-Oophorectomy Specimens
The ovary and fallopian tube should be submitted in toto in patients with BRCA mutations or suspected to be at increased risk of hereditary breast/ovarian cancer, even when grossly normal. This detailed examination results in an approximately 4-fold increase in detection of precursor lesions or early microscopic carcinoma.7 Appropriate handling implies that all ovarian and tubal tissue should be serially sectioned and submitted.8,9 For fallopian tubes, amputate the fimbriated ends and section parallel to the long axis of the fallopian tube to maximize the amount of tubal epithelium available for histological examination (SEE-FIM protocol)10 (Figure 1). The remainder of the fallopian tube is submitted as serial cross-sections. Fixation for 1 to 2 hours prior to sectioning and/or manipulation may help prevent sloughing of the epithelium.

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Figure 1. Protocol for Sectioning and Extensively Examining the Fimbriated End (SEE-FIM) of the Fallopian Tube.
This protocol entails amputation and longitudinal sectioning of the infundibulum and fimbrial segment (distal 2 cm) to allow maximal exposure of the tubal plicae. The isthmus and ampulla are cut transversely at 2- to 3-mm intervals. From Crum et al.10 Copyright © 2007 Lippincott Williams & Wilkins. Reproduced with permission.

Sampling Issues
The recommendation for the number of sections to be taken of an ovarian/adnexal tumor is a general guideline, with the pathologist determining how many sections are necessary. If a tumor is obviously malignant and
homogeneous throughout on gross examination, fewer sections may be needed. In contrast, if there is great variability in the gross appearance of the sectioned surfaces or opened cysts, it may be necessary to take more sections to sample the tumor adequately. In addition, as a general recommendation, borderline serous tumors with micropapillary foci or with microinvasion should be extensively sampled to ensure adequate assessment of the extent of invasion, when present. Mucinous tumors (particularly those with solid areas), solid teratomas, and malignant germ cell tumors often require careful gross examination and judicious sampling. Of note, additional sampling of a tumor that poses problems in differential diagnosis may be more informative than special studies.

**Fallopian Tube(s)**
For patients with high-grade serous carcinoma, if no gross lesion is present in the fimbrial end of each fallopian tube, complete microscopic examination is recommended. If a gross fimbrial lesion is present, representative sections of tumor to determine its distribution and relationship to tubal epithelium are recommended.

For patients with high-grade serous carcinoma, in contrast to other tumor histologic types covered by this protocol, a small, sometimes microscopic focus of tumor may be present in the mucosa of the fallopian tube that is the probable primary site (see Note C). The identification of tubal involvement can usually be accomplished by careful macroscopic examination and, if nothing is identified grossly, by submitting the fimbrial end of the fallopian tubes in toto for microscopic examination using the SEE-FIM protocol.10

**Uterus**
If tumor is grossly present, sections should be taken to determine its extent, including depth of invasion of myometrium if tumor possibly originated in endometrium, and to determine its relation to ovarian tumor (metastatic to, metastatic from, independent primary). If uterine serosa is grossly involved, sections to show this should be taken.

**Omentum**
If tumor is grossly identifiable, representative sections are enough. It is recommended to take multiple sections when no tumor is detected grossly. Although there is no general consensus regarding the number of sections that should be taken on a grossly normal omentum of a patient with an ovarian serous borderline tumor, serous carcinoma, or immature teratoma, a general recommendation would be to take 5 to 10 sections. Implants in serous borderline tumors and immature teratomas may vary from noninvasive to invasive low-grade serous carcinoma11 and from mature to immature,12 respectively. Identification of invasive carcinoma or an immature implant may considerably alter the prognosis and therapy. For borderline tumors or immature teratoma with grossly apparent implants, multiple sections of the implants should be taken.

For patients who have received neoadjuvant chemotherapy for advanced stage tubo-ovarian carcinoma (typically of high-grade serous type), 4 to 6 sections of omentum, to sample the most abnormal areas, are recommended to allow assessment of response to chemotherapy (see Note J).

**Lymph Nodes**
If the lymph nodes are grossly involved by tumor, representative sections are enough. However, if the lymph nodes appear grossly free of tumor, they should be entirely submitted. In either case, the dimension of the largest metastatic deposit should be documented.

**Other Staging Biopsy Specimens**
Staging biopsy tissues should be entirely processed unless grossly positive for tumor. If tumor is grossly seen, representative sections are usually sufficient. For borderline tumors or immature teratomas with grossly apparent implants, multiple sections of the implants should be taken (as in omental sampling).

**Other Organ or Tissue Removed**
Sections should be taken to determine the presence or absence, as well as location and extent, of tumor, if present. Resection margins should be taken, if applicable.
References

B. Rupture of Tumor
It is important to know if the tumor is intact or ruptured, because in the latter situation, malignant cells may have spilled into the abdominal cavity. In tumors that have an admixture of benign, borderline, and/or malignant areas, it may also be important to know which area ruptured.1,2

References

C. Site of Origin
Although determination of primary site for most histologic types of tumor is relatively straightforward, as they present with tumor confined to the ovary, when a tumor involves ovary, fallopian tube, uterus, and multiple intraperitoneal sites, it may be difficult or impossible to determine the primary site of the tumor.

Although historically primary site was assigned based on the dominant mass, this resulted in ovarian metastases from a number of extra-ovarian primary sites (eg, stomach, vermiform appendix, colon, endocervix, endometrium) being mistaken for primary ovarian neoplasms. Increased awareness of the ability of small extra-ovarian primary tumors to metastasize to the ovary and their characteristic morphological features, and the introduction of immunostains that aid in primary site determination, have led to improved recognition of ovarian metastases in practice.

There remain challenges in assignment of primary site in cases of advanced stage high-grade serous carcinoma. Table 1 reflects current recommendations for site assignment in such cases.
Table 1. Criteria for Assignment of Primary Site in Tubo-Ovarian High-Grade Serous Carcinoma (HGSC)\(^1\)-\(^3\)

<table>
<thead>
<tr>
<th>Criteria</th>
<th>Primary Site</th>
<th>Comment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Serous tubal intraepithelial carcinoma (STIC) present</td>
<td>Fallopian tube</td>
<td>Regardless of presence and size of ovarian and peritoneal disease.</td>
</tr>
<tr>
<td>Invasive mucosal carcinoma in tube, with or without STIC</td>
<td>Fallopian tube</td>
<td>Regardless of presence and size of ovarian and peritoneal disease.</td>
</tr>
<tr>
<td>Fallopian tube partially or entirely incorporated into tubo-ovarian mass</td>
<td>Fallopian tube</td>
<td>Regardless of presence and size of ovarian and peritoneal disease.</td>
</tr>
<tr>
<td>No STIC or invasive mucosal carcinoma in either tube in presence of ovarian mass or microscopic ovarian involvement</td>
<td>Ovary</td>
<td>Both tubes should be clearly visible and fully examined by a standardized SEE-FIM protocol. Regardless of presence and size of peritoneal disease.</td>
</tr>
<tr>
<td>Both tubes and both ovaries grossly and microscopically normal (when examined entirely) or involved by benign process in presence of peritoneal HGSC</td>
<td>Primary peritoneal HGSC</td>
<td>As recommended in World Health Organization (WHO) 2014 classification. This diagnosis should only be made in specimens removed at primary surgery prior to any chemotherapy; see below for samples following chemotherapy.</td>
</tr>
<tr>
<td>HGSC diagnosed on small sample, peritoneal/omental biopsy or cytology</td>
<td>Tubo-ovarian</td>
<td>Note: this should be supported by clinicopathological findings including immunohistochemistry to exclude mimics, principally uterine serous carcinoma</td>
</tr>
<tr>
<td>Post-chemotherapy with residual disease</td>
<td>Same criteria as described above</td>
<td></td>
</tr>
<tr>
<td>Post-chemotherapy with no residual disease</td>
<td>Tubo-ovarian</td>
<td></td>
</tr>
</tbody>
</table>

Site assignment as “undesignated” should be avoided as far as possible and used only in the rare event that a case does not fit into any of the above categories and/or there remains doubt over whether it is of tubo-ovarian or endometrial origin.

An adenocarcinoma is primary in the peritoneum when the ovaries and fallopian tubes are not involved or are involved only with minimal surface/serosal implants.

It is important to note that serous carcinomas of endometrium may present with adnexal mass(es). In such cases there is often not the extensive omental involvement characteristic of primary tubo-ovarian high-grade serous carcinoma. Within the endometrium, there may be a co-existent precursor lesion (in situ serous carcinoma, serous endometrial intraepithelial carcinoma), supporting primary endometrial origin of the tumor. WT-1 staining is typically strong and diffuse in tubo-ovarian high-grade serous carcinoma and weak/focal or negative in endometrial serous carcinoma. However, WT-1 is not completely sensitive or specific in determining primary site.\(^1\)\(^2\) Further study is needed to improve the ability to distinguish between high-grade serous carcinoma of endometrial and tubo-ovarian origin; however, it is likely that most instances where high-grade serous carcinoma involves the endometrium, the tumor is primary endometrial serous carcinoma.

References

D. Tumor Location
Distribution of tumor in the ovary may be a clue to its origin. If the tumor is mainly present on the surface of the ovary without forming a discrete lesion, the tumor is more likely to be secondary ovarian involvement. If a tumor is centered or mainly involves the ovarian hilus, it is most likely to be a metastasis. In the case of mucinous neoplasms, if they are bilateral or associated with mucinous ascites or peritoneal/ovarian surface involvement, they are more likely to be metastatic.\(^1\)\(^2\)
Contralateral Ovary

Contralateral ovary refers to the ovary that is non-dominant because it is either (1) involved by a tumor that is similar to but smaller than the dominant ovarian tumor, (2) contains only what appears to be metastatic tumor on gross examination, or (3) is negative for tumor. If the contralateral ovary contains only focal tumor, the gross and microscopic examination should concentrate on determining whether the tumor is an independent primary or it is metastatic from the dominant ovary. Metastatic involvement is supported by the same criteria that are used to distinguish primary and metastatic cancers to the ovary (multiple nodules, surface implants, and hilar vascular space invasion favor metastasis).

Histologic Type

It is recommended that the World Health Organization (WHO) classification and nomenclature of ovarian tumors be used because of its wide acceptance. Histologic type of ovarian carcinoma can be diagnosed with a high degree of reproducibility in routine practice and does have clinical implications. For example, hereditary breast and ovarian cancer syndrome is associated with high-grade serous carcinoma, while Lynch syndrome is associated with endometrioid and clear cell tumors (both tumors that can be seen in association with endometriosis), so accurate diagnosis is important.

The distinction between high-grade serous carcinoma and low-grade serous carcinoma is not an assignment of grade based on a continuum. They differ with respect to risk factors, precursor lesions, response to chemotherapy, and genetic events during oncogenesis, and merit consideration as distinct histologic types. The criteria for distinguishing between high-grade serous carcinoma and low-grade serous carcinoma are primarily based on nuclear variability (>3-fold nuclear size variation). In cases where the distinction is difficult, p16 and p53 immunostaining and assessment of mitotic activity (>12 mitoses/10 high-power fields) can be used. Such a system has molecular and prognostic validity and excellent inter-observer agreement.

High-grade tumors with ambiguous features, such that 1 of the specific histologic types listed cannot be diagnosed, should be classified as “carcinoma, subtype cannot be determined”; however, this is a very infrequent situation, and every effort should be made to subclassify such tumors.

Serous tubal intraepithelial carcinoma (STIC) is an unusual entity. Although an “in situ” neoplasm, it has malignant potential to spread throughout the peritoneal cavity. Therefore, with cases of only a STIC as a primary site and negative staging and negative peritoneal washing, it is recommended to stage such cases as an AJCC pT1a/FIGO IA tumor.

References


Mixtures of Histologic Types of Tumors

The term mixed carcinoma should only be used when 2 or more distinctive subtypes of surface epithelial carcinomas are identified. When a carcinoma is classified as “mixed,” the major and minor types and their relative proportions should be specified.

The diagnosis of mixed carcinoma was relatively common in the past, but with application of current histopathologic criteria, fewer than 1% of tubo-ovarian carcinomas are mixed, and the most common admixture is...
of endometrioid and clear cell carcinoma.\textsuperscript{17} It is now appreciated that high-grade serous carcinomas show a wide range of histopathologic features, and glandular (pseudoendometrioid) differentiation, solid architecture, transitional growth pattern, or clear cell change are now accepted as being within the spectrum of high-grade serous carcinoma, and the presence of these variants does not warrant diagnosis as mixed carcinoma.\textsuperscript{1,2} Therefore, a mixed carcinoma should only be used when there are 2 or more distinct and separate histologic types in the tumor.

Quantitation of various epithelial cell types within a carcinoma, as well as quantitation of tumor types within primitive germ cell tumors, may be prognostically important.\textsuperscript{3}

References

H. Histologic Grade

Epithelial Carcinomas

Clear cell carcinoma and carcinosarcomas are not graded; at present there is no grading system that has consistently been shown to prognosticate for these histologic types. Serous carcinomas are stratified into low grade and high grade. Endometrioid carcinomas may be graded according to the FIGO system used for endometrioid carcinomas of the endometrium, as shown below.

- **Grade 1**: $\leq$5% of nonsquamous, solid growth
- **Grade 2**: 6% to 50% of nonsquamous solid growth
- **Grade 3**: $>50\%$ of nonsquamous, solid growth

Notable nuclear atypia, inappropriate for the architectural grade, raises the grade of a grade 1 or grade 2 tumor by 1 grade.

There are no defined grading systems in widespread use for the remaining histologic types of ovarian carcinoma (eg, mucinous), and a gestalt 3-tier grading system can be used, acknowledging that it is not well validated.

- **Grade X**: Cannot be assessed
- **Grade 1**: Well differentiated
- **Grade 2**: Moderately differentiated
- **Grade 3**: Poorly differentiated (tumors with minimal differentiation seen in very small foci)

Seromucinous tumors are tumors with more than one Müllerian epithelial cell type. Although most commonly seen cell types are serous and mucinous, on occasion clear cell, transitional, or squamous epithelium can be seen. Such tumors can be classified as by the 2-tier (almost always low grade) or the 3-tier grading systems listed above.

Germ Cell Tumors

Immature teratomas are the only malignant germ cell tumors that are graded. They are classically graded on the basis of the quantity of immature/embryonal elements (almost always neuroectodermal tissue) that are present.\textsuperscript{1} Even though in the past a 3-tier system was used to classify immature teratomas (G1 = immature neural tissue occupying $<1$ low-power field [40X] in any slide, and G3 = immature neural tissue occupying $\geq4$ low-power fields in any slide), a 2-tiered grading system (low versus high grade) has been proposed by some experts.\textsuperscript{2} Grade 1 tumors are considered low grade while grade 2 and grade 3 tumors are considered high grade. Also, implants associated with immature teratomas must be assessed for the presence of immature elements, typically glial tissue.

Sertoli-Leydig Cell Tumors
Sertoli-Leydig cell tumors are graded in a 3-part grading system, as described in the WHO 2014 classification. Briefly, in well-differentiated (grade 1) tumors, the Sertoli cells are present in open or closed tubules; in moderately differentiated (grade 2) tumors, the Sertoli cells are present in lobular aggregates, although there may be some tubular architecture present; and in poorly differentiated (grade 3) tumors, there is sarcomatous sheets of stroma; the lobulated Sertoliform growth typical of (grade 2) tumors, if present, is only focal.

References

I. Implants (Serous/Seromucinous Borderline Tumors Only)

In both serous borderline and seromucinous borderline tumors, peritoneal implants must be assessed for invasiveness. Noninvasive implants can be subdivided into epithelial and desmoplastic types and both are typically associated with favorable prognosis. Distinction between subtypes of noninvasive implants is academic and of no clinical significance.

Note that implants with invasive carcinoma (formerly designated as “invasive implants,” as per Bell and Scully criteria) result in a diagnosis of low-grade serous carcinoma or seromucinous carcinoma, based on the WHO 2014 classification, as they are associated with a poor prognosis (identical to that of low-grade serous carcinomas).

References

J. Chemotherapy Response Score

A system for histopathologic assessment of response to neoadjuvant chemotherapy (chemotherapy response score or CRS) for high-grade serous carcinoma has been developed and validated, and shown to be highly reproducible. This 3-tiered scoring system is based on assessment of the section of omentum that shows the least response to chemotherapy. The criteria are shown in Table 2.

Table 2. Criteria of the Chemotherapy Response Score

<table>
<thead>
<tr>
<th>CRS 1: No or minimal tumor response</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mainly viable tumor with no or minimal regression-associated fibro-inflammatory changes, limited to a few foci; cases in which it is difficult to decide between regression and tumor-associated desmoplasia or inflammatory cell infiltration</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>CRS 2: Appreciable tumor response amidst viable tumor, both readily identifiable and tumor regularly distributed</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ranging from multifocal or diffuse regression associated fibro-inflammatory changes, with viable tumor in sheets, streaks, or nodules, to extensive regression associated fibro-inflammatory changes with multifocal residual tumor which is easily identifiable</td>
</tr>
</tbody>
</table>
CRS 3: Complete or near-complete response with no residual tumor OR minimal irregularly scattered tumor foci seen as individual cells, cell groups, or nodules up to 2 mm in maximum size

Mainly regression-associated fibro-inflammatory changes or, in rare cases, no/very little residual tumor in complete absence of any inflammatory response; advisable to record whether “no residual tumor” or “microscopic residual tumor present”

References

K. Pathologic Stage Classification
In view of the role of the pathologist in the staging of cancers, the staging system for ovarian cancer endorsed by the American Joint Committee on Cancer (AJCC) and the International Union Against Cancer (UICC), as well as the parallel system formulated by the International Federation of Gynecology and Obstetrics (FIGO), are recommended.1-4

According to AJCC/UICC convention, the designation “T” refers to a primary tumor that has not been previously treated. The symbol “p” refers to the pathologic classification of the TNM, as opposed to the clinical classification, and is based on gross and microscopic examination. pT entails a resection of the primary tumor or biopsy adequate to evaluate the highest pT category, pN entails removal of nodes adequate to validate lymph node metastasis, and pM implies microscopic examination of distant lesions. Clinical classification (cTNM) is usually carried out by the referring physician before treatment during initial evaluation of the patient or when pathologic classification is not possible.

Pathologic staging is usually performed after surgical resection of the primary tumor. Biopsies of all frequently involved sites, such as the omentum, mesentery, diaphragm, peritoneal surfaces, pelvic nodes, and para-aortic nodes, are required for ideal staging of early disease. For example, a patient can be confidently coded as stage IA (T1 N0 M0), if negative biopsies of all of the aforementioned sites are obtained to exclude microscopic metastases. Pathologic staging depends on pathologic documentation of the anatomic extent of disease, whether or not the primary tumor has been completely removed. If a biopsied tumor is not resected for any reason (eg, when technically infeasible), and if the highest T and N categories or the M1 category of the tumor can be confirmed microscopically, the criteria for pathologic classification and staging have been satisfied without total removal of the primary cancer.

TNM Descriptors
For identification of special cases of TNM or pTNM classifications, the “m” suffix and “y,” “r,” and “a” prefixes are used. Although they do not affect the stage grouping, they indicate cases needing separate analysis.

The “m” suffix indicates the presence of multiple primary tumors in a single site and is recorded in parentheses: pT(m)NM.

The “y” prefix indicates those cases in which classification is performed during or after initial multimodality therapy (ie, neoadjuvant chemotherapy, radiation therapy, or both chemotherapy and radiation therapy). The cTNM or pTNM category is identified by a “y” prefix. The ycTNM or ypTNM categorizes the extent of tumor actually present at the time of that examination. The “y” categorization is not an estimate of tumor before multimodality therapy (ie, before initiation of neoadjuvant therapy).

The “r” prefix indicates a recurrent tumor when staged after a documented disease-free interval and is identified by the “r” prefix: rTNM.
The “a” prefix designates the stage determined at autopsy: aTNM.

**N Category Considerations**

Isolated tumor cells (ITCs) are single cells or small clusters of cells not more than 0.2 mm in greatest dimension. Lymph nodes or distant sites with ITCs found by either histologic examination (eg, immunohistochemical evaluation for cytokeratin) or nonmorphological techniques (eg, flow cytometry, DNA analysis, polymerase chain reaction [PCR] amplification of a specific tumor marker) should be so identified. There is currently no guidance in the literature as to how these patients should be coded; until more data are available, they should be coded as “N0(i+)” with a comment noting how the cells were identified.

**References**


**L. Other Lesions**

The presence of endometriosis, particularly if it is in continuity with either an endometrioid or clear cell carcinoma, is an important clue as to the primary nature of the ovarian tumor.

**M. Special Studies**

Special studies including histochemical, immunohistochemical, and molecular genetic studies may be used in some cases. Evaluation for *BRCA1*/*BRCA2* testing on patients with high-grade serous carcinoma of tubal/ovarian/peritoneal origin should be performed at the discretion of genetic counselors with assessment of other risk factors. Immunohistochemical stains for DNA mismatch repair enzymes MLH1, MSH2, MSH6, and PMS2 for Lynch syndrome screening is recommended on all endometrioid and clear cell carcinomas of the ovary.1,2

**References**