Protocol for the Examination of Excision Specimens From Patients With Melanoma of the Skin

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| **Version:** Melanoma 4.2.0.0 | **Protocol Posting Date:** April 2020 |
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| **CAP Laboratory Accreditation Program Protocol Required Use Date:** May 2020 |
|  |  |
| Includes pTNM requirements from the 8th Edition, AJCC Staging Manual |

**For accreditation purposes, this protocol should be used for the following procedures AND tumor types:**

|  |  |
| --- | --- |
| **Procedure** | **Description** |
| Excision |  |
| **Tumor Type** | **Description** |
| Invasive melanoma  | Limited to melanoma of cutaneous surfaces only |

**This protocol is NOT required for accreditation purposes for the following:**

|  |
| --- |
| **Procedure** |
| Biopsy (consider Skin Melanoma Biopsy protocol)  |
| Primary resection specimen with no residual melanoma (eg, following neoadjuvant therapy) |
| Cytologic specimens |

**The following tumor types should NOT be reported using this protocol:**

|  |
| --- |
| **Tumor Type** |
| Melanoma in situ without invasive melanoma |

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**Accreditation Requirements**

This protocol can be utilized for a variety of procedures and tumor types for clinical care purposes. For accreditation purposes, only the definitive primary cancer resection specimen is required to have the core and conditional data elements reported in a synoptic format.

* Core data elements are required in reports to adequately describe appropriate malignancies. For accreditation purposes, essential data elements must be reported in all instances, even if the response is “not applicable” or “cannot be determined.”
* Conditional data elements are only required to be reported if applicable as delineated in the protocol. For instance, the total number of lymph nodes examined must be reported, but only if nodes are present in the specimen.
* Optional data elements are identified with “+” and although not required for CAP accreditation purposes, may be considered for reporting as determined by local practice standards.

The use of this protocol is not required for recurrent tumors or for metastatic tumors that are resected at a different time than the primary tumor. Use of this protocol is also not required for pathology reviews performed at a second institution (ie, secondary consultation, second opinion, or review of outside case at second institution).

**Synoptic Reporting**

All core and conditionally required data elements outlined on the surgical case summary from this cancer protocol must be displayed in synoptic report format. Synoptic format is defined as:

* Data element: followed by its answer (response), outline format without the paired "Data element: Response" format is NOT considered synoptic.
* The data element should be represented in the report as it is listed in the case summary. The response for any data element may be modified from those listed in the case summary, including “Cannot be determined” if appropriate.
* Each diagnostic parameter pair (Data element: Response) is listed on a separate line or in a tabular format to achieve visual separation. The following exceptions are allowed to be listed on one line:
	+ Anatomic site or specimen, laterality, and procedure
	+ Pathologic Stage Classification (pTNM) elements
	+ Negative margins, as long as all negative margins are specifically enumerated where applicable
* The synoptic portion of the report can appear in the diagnosis section of the pathology report, at the end of the report or in a separate section, but all Data element: Responses must be listed together in one location

Organizations and pathologists may choose to list the required elements in any order, use additional methods in order to enhance or achieve visual separation, or add optional items within the synoptic report. The report may have required elements in a summary format elsewhere in the report IN ADDITION TO but not as replacement for the synoptic report ie, all required elements must be in the synoptic portion of the report in the format defined above.

Summary of Changes

**Version 4.2.0.0**

**The following was modified:**

Required status changed from conditionally reported to optional for Specify location(s) under Peripheral Margins

Distance of melanoma in situ from deep margin was added as an optional element

Updated Explanatory Note G

Surgical Pathology Cancer Case Summary

Protocol posting date: April 2020

# MELANOMA OF THE SKIN: Excision, Re-Excision

**Select a single response unless otherwise indicated.**

## Procedure (select all that apply) (Note A)

\_\_\_ Excision

\_\_\_ Re-excision

\_\_\_ Sentinel node(s) biopsy

\_\_\_ Lymphadenectomy, regional nodes (specify): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

\_\_\_ Other (specify): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

\_\_\_ Not specified

## + Specimen Laterality

+\_\_\_ Right

+\_\_\_ Left

+\_\_\_ Midline

## +\_\_\_ Not specified

## + Tumor Site (Note B): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

## Macroscopic Satellite Nodule(s) (applicable to invasive tumor only)

\_\_\_ Not identified

\_\_\_ Present

\_\_\_ Cannot be determined

## Histologic Type (Note C)

\_\_\_ No residual melanoma identified

Invasive Melanoma

\_\_\_ Superficial spreading melanoma (low-cumulative sun damage (CSD) melanoma)

\_\_\_ Lentigo maligna melanoma

\_\_\_ Desmoplastic melanoma

 +\_\_\_\_ Pure desmoplastic melanoma

 +\_\_\_\_ Mixed desmoplastic melanoma

\_\_\_ Acral melanoma

\_\_\_ Melanoma arising in a blue nevus (blue nevus-like melanoma)

\_\_\_ Melanoma arising in a giant congenital nevus

\_\_\_ Spitz melanoma (malignant Spitz tumor)

\_\_\_ Nodular melanoma

\_\_\_ Nevoid melanoma

\_\_\_ Melanoma, not otherwise classified

\_\_\_ Other histologic type not listed (specify): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

Melanoma In Situ (anatomic level I)#

\_\_\_ Melanoma in situ, superficial spreading type (low-cumulative sun damage (CSD) melanoma in situ)

\_\_\_ Melanoma in situ, lentigo maligna type

\_\_\_ Acral melanoma in situ

\_\_\_ Melanoma in situ arising in a giant congenital nevus

\_\_\_ Melanoma in situ, not otherwise classified

\_\_\_ Other histologic type not listed (specify): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

# *Note: For melanoma in situ, elements that assess the invasive component are not applicable and should not be reported.*

## Maximum Tumor (Breslow) Thickness (applicable to invasive tumor only) (Note D)

Specify (millimeters): \_\_\_ mm

*or*

At least (millimeters): \_\_\_ mm (explain): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

\_\_\_ Cannot be determined (explain): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

## Ulceration (required for invasive tumor only) (Note E)

\_\_\_ Not identified

\_\_\_ Present

+ Extent of ulceration (millimeters): \_\_\_\_ mm

\_\_\_ Cannot be determined

**Microsatellite(s) (applicable to invasive tumor only) (Note F)**

\_\_\_ Not identified

\_\_\_ Present

\_\_\_ Cannot be determined

## Margins (Note G)

**Peripheral Margins*#***

\_\_\_ Negative for invasive melanoma

**+ Distance of invasive melanoma from closest peripheral margin (millimeters):**

+ \_\_\_ Specify \_\_\_ mm

+ \_\_\_ Less than \_\_\_ mm

+ \_\_\_ Greater than \_\_\_ mm

+ \_\_\_ Cannot be determined (explain): \_\_\_\_\_\_\_\_\_\_\_\_\_

 + Specify location(s): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

\_\_\_ Invasive melanoma present at margin

 + Specify location(s): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

\_\_\_ Negative for melanoma in situ

**+ Distance of melanoma in situ from closest peripheral margin (millimeters):**

+ \_\_\_ Specify \_\_\_ mm

+ \_\_\_ Less than \_\_\_ mm

+ \_\_\_ Greater than \_\_\_ mm

+ \_\_\_ Cannot be determined (explain): \_\_\_\_\_\_\_\_\_\_\_\_\_

 + Specify location(s): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

\_\_\_ Melanoma in situ present at margin

 + Specify location(s): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

\_\_\_ Cannot be assessed

### Deep Margin#

\_\_\_ Negative for invasive melanoma

**+ Distance of invasive melanoma from deep margin (millimeters):**

+ \_\_\_ Specify \_\_\_ mm

+ \_\_\_ Less than \_\_\_ mm

+ \_\_\_ Greater than \_\_\_ mm

+ \_\_\_ Cannot be determined (explain): \_\_\_\_\_\_\_\_\_\_\_\_\_

\_\_\_ Invasive melanoma present at margin

\_\_\_ Negative for melanoma in situ

\_\_\_ Melanoma in situ present at margin

**+ Distance of melanoma in situ from deep margin (millimeters):**

+ \_\_\_ Specify \_\_\_ mm

+ \_\_\_ Less than \_\_\_ mm

+ \_\_\_ Greater than \_\_\_ mm

+ \_\_\_ Cannot be determined (explain): \_\_\_\_\_\_\_\_\_\_\_\_\_

\_\_\_ Cannot be assessed

*#Note: Margin involvement by melanoma in situ should be recorded if in situ disease is present in the specimen, and if margins are uninvolved by invasive melanoma.*

## Mitotic Rate (applicable to invasive tumor only) (Note H)

\_\_\_ None identified

\_\_\_ Specify (mitoses/mm2): \_\_\_\_\_\_ mitoses/mm2

\_\_\_ Cannot be determined

**+ Anatomic (Clark) Level (applicable to invasive tumor only) (Note D)**

+ \_\_\_ At least level \_\_\_\_\_ (explain): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

+ \_\_\_ II (melanoma present in but does not fill and expand papillary dermis)

+ \_\_\_ III (melanoma fills and expands papillary dermis)

+ \_\_\_ IV (melanoma invades reticular dermis)

+ \_\_\_ V (melanoma invades subcutis)

+ \_\_\_ Cannot be determined

## Lymphovascular Invasion (applicable to invasive tumor only) (Note I)

\_\_\_ Not identified

\_\_\_ Present

\_\_\_ Cannot be determined

## Neurotropism (applicable to invasive tumor only) (Note J)

\_\_\_ Not identified

\_\_\_ Present

\_\_\_ Cannot be determined

## + Tumor-Infiltrating Lymphocytes (applicable to invasive tumor only) (Note K)

+ \_\_\_ Not identified

+ \_\_\_ Present, nonbrisk

+ \_\_\_ Present, brisk

+ \_\_\_ Cannot be determined

## Tumor Regression (Note L)

\_\_\_ Not identified

\_\_\_ Present, involving less than 75% of lesion

\_\_\_ Present, involving 75% or more of lesion

\_\_\_ Cannot be determined

## Regional Lymph Nodes (applicable to invasive tumor only) (Note M)

*Note: If nodes from more than one nodal basin are included, each nodal basin should be reported separately.*

\_\_\_ No lymph nodes submitted or found

\_\_\_ Uninvolved by tumor cells

**Total Number of Lymph Nodes Examined: \_\_\_\_**

**Number of Sentinel Nodes Examined (if applicable): \_\_\_\_**

\_\_\_ Involved by tumor cells

**Total Number of Lymph Nodes Involved: \_\_\_\_**

\_\_\_ Number cannot be determined (explain): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

+ Location (specify)*#*: \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

 *#Note: Locations may include subcapsular, intramedullary, and other locations.*

**Number of Sentinel Nodes Involved (required only if sentinel nodes examined and involved): \_\_\_\_**

\_\_\_ Number cannot be determined (explain): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

**+ Size of Largest Metastatic Deposit*#* \_\_\_\_mm**

## *# Note: Relevant only if larger than sentinel lymph node metastatic deposits.*

**+ Size of Largest Metastatic Deposit in Sentinel Lymph Node \_\_\_\_mm**

**+ Extranodal Extension**

+ \_\_\_ Not identified

+ \_\_\_ Present

+ \_\_\_ Cannot be determined

**Matted Nodes**

\_\_\_ Not identified

\_\_\_ Present

**Total Number of Lymph Nodes Examined: ­­­­­\_\_\_\_**

\_\_\_ Number cannot be determined (explain): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

**Number of Sentinel Nodes Examined (if applicable): \_\_\_\_**

## Pathologic Stage Classification (pTNM, AJCC 8th Edition) (Note N)

*Note: Reporting of pT, pN, and (when applicable) pM categories is based on information available to the pathologist at the time the report is issued. Only the applicable T, N, or M category is required for reporting; their definitions need not be included in the report. The categories (with modifiers when applicable) can be listed on 1 line or more than 1 line.*

**+ \_\_\_ Classification assigned in this report includes information from a prior procedure (explain): \_\_\_\_\_\_\_\_\_\_**

*Note: In general, CAP cancer protocol case summaries are intended to guide reporting on the specimen that the pathologist is evaluating at that time. However, melanoma cases frequently include multiple procedures. Because of this, a prior procedure that was performed may affect the pathologic classification of the tumor.*

*In order to represent this appropriately in the pathology report, information from prior procedures may be incorporated into the assignment of pathologic classification if it is available. When information from a prior procedure is included in this report, details of that procedure should be documented in the report as well.*

**TNM Descriptors (required only if applicable) (select all that apply)**

\_\_\_ m (multiple)

\_\_\_ r (recurrence or retreatment)

\_\_\_ y (posttherapy or post-neoadjuvant therapy)

### Primary Tumor (pT)

\_\_\_ pTX: Primary tumor thickness cannot be assessed (eg, diagnosis by curettage)
(explain): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

\_\_\_ pT0: No evidence of primary tumor (eg, unknown primary or completely regressed melanoma)

\_\_\_ pTis: Melanoma in situ (ie, not an invasive tumor: anatomic level I)

\_\_\_ pT1: Melanoma 1.0 mm or less in thickness, ulceration status unknown or unspecified (see Note D)

\_\_\_ pT1a: Melanoma <0.8 mm in thickness, no ulceration

\_\_\_ pT1b: Melanoma <0.8 mm in thickness with ulceration OR melanoma 0.8 to 1.0 mm in thickness with or without ulceration

\_\_\_ pT2: Melanoma >1.0 to 2.0 mm in thickness, ulceration status unknown or unspecified

\_\_\_ pT2a: Melanoma >1.0 to 2.0 mm in thickness, no ulceration

\_\_\_ pT2b: Melanoma >1.0 to 2.0 mm in thickness, with ulceration

\_\_\_ pT3: Melanoma >2.0 to 4.0 mm in thickness, ulceration status unknown or unspecified

\_\_\_ pT3a: Melanoma >2.0 to 4.0 mm in thickness, no ulceration

\_\_\_ pT3b: Melanoma >2.0 to 4.0 mm in thickness, with ulceration

\_\_\_ pT4: Melanoma >4.0 mm in thickness, ulceration status unknown or unspecified

\_\_\_ pT4a: Melanoma >4.0 mm in thickness, no ulceration

\_\_\_ pT4b: Melanoma >4.0 mm in thickness, with ulceration

### Regional Lymph Nodes (pN) (applicable to invasive tumor only)

\_\_\_ pNX: Regional lymph nodes not assessed (e.g., SLN biopsy not performed, regional nodes previously removed for another reason)

\_\_\_ pN0: No regional lymph node metastasis detected

\_\_\_ pN1: One tumor-involved node or in-transit, satellite, and/or microsatellite metastases with no tumor-involved nodes

\_\_\_ pN1a: One clinically occult tumor-involved node (ie, detected by sentinel node biopsy) with no in-transit, satellite and/or microsatellite metastases

\_\_\_ pN1b: One clinically detected tumor-involved node with no in-transit, satellite and/or microsatellite metastases***#***

\_\_\_ pN1c: Presence of in-transit, satellite and/or microsatellite metastases with no regional lymph node disease

\_\_\_ pN2: Metastasis in two to three regional nodes or in-transit, satellite, and/or microsatellite with one tumor-involved node

\_\_\_ pN2a: Two to three clinically occult tumor-involved node (ie, detected by sentinel node biopsy) with no in-transit, satellite and/or microsatellite metastases

\_\_\_ pN2b: Two to three tumor-involved nodes at least one of which was clinically detected with no in-transit, satellite and/or microsatellite metastases***#***

\_\_\_ pN2c: One clinically occult or clinically apparent tumor-involved node with presence of in-transit, satellite and/or microsatellite metastases

\_\_\_ pN3: Metastasis in four or more regional lymph nodes, or in-transit, satellite or microsatellite metastases with two or more tumor-involved nodes or any number of matted nodes without or with in-transit, satellite or microsatellite metastases

\_\_\_ pN3a: Four or more clinically occult tumor-involved nodes (ie, detected by sentinel node biopsy) with no in-transit, satellite and/or microsatellite metastases

\_\_\_ pN3b: Four or more tumor-involved nodes, at least one of which was clinically detected, with no in-transit, satellite and/or microsatellite metastases***#***

\_\_\_ pN3c: Two or more clinically occult or clinically detected tumor-involved nodes with in-transit, satellite and/or microsatellite metastases and/or any number of matted nodes with in-transit, satellite and/or microsatellite metastases

*# Note: pN1b, 2b, and 3b subcategories are dependent on clinical information that may be unavailable to the pathologist. If this information is not available, the parent category (pN1, pN2 or pN3) should be selected.*

### Distant Metastasis (pM) (required only if confirmed pathologically in this case)

*Note: AJCC pM category suffixes “(0)” and “(1)”, which denote LDH level of elevation, are NOT included in the surgical pathology report. LDH levels, as with other clinical parameters, may be included in the final classification by clinicians with access to this data.*

\_\_\_ pM1: Distant metastasis (documented in this specimen)

\_\_\_ pM1a: Distant metastasis in skin, subcutaneous tissues, soft tissues including muscle and/or nonregional lymph nodes

\_\_\_ pM1b: Distant metastasis to lung with or without M1a sites of disease

\_\_\_ pM1c: Distant metastasis to non-CNS visceral sites with or without M1a or M1b sites of disease

\_\_\_ pM1d: Distant metastasis to CNS with or without M1a, M1b or M1c sites of disease

 Specify site(s), if known: \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

## + Additional Pathologic Findings (select all that apply)

+ \_\_\_ Associated nevus (specify type): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

+ \_\_\_ Other (specify): \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

**+ Ancillary Studies**

*Note: For molecular genetic reporting, the CAP Melanoma Biomarker Template should be used. Pending biomarker studies should be listed in the Comments section of this report.*

## + Comment(s)

Explanatory Notes

## A. Procedure

Optimal pathologic evaluation of melanocytic lesions requires complete excision that incorporates the full thickness of the lesion removed intact.1 "Shave" procedures that do not include the intact base of the lesion are suboptimal for pathologic evaluation and should be avoided unless clinically indicated. Similarly, “punch” procedures may not include intact peripheral borders of the lesion thereby limiting assessment of symmetry and lateral circumscription, which can be essential for distinction of melanoma from melanocytic nevus.2,3 Partial biopsies of melanocytic tumors are associated with an increased risk of misdiagnosis with possible consequent adverse clinical outcomes.4 Nevertheless, clinical factors are also important in determining the most appropriate biopsy technique for any lesion. For example, an excision biopsy of a large lesion on a cosmetically or functionally sensitive site may cause cosmetic disfigurement or alter reconstructive options.

The use of frozen sections for evaluation of biopsy or excision of melanocytic lesions is strongly discouraged.5,6 Optimal histologic evaluation of cutaneous melanoma requires well-cut, well-stained hematoxylin-and-eosin (H&E) sections prepared from formalin-fixed paraffin-embedded tissue. Frozen sections of sentinel lymph nodes are similarly discouraged, because the manipulation required for intraoperative handling may decrease the sensitivity of the procedure.7

References

1. Sober AJ, Chuang TY, Duvic M, et al. Guidelines of care for primary cutaneous melanoma. *J Am Acad Dermatol.* 2001;45(4):579-586.

2. Stell VH, Norton HJ, Smith KS, Salo JC, White RL, Jr. Method of biopsy and incidence of positive margins in primary melanoma. *Ann Surg Oncol.* 2007;14(2):893-898.

3. Sober AJ, Balch CM. Method of biopsy and incidence of positive margins in primary melanoma. *Ann Surg Oncol.* 2007;14(2):274-275.

4. Ng JC, Swain S, Dowling JP, Wolfe R, Simpson P, Kelly JW. The impact of partial biopsy on histopathologic diagnosis of cutaneous melanoma: experience of an Australian tertiary referral service. *Arch Dermatol.* 2010;146(3):234-239.

5. Smith-Zagone MJ, Schwartz MR. Frozen section of skin specimens. *Arch Pathol Lab Med.* 2005;129(12):1536-1543.

6. Prieto VG, Argenyi ZB, Barnhill RL, et al. Are en face frozen sections accurate for diagnosing margin status in melanocytic lesions? *Am J Clin Pathol.* 2003;120(2):203-208.

7. Scolyer RA, Thompson JF, McCarthy SW, Gershenwald JE, Ross MI, Cochran AJ. Intraoperative frozen-section evaluation can reduce accuracy of pathologic assessment of sentinel nodes in melanoma patients. *J Am Coll Surg.* 2005;201(5):821-823; author reply 823-824.

## B. Anatomic Site

For cutaneous melanoma, prognosis may be affected by primary anatomic site.1,2

References

1. Balch CM, Soong SJ, Gershenwald JE, et al. Prognostic factors analysis of 17,600 melanoma patients: validation of the American Joint Committee on Cancer melanoma staging system. *J Clin Oncol.* 2001;19(16):3622-3634.

2. Elder DE, Massi D, Scolyer RA, Willemze R. eds. *WHO Classification of Skin Tumors. World Health Organization of Tumors, 4th ed Volume 11.* Lyon France; 2018, ISBN-13 978-92-832-2440-2.

## C. Melanoma Subtypes

Superficial spreading melanoma (low cumulative sun damage (CSD) melanoma)

Lentigo maligna melanoma

Desmoplastic melanoma (pure and mixed)

Acral melanoma

Melanoma arising in a blue nevus (blue nevus-like melanoma)

Melanoma arising in a giant congenital nevus

Spitz melanoma (malignant Spitz tumor)

Nodular melanoma

Nevoid melanoma

Melanoma, not otherwise classified

The recent WHO 2018 classification introduced multidimensional pathway classification of melanocytic tumors based on the extent of ultraviolet (UV) radiation damage, the cell of origin, and characteristic genomic findings (Table 1).

Table 1. Classification of melanoma

|  |  |
| --- | --- |
| **Extent of UV radiation damage** | **Subtypes**  |
| Melanomas found in skin with low cumulative sun damage (low-CSD) | Superficial spreading melanoma (low cumulative sun damage (CSD) melanoma) |
| Melanomas found in skin with high cumulative sun damage (high-CSD) | Lentigo maligna melanoma |
| Desmoplastic melanoma |
| Melanomas on site with no sun exposure or without known etiological associations with sun exposure | Malignant Spitz tumor (Spitz melanoma) |
| Acral melanoma |
| Mucosal melanoma |
| Melanoma arising in congenital nevus |
| Melanoma arising in a blue nevus |
| Uveal melanoma  |
| Various sun exposure | Nodular melanoma |
| Nevoid melanoma  |

References

1. Elder DE, Massi D, Scolyer RA, Willemze R. eds. WHO Classification of Skin Tumors. World Health Organization of Tumors, 4th ed Volume 11. Lyon France; 2018, ISBN-13 978-92-832-2440-2.

## D. Primary Tumor (Breslow) Thickness) and Anatomic (Clark) Levels

## Maximum tumor thickness is measured with a calibrated ocular micrometer at a right angle to the adjacent normal skin. The upper point of reference is the upper edge of the granular layer of the epidermis of the overlying skin or, if the lesion is ulcerated overlying the entire dermal component, the base of the ulcer. The lower reference point is the deepest point of tumor invasion (ie, the leading edge of a single mass or an isolated group of cells deep to the main mass).

If the tumor is transected by the deep margin of the specimen, the depth may be indicated as “at least \_\_ mm” with a comment explaining the limitation of thickness assessment. For example, “The maximum tumor thickness cannot be determined in this specimen because the deep plane of the biopsy transects the tumor.” Tumor thickness measurements should not be based on periadnexal extension (either periadnexal adventitial or extra-adventitial extension), except when it is the only focus of invasion. In that circumstance, Breslow thickness may be measured from the inner layer of the outer root sheath epithelium or inner luminal surface of sweat glands, to the furthest extent of infiltration into the periadnexal dermis. Microsatellites or foci of neurotropism or lymphovascular invasion should not be included in tumor thickness measurements.

In the 8th edition of the AJCC melanoma staging system,1 it is recommended that tumor thickness measurements be recorded to the nearest 0.1 mm, not the nearest 0.01 mm, because of the impracticality and imprecision of measurements, particularly for tumors greater than 1 mm thick. Tumors less than or equal to 1 mm thick may be measured to the nearest 0.01 mm if practical, but should be reported to the nearest 0.1mm (eg, melanomas measured to be in the range of 0.75 mm to 0.84 mm are reported as 0.8 mm in thickness and hence T1b, and tumors 1.01 to 1.04 mm in thickness are reported as 1.0 mm).

While the principal T category tumor thickness ranges have been maintained in the AJCC 8th edition, T1 is now subcategorized by tumor thickness strata at a 0.8 mm threshold. Tumor mitotic rate as a dichotomous variable is no longer used as a staging category criterion for T1 tumors. T1a melanomas are now defined as non-ulcerated and less than 0.8 mm in thickness. T1b melanomas are defined as 0.8-1.0mm in thickness or ulcerated melanomas less than 0.8 mm in thickness.

Anatomic (Clark) levels are defined as follows:

I Intraepidermal tumor only (ie, melanoma in situ)

II Tumor present in but does not fill and expand papillary dermis

III Tumor fills and expands papillary dermis

IV Tumor invades into reticular dermis

V Tumor invades subcutis

Anatomic (Clark) level of invasion remains an independent predictor of outcome and is recommended by the AJCC to be reported as a primary tumor characteristic.1 However, assessment of Clark levels is less reproducible among pathologists than is tumor thickness, and Clark levels are not used in the AJCC staging system for pT status. Accordingly, Clark levels are included in this checklist as an optional data item.

References

1. Gershenwald JE, Scolyer RA, Hess KR, et al. Melanoma of the skin, In: Amin MB, Edge SB, Greene FL, et al. eds. *AJCC Cancer Staging Manual.* 8th ed. New York, NY: Springer; 2017.

## E. Ulceration

Primary tumor ulceration has been shown to be a dominant independent prognostic factor in invasive cutaneous melanoma,1,2 and if present, changes the pT stage from T1a to T1b, T2a to T2b, etc., depending on the thickness of the tumor. The presence or absence of ulceration must be confirmed on microscopic examination.2 Melanoma ulceration is defined as the combination of the following features: full-thickness epidermal defect (including absence of stratum corneum and basement membrane); evidence of reactive changes (ie, fibrin deposition, neutrophils); and thinning, effacement, or reactive hyperplasia of the surrounding epidermis in the absence of trauma or a recent surgical procedure. Ulcerated melanomas typically show invasion through the epidermis, whereas nonulcerated melanomas tend to lift the overlying epidermis.

Only nontraumatic (“tumorigenic”) ulceration should be recorded as ulceration. If ulceration is present related to a prior biopsy, the tumor should not be recorded as ulcerated for staging purposes. If a lesion has been recently biopsied or there is only focal loss of the epidermis, assessment of ulceration may be difficult or impossible; in this instance it may be difficult to determine whether the epidermal deficiency is due to true ulceration or to sectioning artifact.2 Absence of fibrin, neutrophils, or granulation tissue from putative areas of ulceration would be clues that the apparent ulceration is actually due to sectioning of only part of the epidermis and this should not be designated as ulceration. If non-traumatic (“tumorigenic”) ulceration is present in either an initial partial biopsy or a re-excision specimen, then for staging purposes, the tumor should be recorded as ulcerated.

Ulceration may be present in an in situ melanoma but does not affect the staging.

A number of studies have demonstrated that the extent of ulceration (measured either as a percentage of the width of the dermal invasive component of the tumor or as a diameter) more accurately predicts outcome than the presence or absence of ulceration alone.3,4

References

1. Gershenwald JE, Scolyer RA, Hess KR, Sondak VK, Long GV, Ross MI et al. Melanoma staging: Evidence-based changes in the American Joint Committee on Cancer eighth edition cancer staging manual. CA Cancer J Clin. 2017;67(6):472-92.

2. Gershenwald JE, Scolyer RA, Hess KR, et al. Melanoma of the skin, In: Amin MB, Edge SB, Greene FL, et al. eds. *AJCC Cancer Staging Manual.* 8th ed. New York, NY: Springer; 2017.

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**F. Microsatellite(s)**

A microsatellite(s) is defined as the presence of a microscopic cutaneous metastasis found adjacent or deep to a primary melanoma on pathological examination of the primary tumor site.1 The metastatic tumor cells must be discontinuous from the primary tumor. If the tissue between the apparently separate nodule and the primary tumor is only fibrous scarring and/or inflammation, this does not indicate a microsatellite, because the aforementioned changes may represent regression of the intervening tumor (but not separated only by fibrosis or inflammation because the features could signify regression of the intervening tumor). There is no minimum size threshold or distance from the primary tumor to define a microsatellite. Before diagnosing the presence of a microsatellite, it is generally recommended that multiple sections from the same tissue block being examined to verify that the microsatellite is indeed discontinuous from the primary tumor. For example, periadnexal extension of tumor or the irregular shape of the peripheral or deep extent of the tumor may result in tumor that is contiguous with the primary tumor appear discontiguous on single sections.

References

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## G. Margins

Microscopically measured distances between tumor and labeled peripheral (lateral) or deep margins are appropriately recorded for melanoma excision specimens, whenever possible.If a margin is involved by tumor, it should be stated whether the tumor is in situ or invasive. Occasionally, in situ melanoma can extend down an adnexal structure like a hair follicle and cause a deep positive margin.1

References

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## H. Mitotic Rate

Tumor mitotic rate (of the invasive component of a melanoma) is a strong independent predictor of outcome across its dynamic range in all thickness categories and should be assessed and recorded in all primary melanomas including in both initial and excision biopsies (the highest value in any specimen should be used for prognostic purposes). Although tumor mitotic rate is no longer used as a T1-category criterion in the 8th edition of the AJCC melanoma staging system (due to the more significant prognostic significance of the new tumor thickness strata within T1 melanoma), mitotic rate will likely be an important parameter in prognostic models developed in the future that will provide personalized prediction of prognosis for individual patients1. The method recommended for enumerating the tumor mitotic rate in the 8th edition of the AJCC staging system is provided below:

“The recommended approach to enumerating mitoses is to first find the regions in the dermis containing the most mitotic figures, the so-called "hot spot" or "dermal hot spot." After counting the mitoses in the initial high‐power field, the count is extended to immediately adjacent nonoverlapping fields until an area of tissue corresponding to 1 mm2 is assessed. If no hot spot is found and mitoses are sparse and/or randomly scattered throughout the lesion, then a representative mitosis is chosen and, beginning with that field, the count is then extended to immediately adjacent nonoverlapping fields until an area corresponding to 1 mm2 of tissue is assessed. The count then is expressed as the (whole) number of mitoses/mm2. If the invasive component of the tumor involves an area less than 1 mm2, the number of mitoses should be assessed and recorded as if they were found within square millimeter. For example, if the entire dermal component of a tumor occupies 0.5 mm2 and only one mitosis is identified, the mitotic rate should be recorded as 1/mm2 (not 2/mm2). The number of mitoses should be listed as a whole number per square millimeter. If no mitoses are identified, the mitotic rate may be recorded as “none identified” or “0/mm2.” Only mitotic figures in *invasive* melanoma cells should be counted. This methodology for determining the mitotic rate of a melanoma has been shown to have excellent interobserver reproducibility, including among pathologists with widely differing experience in the assessment of melanocytic tumors.2

To obtain accurate measurement, calibration of individual microscopes is recommended using a stage micrometer to determine the number of high-power fields that equates to a square millimeter.

The data that demonstrated the strong prognostic significance of mitotic rate were obtained from the melanoma pathology reports of routinely assessed H&E stained sections. It therefore is recommended that no additional sections be cut and examined in excess of those that would normally be used to report and diagnose the melanoma to determine the mitotic count (ie, no additional sections should be cut and examined for the sole purpose of determining the mitotic rate, including in situations in which no mitoses are identified on the initial, routinely examined sections). Immunohistochemical stains for identifying mitoses are not used for determining mitotic rate for staging and/ or reporting purposes. A possible exception is the use to dual immunohistochemistry (eg, MART1 and pHH3) to determine if a cell in mitosis is a melanocyte or not (macrophage, endothelial cell, etc).3

Although the AJCC recommends reporting “0” rather than “none identified” or “less than 1,” for the purposes of cancer registry reporting all of these terms should be considered equivalent.

References

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## I. Lymphovascular Invasion

Lymphovascular invasion is identified by the demonstration of melanoma cells within the lumina of blood vessels or lymphatics, or both.1 Immunohistochemistry for vascular endothelial cell markers CD31, CD34 or ERG or the lymphatic marker D2-40 may assist in the identification of the presence of intravascular or intralymphatic tumor by highlighting vascular lumina. Vascular invasion by melanoma correlates independently with worsened overall survival.2 The detection of LVI is increased in primary melanomas when double labeling of tumor cells and lymphatic endothelium is applied.3

References

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**J. Neurotropism**

Neurotropism is defined as the presence of melanoma cells abutting nerve sheaths usually circumferentially (perineural invasion) or within nerves (intraneural invasion).1 Occasionally, the tumor itself may form neuroid structures (termed ‘neural transformation’ and this is also regarded as neurotropism). Neurotropism is best identified at the periphery of the tumor; the presence of melanoma cells around nerves in the main tumor mass caused by “entrapment” of nerves in the expanding tumor does not represent neurotropism.

Neurotropism is most commonly identified in desmoplastic melanomas (sometimes termed desmoplastic neurotropic melanoma) but may occur in any melanoma subtype.2 Neurotropism may correlate with an increased risk for local recurrence.

References

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## K. Tumor-Infiltrating Lymphocytes

A paucity of tumor-infiltrating lymphocytes (TILs) is an adverse prognostic factor for cutaneous melanoma.1 Tumor-infiltrating lymphocytes may be assessed in a semiquantitative way, as defined below. To qualify as TILs, lymphocytes need to surround and disrupt tumor cells of the invasive component of the tumor.

TILs Not Identified: No lymphocytes present, or lymphocytes present but do not infiltrate tumor at all.

TILs Nonbrisk: Lymphocytes infiltrate melanoma only focally or not along the entire base of the invasive tumor.

TILs Brisk: Lymphocytes diffusely infiltrate the entire base of the invasive tumor (Figure 1, A) or show diffuse permeation of the invasive tumor (Figure 1, B).

References

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**Figure 1.** Brisk tumor-infiltrating lymphocytes. A. Lymphocytes diffusely infiltrate the entire base of the invasive tumor. B. Lymphocytes diffusely infiltrate the entire invasive component of the melanoma.

## L. Tumor Regression

Characteristic features of regression include replacement of tumor cells by lymphocytic inflammation, as well as attenuation of the epidermis and nonlaminated dermal fibrosis with inflammatory cells, melanophagocytosis, and telangiectasia.

## M. Regional Lymph Nodes

Removal of sentinel lymph nodes may be performed for patients with clinically localized primary cutaneous melanomas with a thickness of 1 mm or greater, or in selected patients with thinner tumors with other adverse prognostic features.1,2 Frozen section analysis of sentinel lymph nodes is not advised.1 Review of the H&E-stained slides from multiple levels through serially sliced sentinel lymph nodes increases the sensitivity of detecting microscopic melanoma metastasis; routine analysis (H&E-stained sections of the cut surfaces of a simply bisected lymph node) may lead to a false-negative rate of 10%-15%. The use of immunohistochemical stains (eg, for HMB-45 or MART-1) further increases the sensitivity of detection of microscopic melanoma metastases and should also be considered in the examination of sentinel lymph nodes. Although immunohistochemical staining should be used in conjunction with and not in place of standard H&E histologic examination, immunohistochemically identified micrometastases are accepted as representing greater than N0 disease by the 8th edition of the AJCC staging system (as in the 7th edition), ie, a lymph node in which any metastatic tumors cells are identified, irrespective of the number of cells or whether they were identified on H&E or immunostained sections, should be designated as a tumor-positive node.2

For histologic examination, whether for sentinel node analysis or for routine regional lymph node evaluation, the entire node, except tissue collected for consented research protocols, should be submitted. For routine evaluation, large lymph nodes may be bisected or sliced at 2-3 mm intervals, whereas smaller nodes (less than 5 mm) may be submitted whole.

Data from multiple studies3-5 indicated that the sentinel lymph node tumor burden and/or the microanatomical region/compartment of the sentinel node occupied by the metastasis may be useful in predicting patients who have additional disease in nonsentinel nodes as well as disease outcome. Because sentinel node tumor burden is considered a regional disease prognostic factor, it should be reported in all patients with a positive sentinel node but it is not used to determine N-category groupings in the 8th edition of the AJCC staging system. The current National Comprehensive Cancer Network (NCCN) guidelines6 also recommend recording the size and location of tumor present in a positive sentinel node.

References

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## N. Pathologic Stage Classification

Changes in the 8th edition *AJCC Cancer Staging Manual1* of importance to practicing pathologists include:

* T1a melanoma now defined as non-ulcerated melanomas less than 0.8mm thick
* T1b melanomas now defined as melanoma between 0.8mm and 1.0mm in thickness OR ulcerated melanomas less than 0.8mm thick
* Tumor mitotic rate no longer used as a T category criterion but remains an important prognostic factor and should be reported in all invasive primary melanomas
* Recommendation to record tumor thickness to the nearest 0.1mm (not the nearest 0.01mm)
* Regarding regional lymph node metastasis, the previously empirically defined terms “microscopic” and “macroscopic” replaced with “clinically occult” (ie, detected by sentinel node biopsy) and “clinically detected”
* Non-nodal regional metastatic disease (ie, microsatellites, satellites and in transit metastases) now formally stratified by N category according to the number of tumor-involved nodes
* Gross extranodal extension no longer used as an N-category criterion (but presence of “matted nodes” retained)
* M1 now defined by both anatomic site of distant metastasis and serum LDH levels for all anatomic subsite categories of metastasis
* New M1d designation added for distant metastasis to central nervous system
* pT1bN0M0 is now pathologic stage IA in contrast to cT1N0M0 which remains clinical stage IB disease
* N category now defines four stage subgroups and takes into account both T category elements and N category elements

Pathologic staging includes microstaging of the primary melanoma and pathologic information about the regional lymph nodes after partial or complete lymphadenectomy.

In virtually all studies of cutaneous melanoma, tumor thickness has been shown to be a dominant prognostic factor,and it forms the basis for the stratification of pT. Although anatomic (Clark) levels are also commonly used to indicate depth of invasion of the primary tumor, they are less predictive of clinical outcome than mitotic activity or ulceration. 1,2,3

By AJCC convention, the designation “T” refers to a primary tumor that has not been previously treated. Similarly by convention, clinical staging is performed after biopsy of the primary melanoma (including utilizing pathologic information on microstaging of the primary melanoma) with clinical or biopsy assessment of regional lymph nodes and distant sites. Pathologic staging uses information gained from pathologic evaluation of both the primary melanoma after biopsy *and* wide excision as well as pathological evaluation of the regional node basin after SLN biopsy (required for N categorization of all greater than T1 melanomas) and/or complete regional lymphadenectomy.1,2 In additional, for pathological staging, if information from any prior biopsy is known and is relevant for staging, this should be documented in the pathology report (in the staging section) and used for assigning T, N and M categories and staging purposes.

**T Category Considerations**

Pathologic (microscopic) assessment of the primary tumor is required for accurate staging. Therefore, excision of the primary tumor, rather than incisional biopsy, is advised. The T classification of melanoma is based on the thickness of the primary tumor and presence or absence of ulceration (see also Notes D, and E).

**N Category Considerations (see also Note M)**

The regional lymph nodes are the most common sites of metastasis. The widespread use of cutaneous lymphoscintigraphy, lymphatic mapping, and sentinel lymph node biopsies has greatly enhanced the ability to identify the presence of lymph node metastasis. By convention, the term regional lymph nodal metastasis refers to disease confined to one draining nodal basin or 2 contiguous draining nodal basins, as in patients with nodal disease in combinations of femoral/iliac, axillary/supraclavicular, cervical/supraclavicular, axillary/femoral, or bilateral axillary or femoral metastases. In some patients, lymphoscintigraphic imaging may define multiple regional lymph node basins and disease identified in any of such basins also constitutes regional disease. Metastasis to nondraining nodal basin(s) is considered M1 disease.

Sentinel Lymph Nodes

Sentinel lymph node identification and evaluation is often included in the surgical approach to cutaneous melanoma.1,2 A sentinel lymph node is defined as any lymph node receiving direct lymphatic drainage from a primary tumor site. There is often more than 1 sentinel node per draining basin and in some patients, lymphatic fluid may drain to more than one regional nodal basin.4 The clinical rationale for sentinel lymph node identification and separate evaluation assumes that metastatic involvement of a sentinel node is more likely than in regional nodes in the same nodal basin that do receive lymphatic drainage from the primary melanoma. Conversely, if sentinel nodes are negative, other regional nodes in the same basin would be less likely to contain metastasis.

**M Category Considerations**

The category “MX” has been eliminated from the AJCC/UICC TMN system.1 Unless there is clinical or pathologic evidence of distant metastasis the stage is classified as clinical M0 (ie, no distant metastasis). pM should *only* be reported when metastases have been documented by pathologic examination, that is, pM1 disease. pMX and pM0 should *not* be reported by the pathologist.

## TNM Descriptors

For identification of special cases of TNM or pTNM classifications, the “y,” “r,” and “a” prefixes are used. Although they do not affect the stage grouping, they indicate cases needing separate analysis.

Post-therapy or post neoadjuvant therapy classification (yTNM) documents the extent of the disease for patients whose first course of therapy includes systemic or radiation treatment prior to surgical resection or when systemic therapy or radiation is primary treatment with no surgical resection. The extent of disease is classified using the same T, N, and M definitions and identified as post-treatment with a “yc” or “yp” prefix (ycT, ycN, ycTNM; ypT, ypN, ypTNM).

Recurrence or retreatment classification (rTNM) may be used to define used because information gleaned from therapeutic procedures and from extent of disease defined clinically may be prognostic for patients with recurrent cancer after a disease-free interval. It is important to understand that the rTNM classification does not change the original clinical or pathologic staging of the case and that this classification schema is not yet widely used in melanoma.1,2

Autopsy classification (aTNM) is used to stage cases of cancer not recognized during life and only recognized postmortem.

## Lymphovascular Invasion

## Lymphovascular invasion (LVI) indicates whether microscopic lymphatic or vascular invasion is identified and includes lymphatic invasion, blood vessel invasion, or invasion of a vessel for which it cannot be determined whether it is a blood or lymphatic vessel lLymphovascular invasion). By AJCC/UICC convention, LVI does not affect the T category indicating local extent of tumor, ie, foci of Lymphovascular invasion should not be included in the measurement of tumor (Breslow) thickness.

## Pretreatment Serum Lactate Dehydrogenase and Serum Albumin

Data from numerous studies have indicated that an elevated serum level of lactate dehydrogenase (LDH) is an independent predictor of decreased survival in AJCC stage IV melanoma patients. In the AJCC eight edition, serum LDH is now recorded as “non-elevated” or “elevated” for each M category using the suffix, “(0)” or “(1)”, respectively. 1,2

References

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